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# **Cardiovascular** control mechanisms during anaesthesia and surgery

17

with special reference to muscle nerve sympathetic activity

### Johan Seligren

Göteborg 1993



### Cardiovascular control mechanisms during anaesthesia and surgery

with special reference to muscle nerve sympathetic activity

#### AKADEMISK AVHANDLING

som för avläggande av medicine doktorsexamen vid Göteborgs Universitet kommer att offentligen försvaras i föreläsningssal F3, Sahlgrenska Sjukhuset fredagen den 12 mars 1993, kl 09.00

av

Johan Sellgren leg. läkare

Avhandlingen baseras på följande delarbeten:

- I Anesthetic modulation of the cardiovascular response to microiaryngoscopy. A comparison of propofol and methohexital with special reference to leg blood flow, catecholamines and recovery. Sellgren J, Ejnell H, Pontén J, Sonander HG. Submitted for publication.
- $\mathbf I$  The effects of propofol, methohexitone and isoflurane on the baroreceptor reflex in the cat. Sellgren }, Biber B, Henriksson B-Å, Martner }, Pontén J. Acta Anaesthesiol Scand 1992; 36: 784-790.
- DI Characteristics of muscle nerve sympathetic activity during general anaesthesia in humans. Sellgren J, Pontén J, Wallin BG. Acta Anaesthesiol Scand 1992; 36: 336-345.
- IV Percutaneous recording of muscle nerve sympathetic activity during propofol, nitrous oxide and isoflurane anesthesia in humans. Sellgren J, Pontén J, Wallin BG. Anesthesiology 1990; 73: 20-27.
- V Sympathetic muscle nerve activity, peripheral blood flows and baroreceptor reflexes in humans during propofol anesthesia and surgery. Sellgren J, Ejnell H, Elam M, Pontén J, Wallin BG. Submitted for publication.

### Cardiovascular control mechanisms during anaesthesia and surgery

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Thesis defended March 12th, 1993.

#### Abstract.

Knowledge of effects of anaesthetics on cardiovascular control is based mainly on studies during undisturbed anaesthesia. However, in the clinical situation the cardiovascular characteristics of different anaesthetics are related to the balance between the dose of the anaesthetic and the intensity of surgical stimulation. Therefore, a general aim of the present studies was to investigate the effects of anaesthetics on cardiovascular control mechanisms both during undisturbed anaesthesia and during surgery. Since the trend in general anaesthesia is to use anaesthetics with short duration, the studies were focused on propofol in particular but also on methohexitone and isoflurane. Microlaryngoscopy was used as a surgical stress model since microlaryngoscopy evokes an intense and relatively stable afferent stimulation associated with a reproducible pressor response.

Methods: A main method in the human studies was microneurography of sympathetic vasoconstrictor nerve traffic to skeletal muscle blood vessels. It was thereby possible to differentiate between neurogenic effects and direct effects on the blood vessels from circulating factors including the anaesthetics themselves. Cardiac output (impedance cardiography) and regional blood flows (leg plethysmography, skin laser Doppler flowmetry, photoelectric pulse plethysmography) were recorded. Arterial catecholamine concentrations were measured. In addition, an experimental open loop baroreflex model (isolated carotid sinuses) was studied in the cat.

Results: Sympathetic activity to skeletal muscle (MSA) was depressed by propofol, methohexitone and isoflurane, whereas nitrous oxide was associated with an increase in MSA. The depression of MSA during undisturbed propofol infusion was to a large extent restored during microlaryngoscopy in spite of a more than three times increased propofol infusion rate. Vasodilation during propofol anaesthesia was caused by an inhibition of central sympathetic outflow and probably also by a direct vascular effect. In a comparative study during microlaryngoscopy, propofol was a better alternative than equianaesthetic doses of methohexitone, which in a low infusion dose was insufficient to control the microlaryngoscopyinduced pressor response and in a high infusion dose was associated with prolonged recovery. A large difference in leg blood flow was noted between the low and high-dose methohexitone groups whereas no difference was observed between the low and high-dose propofol groups. In the cat, the baroreflex sensitivity was better maintained during anaesthesia with propofol than with methohexitone or isoflurane. In humans, both cardiac and muscle sympathetic baroreflex sensitivities were depressed by propofol. The further depression of the cardiac baroreflex that was observed during surgery may have been due to a central vagal inhibition similar to that found in animals during defence area stimulation. The muscle nerve sympathetic baroreflex sensitivity was determined by a balance between an augmented central sympathetic outflow due to surgical stress and inhibition due to the anaesthetic.

Conclusions: Sympathetic activity to skeletal muscle is profoundly influenced by the choice of anaesthetic agent. A suppression of activity is more common than an increase. A decrease in MSA is counteracted by surgical stress. During propofol, methohexitone and isoflurane anaesthesia, the muscle nerve sympathetic baroreflex is qualitatively operative but the baroreflex sensitivity is depressed to a variable extent depending on the anaesthetic agent and depth of anaesthesia.

Key words: Anesthetics intravenous, propofol, methohexitai; anaesthetics volatile, isoflurane, nitrous oxide; surgery, larynx; sympathetic nervous system; arterial baroreceptors; sympathetic microneurography; plethysmography, leg blood flow; skin, blood flow; cats; humans.

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To

Marie-Louise, Anna, Erik and Fredrik

### Original papers

This thesis is based on the following papers, which will be referred to in the text by their Roman numerals:

**I** Anesthetic modulation of the cardiovascular response to microlaryngoscopy. A comparison of propofol and methohexital with special reference to leg blood flow, catecholamines and recovery. Sellgren J, Ejnell H, Pontén J, Sonander HG.

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- **III** Characteristics **of** muscle nerve sympathetic activity during general anaesthesia in humans. Sellgren J, Pontén J, Wallin BG. Acta Anaesthesiol Scand 1992; 36: 336-345.
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- **V** Sympathetic muscle nerve activity, peripheral blood flows and baroreceptor reflexes in humans during propofol anesthesia and surgery. Sellgren J, Ejnell H, Elam M, Pontén J, Wallin BG. Submitted for publication.

### Contents



## Abbreviations



### 1. Introduction

#### Physiological background

Cardiovascular homeostasis includes maintenance of an adequate blood pressure and optimal local tissue blood flows according to present metabolic demands. The cardiovascular control mechanisms are complex and involve many components<sup>1,2.</sup> The heart and the vascular smooth muscles serve as effector organs in the dynamic short-term circulatory regulation, which is the

subject of this thesis. The autonomic nervous system plays an important role in this regulation through cardiovascular reflexes mediated by neurohormonal pathways (fig. 1). Myogenic mechanisms 3,4 , local metabolites 5,6 , release of vasoactive substances such as histamine, bradykinin and prostaglandins<sup>7</sup>, and local nervous reflexes<sup>8</sup> also contribute to the control of vascular tone and regional blood flow.



The autonomic cardiovascular control centres are located in the medulla oblongata. Both the cardioinhibitory centres generating parasympathetic activity and the vasomotor centres responsible for sympathetic activity are affected by descending supramedullary differentiated activity. Efferent autonomic neural activity leaving the medulla is continuously modulated by afferent impulses from different sensory receptors. Arterial baroreceptors located in the carotid sinuses and aortic arch and cardiopulmonary baroreceptors in the caval vein, right atrium, right ventricle and lungs are mechanoreceptors responding to changes in blood pressure and ventilatory manoeuvres. A sudden hypotension decreases the afferent inhibitory effect of the baroreceptors on the vasomotor centre and causes an increase in efferent sympathetic nervous activity, which increases vascular resistance and restores blood pressure. The vasoconstrictor impulses are conducted in pre- and postganglionic sympathetic nerves to smooth muscles in the blood vessel walls. The nerve action-potentialinduced release of noradrenaline (NA) in the synaptic cleft induces an a-receptor-mediated vascular muscle contraction. The released NA is removed mainly by re-uptake or local metabolic breakdown but some spillover to the systemic circulation also occurs. The strength of the sympathetic neural outflow varies among different organs. The sympathetic response also includes a release of catecholamines, mainly adrenaline (A), from the adrenal medulla to the systemic circulation. In small concentrations, adrenaline induces vasodilation through  $\beta_2$ -receptor activation but with high concentrations the a-receptor-induced vasoconstriction dominates. The cardiac baroreflex response to a sudden hypotension is due to some extent to increased cardiac sympathetic activity but mainly to decreased parasympathetic activity<sup>9</sup>. The parasympathetic impulses are conducted by cholinergic fibres in the vagal nerves to the sinus and atrioventricular nodes for heart-rate modulation.

Arterial chemoreceptors are located in the carotid and aortic bodies and respond to changes in  $P_aCO_2$  and  $P_aO_2$ . Hypoxia or hypercapnoea will, in addition to the respiratory reflex response (increased minute ventilation), also have circulatory effects including increases in heart rate and sympathetic vasoconstrictor activity to skeletal muscle<sup>10,11</sup>. These increases are more prominent during hypercapnoea than during hypoxia. Simultaneous hypercapnoea and hypoxia has a synergistic effect on sympathetic activity.

Somatic afferent stimuli, like pain and cold, also evoke cardiovascular responses mediated by the autonomic nervous system<sup>12</sup>. Both medullary and supramedullary pathways are involved in these somatosympathetic reflexes. The efferent responses include increased heart rate and increased sympathetic nervous activity. However, in contrast to the baroreflex response, the somatosympathetically induced vasoconstriction is more prominent in the visceral and renal vascular beds than in the skeletal muscle vasculature<sup>13</sup>. This response seems to be functionally related to the defence reaction, which is evoked by rage or fear and prepares the organism for fight or flight. Experimentally, this reaction can be evoked by electrical stimulation of the hypothalamic defence area<sup>13-18</sup>. The increased heart rate, blood pressure and skeletal muscle blood flow are appropriate for instant physical activity. Normally, the increase of arterial blood pressure is modulated by a decrease of heart rate. However, during the somatosympathetic reflex response and defence reaction, the cardiac baroreflex is inhibited to improve cardiac performance<sup>13-15</sup>.

The hypothalamic and medullary cardiovascular control centres are influenced continuously by several afferent stimuli. The efferent autonomic activity is therefore an integration of different reflex responses, which together with local factors regulate the blood flow distribution. There are large differences in regional blood flows (fig. 2)<sup>5</sup>. Although the total blood flows to the kidneys and skeletal muscles are similar at rest, each with about 20 % of cardiac output, their respective regional blood flows are quite different. The kidneys have a constant low vascular resistance and a high blood flow (300-400 ml·min<sup>-1.</sup>  $100 \text{ g}^{-1}$ ) in order to support the renal clearance function while the skeletal muscles, which represent 45 % of total body mass, at rest have a low blood flow  $(2-5 \text{ ml-min-1.100 g<sup>-1</sup>)$  due to relative vasoconstriction. However, the renal blood flow is already close to its maximal flow, whereas the blood flow to skeletal muscle can increase more than 20-fold

during physical exercise. Cholinergic vasodilatory nerve fibres, known to exist in cats, have not been found in humans and vasodilation in human skeletal muscle is therefore achieved by local metabolic factors and by inhibition of sympathetic vasoconstrictor activity. Skeletal muscle vessels are also more responsive to sympathetic vasoconstrictor impulses than renal vessels, although the sympathetically mediated increase in vascular resistance is most prominent in cutaneous vessels 2 . Due to the large skeletal muscle mass, even small changes in vascular resistance governed by efferent sympathetic activity are of importance for modulating the systemic vascular resistance and arterial blood pressure. Sympathetic control of the splanchnic circulation contributes to this response but in this vascular bed also has a major vasoconstrictor effect on the capacitance vessels 2 . Central blood volume is thereby restored and the preload and cardiac output are increased.





#### Influence of anaesthesia and surgery

Anaesthetics interact with the cardiovascular regulation in several ways<sup>19,20</sup>. Interaction with the autonomic nervous system is common and can include both effects on supramedullary centres responsible for the tonic sympathetic outflow and effects on different cardiovascular reflexes. In addition, most anaesthetics have direct effects on heart rate, myocardial performance and vascular contractility. Since different anaesthetics affect the cardiovascular regulatory system at different sites and therefore have different circulatory characteristics, anaesthetic techniques may be individualized with regard to the type of operation and physical condition of the patient.

During anaesthesia, a general aim concerning circulation is to depress surgically induced somatosympathetic reflexes. These reflexes, inducing increases in blood pressure and heart rate, are similar to the fight-and-flight reaction<sup>12</sup> but inappropriate in the operating situation and may be dangerous in patients with coronary artery insufficiency or aneurysms in the aorta or the intracranial vessels. Although all anaesthetics, to varying extent, depress this somatic pressor response, the effect is not always related to the depth of anaesthesia. An effective reflex depression can also be achieved without general anaesthesia by, for example, regional blockade of the afferent somatosensoric nerve activity. Barbiturates have been shown to inhibit somatosympathetic reflexes at supramedullary level<sup>21</sup>, whereas inhalation anaesthetics also have been shown to depress sympathetic ganglionic transmission<sup>22</sup>. The opiates act mainly through the opiate receptor endorphin system, which may explain their minor cortical and cardiovascular effects<sup>23,24</sup>.

The effects of different anaesthetics on the baroreceptor reflexes vary. Whether it is desirable or not to depress these reflexes depends on the state of the patient and the surgical situation. For example, in a hypovolaemic patient, anaesthesia with a baroreflex-depressing agent such as thiopentone can induce severe hypotension. Similarly, hypotension can be caused by a high spinal or epidural anaesthesia which blocks efferent sympathetic vasoconstrictor reflex discharge. However, depression of the baroreflex can also be used as a tool to balance the somatosympathetic reflex induced by surgery. In several studies the baroreflex has been depressed by barbiturates<sup>25-27</sup>, halothane<sup>28-31</sup>, enflurane<sup>32,33</sup> and isoflurane<sup>22,32,34,35</sup>, whereas nitrous oxide, and the previously used inhalation agents ether<sup>36</sup> and cyclopropane<sup>37</sup> have been associated with increased baroreflex responses. Opiates have been shown to have minor or no effects on the baroreflex response<sup>26</sup>

The trend in general anaesthesia is to use anaesthetics with short duration. It is thereby easier to obtain cardiovascular stability by adjustments of the dose when needed. The rapid recovery with these agents is also an advantage both for patient comfort and economically. In our studies, we have focused our interest on three short-acting anaesthetics: propofol, methohexitone and isoflurane. Propofol, which is a phenol solved in fat emulsion, was introduced in Sweden in 1987. Propofol is characterised by a very short duration, mainly due to redistribution but also to a high clearance rate, and is thereby associated with a rapid recovery<sup>38-40</sup>. Propofol has potent circulatory effects, with more pronounced hypotension during induction of anaesthesia than other intravenous induction agents<sup>41-44</sup>. No studies concerning the effects of propofol on the baroreceptor reflex were available when our studies started.

Methohexitone is the most short-acting barbiturate and therefore, in contrast to thiopentone, it has been used for both induction and maintenance of anaesthesia<sup>45</sup> . The main disadvantage compared with thiopentone has been excitatory effects such as movements, hiccups and laryngospasm. The circulatory effects of methohexitone are accompanied by depression of the baroreceptor reflex $25-27$ . The inhalation agent isoflurane is, due to its comparatively lower blood gas solubility and low degree of biotransformation, associated with more rapid drug uptake and emergence from anaesthesia than halothane and enflurane<sup>46-48</sup>. Typical circulatory effects of isoflurane are a decrease in systemic vascular resistance and an increase in heart rate but also a depression of the baroreceptor reflex<sup>22,32,34,35</sup>.

Knowledge of the effects of anaesthetics on cardiovascular control is based mainly on studies during undisturbed anaesthesia. However, in the clinical situation the cardiovascular characteristics of different anaesthetics are related to the balance between the dose and the intensity of surgical stress. A general aim in our studies was therefore to investigate the effects of several anaesthetics (propofol in particular but also methohexitone and isoflurane) on cardiovascular control mechanisms both during undisturbed anaesthesia and during surgery. Microlaryngoscopy was used as a surgical stress model since microlaryngoscopy evokes an intense and relatively stable somatic afferent stimulation associated with a reproducible pressor response. The main method in these studies was microneurography of sympathetic nervous activity to skeletal muscle vessels, a method enabling us to monitor the efferent vasoconstrictor activity. It was thereby possible to evaluate neurogenic effects of anaesthetics on the blood flow to skeletal muscle. The dynamic properties of the muscle sympathetic part of the baroreflex were also studied in relation to anaesthesia and surgery. Although the microneurographic technique of recording sympathetic activity was presented already in 1968, the present studies represent the first recordings of MSA during general anaesthesia<sup>49</sup>.



### 2. Aims of the study

- To study the differentiation in systemic and peripheral (leg) blood flows during surgery and anaesthesia with either propofol or methohexitone.
- To evaluate the relative effects of propofol, methohexitone and isoflurane on the baroreceptor reflex in an experimental open loop model.
- To describe the general characteristics of muscle nerve sympathetic activity during induction and maintenance of anaesthesia with different anaesthetics and the effects of different reflex stimuli and surgical stress.
- To study the effects on muscle nerve sympathetic activity of induction of  $\bullet$ anaesthesia with propofol and maintenance of anaesthesia with nitrous oxide and/or isoflurane.
- To study the effects on muscle nerve sympathetic activity and peripheral blood flows of undisturbed steady-state propofol anaesthesia and of surgical stress.
- To study the cardiac and muscle sympathetic baroreflex sensitivity during undisturbed steady-state propofol anaesthesia and during surgical stress.



### 3. **Methodological** considerations

#### Experimental carotid baroreflex open loop model

The baroreceptor reflexes are normally closed feed-back loops; that is, the baroreflex-induced change in blood pressure affects in itself the baroreceptor afferent activity. This regulatory feedback system is essential for haemodynamic stability. However, when one is studying baroreflex responses the feedback may be a disturbing factor. Therefore, a carotid baroreflex open loop model was studied in chloraloseanaesthetized cats (II). This model is well established in our laboratory<sup>50-52</sup>. Chloralose is a suitable basal anaesthetic in experimental cardiovascular studies since it preserves the baroreceptor reflexes<sup>26,53</sup>. If anything, the baroreflex sensitivity has been shown to be slightly increased by chloralose<sup>54</sup>.

Both the carotid sinuses were partly isolated by ligation of the external and sometimes the internal carotid artery as well as all other arterial branches that could be ligated without endangering the integrity of the sinus nerve by the dissecting procedure. Catheters were inserted into the common carotid arteries bilaterally, allowing the carotid sinuses to be perfused with blood from the femoral arteries at any desired level of pulsatile pressure by means of a roller pump (fig. 3). In order to interrupt all closed baroreflex loops, the cardiopulmonary and aortic baroreceptors were denervated by bilateral sectioning of the vagal nerves. The remaining cardiac innervation was thus of sympathetic origin. This is of impor-

tance for interpretation of our data since vagal influence normally overrides sympathetic for the modulation of heart rate<sup>9</sup>. In order to allow adjustments of the carotid sinus perfusion pressure, when perfused by the pump, without changing the pump frequency (set at 175 rpm/min), an adjustable arteriovenous shunt was inserted between the carotid sinuses inflow tube and an external jugular vein. By means of a by-pass circuit parallel to the pump, the carotid sinuses could also be exposed to prevailing femoral arterial pressure. The carotid sinus perfusion pressure was recorded from a side branch of the tubing system. Systemic arterial pressure was recorded through a catheter inserted in a brachial artery. The systemic MAP was lower when the carotid sinuses were perfused by artificial pumping at



*Figure. 3. The model for carotid sinus perfusion. The carotid sinuses were perfused with blood from the femoral arteries either at prevailing systemic arterial pressure (by-pass open, pump off, jugular vein shunt closed) or at any desired pump- and jugular-shunt-controlled pressure level (by-pass closed, pump on and jugular-shunt variably open).* 

a perfusion pressure equivalent to prevailing baseline MAP than when perfused by the cardiac-generated pressure (via the by-pass). This difference may to some extent depend on the pressure gradient (6 ±2 SEM mm Hg) in the femoral-carotid shunt tubes. However, the altered pulse pressure amplitude and pulse wave-form during pump-generated carotid perfusion as compared to when the pressure was generated by the heart is probably more important. Since these conditions were essentially constant during the experiment, it is likely that such factors did not influence the results.

Baseline data were obtained by exposing the carotid sinuses to the prevailing femoral arterial pressure through the bypass. Baroreceptor function was assessed by investigating how induced changes in the artificially pump-controlled carotid sinus perfusion pressure influenced systemic mean arterial pressure (MAP) and heart rate (HR). Carotid sinus pressure was thereby varied in steps of 25 mm Hg over a pressure range of 50- 200 mm Hg and for each level of sinus pressure corresponding data for AP and HR were recorded after steady-state had been attained (5-20 sec). In this way the open loop gain, i.e. the inverse change in systemic arterial pressure or heart rate per unit change in carotid sinus pressure, could be calculated for each step of 25 mm Hg carotid sinus pressure change throughout the investigated range. Data obtained during basal chloralose anaesthesia alone, before administration of each anaesthetic, were used as control values.

In order to study the dynamic range of the baroreceptor reflex response, systemic mean arterial pressure and heart rate at carotid sinus pressures of 50-150 mm Hg were analysed in detail. The influence of the different anaesthetic agents on the baroreceptor reflex was evaluated by assessing the effects on the following characteristics of baroreceptor reflex function:

- 1. The overall baroreflex capacity to regulate systemic mean arterial pressure and heart rate (when carotid sinus pressure was varied from 50 to 150 mm Hg).
- 2. The baroreflex set point evaluated by identifying the carotid sinus pressure interval associated with the maximal open loop gain in systemic mean arterial pressure and heart rate response.
- 3. The baroreflex sensitivity defined as the efficiency of the baroreceptor reflex to influence systemic mean arterial pressure and heart rate as derived from the carotid sinus pressure interval which was associated with the maximal open loop gain. Sensitivity was expressed as changes in systemic MAP or HR induced by 1 mm Hg change in carotid sinus pressure.

### Mi croneurography

Direct microelectrode recordings of sympathetic discharges in human extremity nerves were first presented by Hagbarth and Vallbo in 1968<sup>55</sup>. This efferent autonomic activity is conducted in unmyelinated C-fibres located intrafascicularly in somatic nerves. Sympathetic nervous activity has been recorded in nerves conducting vasoconstrictor impulses to skeletal muscle (muscle nerve sympathetic activity; MSA)<sup>56,57</sup> and in nerves conducting both vasoconstrictor and sudomotor impulses to skin (skin nerve sympathetic activity; SSA)<sup>58,59</sup>. Most recordings have been made from the peroneal, tibial and median nerves.

#### *Recording technique*

The recording microelectrode is made of lacquer-insulated tungsten wire with a diameter of  $200 \mu$ , which has been electrolytically pointed to a diameter of a few  $\mu$ . The present recordings have been made in the peroneal nerve at the fibular head. After locating the nerve with weak cutaneous electrical stimuli, the electrode was inserted manually through intact skin into the underlying nerve. A reference electrode was placed subcutaneously about 2 cm from the recording electrode, which was advanced during simultaneous weak electrical stimulation (1-3 V, 1 Hz). The nerve contains fascicles innervating skeletal muscle and skin which can be differentiated on the basis of the effects evoked by stimulation through the needle and the afferent responses induced by certain peripheral stimuli. After a muscle nerve fascicle had been located, the postganglionic sympathetic nerve fibres were found by small adjustments of the recording electrode within the fascicle (fig. 4).

The criteria for acceptable recordings of MSA, upon which our studies were focused, were as follows:

- weak electrical stimulation through the electrode elicited involuntary muscle contraction of appropriate muscles but not paraesthesiae,
- tapping or stretching muscles or tendons innervated by the impaled fascicle evoked afferent mechanoreceptor discharges but similar activity was not elicited by stroking the skin,
- spontaneous pulse-synchronous bursts of sympathetic impulses occurred intermittently and increased during expiratory apnoeas and during the hypotension induced by the Valsalva manoeuvre (phase II and III).



*Figure. 4. A schematic view of the recording electrode inserted intrafascicularly in the peroneal nerve. The unmyelinated sympathetic C-fibres are depicted as light inclusions in the Schwann cell.* 



*Figure.* 5. *Muscle nerve sympathetic activity (MSA) from a peroneal recording in the awake state.* MSA *bursts are related to decreases in the arterial blood pressure. Note the large M SA burst evoked by the prolonged diastole after an extrasystolic heart beat (at arrow).* 

Evidence that the recorded nerve activity is of sympathetic origin comprises the following observations <sup>60</sup>:

- injection of local anaesthetic around the nerve proximal but not distal to the recording site eliminates the activity,
- the conduction velocity of the impulses is approximately 1 m/s, which is appropriate for unmyelinated C-fibres,
- intravenous infusion of a ganglionblocking drug such as trimetaphan reversibly eliminates the activity,
- changes in the intensity of the nerve activity are followed within a few seconds by sympathetically mediated responses, such as blood pressure, leg or forearm blood flow (mainly MSA), skin blood flow (SSA) and skin electrical resistance (SSA).

#### *Microneurographic findings*

The old view of a generalized sympathetic tone has been rejected by, for instance, studies of MSA and SSA which have shown pronounced differentiation<sup>56,59,61,62</sup>. Short lasting emotional excitement and mental stress evoked by arithmetic problems have been associated with strong SSA

responses without changes in MSA<sup>59</sup>. SSA is important for temperature regulation and has been shown to increase both at ambient temperatures below (vasoconstrictor impulses) and above (sudomotor impulses) thermoneutral conditions 63 . Thiopentone and halothane anaesthesia have been associated with dose-dependent reductions of SSA<sup>64</sup>.

MSA, in contrast to SSA, is an efferent part of the baroreceptor reflex and is therefore modulated by changes in blood pressure<sup>65</sup> . A decrease in arterial blood pressure evokes MSA bursts (fig. 5) aiming to restore blood pressure by an increase in skeletal muscle vascular resistance. The sensitivity and speed of this response is illustrated by the large burst evoked by the prolonged diastole after an extrasystolic heart beat (fig. 5). The response delay of approximately 1.3 s between the ventricular depolarisation in the ECG and the MSA burst inhibition in the peroneal nerve represents to a large extent the C-fibre conduction velocity<sup>57,66</sup>.

The characteristic pulse synchrony in MSA is due to baroreflex modulation: each systolic blood pressure peak causes a short-lasting inhibition of sympathetic outflow. The importance of the inhibitory afferent nerve activity from the arterial baroreceptors for the pulse synchrony has been illustrated by the effects of bilateral local anaesthesia of the glossopharyngeal and vagal nerves in the neck<sup>67</sup>. This procedure evoked a pronounced increase in MSA (and arterial blood pressure) and the pulse synchrony was replaced by a fast and irregular burst rhythm. Changes in MSA and arterial blood pressure have been shown to be positively correlated to diastolic but not to systolic blood pressure<sup>65</sup>. At a given blood pressure, the sympathetic discharges are stronger when pressure is decreasing than when it is increasing<sup>65</sup>, i.e. the direction of the pressure change is important for the MSA response. The MSA responses due to dynamic arterial baroreflex function buffer short-term blood pressure changes, whereas preload changes affecting cardiopulmonary low-pressure baroreceptors exert more tonic reflex effects on MSA. The importance of these low pressure baroreceptors has been shown when central venous pressure (CVP) but not arterial pressure has been decreased by applying subatmospheric pressure around the lower body<sup>68</sup>. Arterial pressure and CVP often change in the same direction, for example during a Valsalva manoeuvre, and the MSA response is therefore due to a combination of arterial and low-pressure baroreflex effects. When changing posture from the supine to the upright position, the arterial baroreflex contributes to the initial MSA increase whereas the persisting higher level of MSA is due to unloading of cardiopulmonary baroreceptors.

During resting conditions, the MSA burst frequency shows large interindividual differences (from less than 10 to more than 90 bursts/100 heart beats)<sup>69</sup>. However, in the same individual, simultaneous recordings from different

nerves and repeated recordings also after several months show similar burst frequency 69 . Interindividual comparisons of burst frequency have shown a weak positive correlation to age but no correlation to the blood pressure level in normotensive subjects.

#### *Nerve signal processing and data analysis*

The nerve signal was amplified with a gain of 50000 and the signal-to-noise ratio was improved by using a 700-2000 Hz bandpass filter and an amplitude discriminator (fig. 6). An RC-integrating network with a time constant of 0.1 sec was used to obtain a mean voltage display of the multi-unit nerve activity (fig. 6). MSA can be presented as burst frequency (per minute or per 100 heart beats)( $III, IV, V$ ) or as "total MSA", i.e. the product of burst frequency/minute and either the burst mean amplitude (III,IV) or area (V). The MSA burst detection and quantification can be done manually with computer support (III,IV) or by computer only after setting detection criteria  $(V)$ . The computer program<sup>70</sup> emulates the manual analysis by detecting each MSA burst in the mean voltage neurogram and then calculates its area. A sympathetic burst was identified on the basis of a monotonie rise and fall with a local maximum occurring in the MSA signal within 1000-1800 ms from a preceding R-wave in the ECG. The parameters defining a burst could be changed manually and if the automatically detected bursts did not agree with the visually defined ones, parameters were changed and the automatic burst detection was restarted. The area of a burst was calculated from a relative baseline (mean value of background activity between bursts) set by the computer. All recorded circulatory parameters in the respective study protocol were stored on tape and subsequently analysed by computer. For



*Figure. 6. Original, discriminated and integrated neurogram of muscle nerve sympathetic activity (MSA) and arterial blood pressure. The recording is from the same sequence as in figure* 5 *but here shown at a faster speed.* 

further analysis, all data in relation to each corresponding heart beat were exported to Excel spreadsheets (Microsoft, USA).

#### *Recording equipment and success rate*

All microneurographic recordings were performed in the operating room. Besides a nerve amplifier, tape recorder and ink-jet recorder for measuring and storing MSA, the recording set-up also included equipment for continuous measurements of ECG, invasive arterial blood pressure, laser Doppler skin blood flow, finger pulse amplitude, respiratory movements, end-tidal CO<sub>2</sub> concentration,  $F_iO_2$ , end-tidal isoflurane concentration (paper IV) and intermittent plethysmographic measurements of leg blood flow (fig. 7). The patients rested as comfortably as possible on the operating table and had extra blankets if needed to prevent cooling and discomfort. The ambient temperature in the room was kept constant during the experiment. Due to the complexity of the microneurographic technique when recordings are made during general anaesthesia, the number of patients included in the studies was lower than the total number of patients investigated (table 1).

#### *CUSUM-technique*

To clarify the time relations between the start of changes in MSA and MAP after





*Figure.* **7.** *The recording setup in the operation room during the microneurographic studies.* 



injection of a drug, a cumulative sumtechnique  $(CUSUM)^{71}$  was used (III). Total MSA and MAP were averaged for every 5 heart beats. During a 25-50 sec control period before the manoeuvre (for example the induction of anaesthesia) averaged values of MSA and MAP were set to 100%. The differences between the values for each period of 5 heart beats and the respective average control value were then cumulated during the control period and the initial period after the injection. Taking a reference level equal to the mean of a control period gives a CUSUM of zero slope. With the cumulation, the onset of a change in MSA or MAP was magnified and thus easy to detect.

#### *Baroreflex tests*

In humans, cardiac baroreflex sensitivity is usually determined by plotting arterial blood pressure against the corresponding RR-interval after intravenous injection of a pressor drug such as phenylephrine or angiotensin. The baroreflex sensitivity is represented by the slope when the x-y-plot has been subjected to linear regression analysis. This "slope method" was originally described by Smyth et al. 1969<sup>72</sup>. Later, depressor drugs such as

nitroprusside (SNP) or nitroglycerine have also been used in a corresponding way 73 . During anaesthesia MSA is low. A pressor test therefore reduces MSA below the detection limit. Consequently, in our anaesthesia-related studies, we used only depressor tests, which activate MSA. The depressor test period used by us for analysis was delimited by the start of the SNP-induced decrease in MAP and the point within 60 s when the lowest MAP was achieved (fig. 8). Since not all heart beats are associated with MSA bursts, it is not possible to plot the individual MSA burst area against the corresponding blood pressure value for every heart beat and therefore the x-y-plot method had to be slightly modified. The approach used was to perform several depressor tests under the same experimental condition and to utilize all heart beats from all tests to construct the diagram. The heart beats were sorted according to blood pressure values. The blood pressure range was divided in ten equal intervals each comprising an equal number of heart beats and the mean of all blood pressure values in each interval was plotted against corresponding values of MSAA and RR-interval, respectively. DAP was used for plotting against MSAA<sup>65</sup>

whereas MAP was used for plotting against RR-intervals. Regression lines with a correlation coefficient below 0.5 were excluded. This limit is slightly lower than a correlation coefficient of 0.63, which corresponds to p<0.Q5, originally used by Smyth. In a recent paper, Sleight has discussed whether or not slopes with lower correlation coefficients also ought to contribute to the average slope value. Sleight proposes



*Figure. 8. (A) Original recording. Muscle sympathetic neurogram and haemodynamic recordings from a typical depressor test in the awake state in one patient (intravenous injection of sodium nitroprusside, SNP, 2 figkg' <sup>1</sup>marked with an arrow). The depressor test period as delimited by start of decrease in MAP and the lowest achieved MAP is marked with lines. Note the single large MSA bursts caused by a sudden decrease in DAP after two spontaneous extrasystolic heart beats. (B) Baroreflex slope. X-y-plot of MSAA vs. DAP and linear regression analysis of pooled data from four depressor tests before anaesthesia in the same patient. MSAA is expressed as percent of basal MSAA value during the awake control period. The MSAA set point, which corresponds to the preSNP reference blood pressure is marked with a dotted line.* 

weighting of the slopes by their correlation coefficients <sup>74</sup> . This procedure is, however, not useful for a group of patients if statistical comparisons are to be made, since standard deviations cannot be determined. If all slopes (irrespective of correlation coefficients) had been included in study V, the results would not have been different from those presented in the paper.

In 2 of the 9 patients in study V some depressor tests during anaesthesia and surgery increased the baseline of the integrated neurogram. Reintegration of the original neurograms using a shorter time constant (0.05 s) eliminated only part of the baseline elevation. This indicates that the sympathetic nerve activity had become continuous and systolic baroreflex inhibitions were incomplete. Since the continuous nerve activity was not included when calculating MSAA, sympathetic baroreflex sensitivity was somewhat underestimated in these patients, especially during surgery, when the elevations of baseline were most marked.

Other manoeuvres used in study III were ventilator-induced sighs, valsalva-like manoeuvres and transitory increases of PEEP. These ventilatory manoeuvres induce combined responses from both arterial and cardiopulmonary baroreceptors as well as mechanical stretch receptors in the lungs.

#### Strain gauge plethysmography

Venous occlusion plethysmography for measurements of arterial blood flow in a limb was first presented by Brodie and Russell in  $1905^{75}$  and during the first decades of the 20th century the technique was further developed. The recorded limb was sealed in a solid jacket and the sealed space was filled with air or water and connected to a volume recorder. Some disadvantages of the original technique were reduced by Whitney when he, in 1949, introduced the mercury-in-rubber strain gauge plethysmograph. This improved method was fully described by Whitney in 1953<sup>76</sup> . The silicon rubber tube (bore diameter 0.5 mm and wall thickness 0.8 mm) is placed around the thickest part of the calf or forearm and slightly extended. A Wheatstone bridge circuit (50 mA) is connected to the tube ends which are closed with metal plugs and thus in electrical contact with the mercury. A change in electrical resistance between the ends of the tube is directly proportional to the change in length of the tube. Calibration of the strain gauge is performed by a 3 mm extension. To reduce artefacts, the forearm or calf must be positioned slightly above heart level and not in contact with any surface. In order to estimate the arterial blood flow in the limb, the venous return is temporarily stopped by an occlusion cuff (40 mm Hg) placed proximal to the strain gauge. During the first 5-10 s the increase in





*Figure. 9. Strain gauge plethysmography. The left part of the figure shows calibration of electrical resistance corresponding to* 3 *mm extension of the mercury-filled silicon rubber tube. The right part of the figure shows the effect on electrical resistance of sudden venous occlusion.* 

electrical resistance represents the arterial blood flow (fig. 9). During longer venous occlusion the arterial blood flow decreases due to gradually increased venous blood pressure. Plethysmography of the forearm and calf mainly represents skeletal muscle blood flows. Influences of skin blood flow from the hand or the foot can be eliminated by an arterial occlusion cuff at the wrist or ankle. Distal occlusion of the hand is most important due to the relatively small muscle volume in the forearm $77$ .

In the formula used for calculation of blood flow "a" represents the calibration change in electrical resistance, corresponding to 3 mm extension. The slope expressed by "d" is related to extrapolation of the first linear part of the curve (5-10 s) to a period of 30 s. A factor 2 is added to the numerator in order to obtain flow/minute and another factor 2 in order to transform the change in circumference to a change in volume. The factor 100 is added to obtain flow/100 ml tissue. Leg (calf) blood flows were in our studies based on the average of 2-5 measurements. Reference values of leg blood flow range from 1.4 to 3.6 ml $\cdot$ min<sup>-1</sup> $\cdot$ 100 ml tissue<sup>-178</sup>. Although, there is a high correlation between the different plethysmography techniques, the strain gauge method will give a 9% underestimation compared with water plethysmography 79 . We have therefore only expressed our data in terms of relative changes related to measurements during a control period in each patient.

#### Impedance cardiography

Impedance cardiography for measurements of cardiac stroke volume was developed by Kubicek et al during the early 1960s<sup>80</sup>. This noninvasive method was originally developed for space research but later it has also been used in clinical research and for routine



*Figure 10. Impedance cardiography. The maximum value of the first derivative of thoracic impedance (dZ/dt<sub>max</sub>)* and left ventricular *ejection time (LVET) are used in Kubiceks formula for calculation of stroke volume (SV).* 

haemodynamic monitoring. Two circular electrodes (metal tape, 3M) are placed around the neck and two around the lower chest. The two outer electrodes are connected to a constant current source providing a 100 kHz sinusoidal current. The resulting voltage is monitored from the two inner electrodes by a high impedance amplifier and a detection circuit. The method is based on the assumption that the chest is a cylinder. When a rapid sinusoidal current is transmitted across this cylindrical distensible fluid, an increase of fluid in the cylinder causes a decrease in impedance which is directly proportional to the increase in cylinder volume. The change in impedance and its first derivative (dZ/dt), ECG and phonocardiogram (in order to detect the aorta valve closure) in relation to the cardiac cycle are shown in figure 10.

In Kubicek's formula for calculation of stroke volume the electrical resistivity of

blood (r) is positively correlated to the haematocrit value. Since no changes in haematocrit values were expected during the recording period in our patients, the resistivity value was set to 127 (ohm-cm), which refers to a haematocrit value of 40% at 37.5 $^{\circ}$  C<sup>81,82</sup>. L is the mean distance between the two inner electrodes (cm).  $Z<sub>0</sub>$  is the basic impedance across the chest. LVET is the left ventricular ejection time (s) measured from the shift in the derived impedance curve indicating the start of the ejection to the start of the second heart sound. dZ/dt (ohm $-cm·s<sup>-1</sup>$ ) is the difference between the basic impedance level and the maximum value in the derived impedance curve.

We used an impedance cardiograph model 400 (Instrumentation for Medicine Inc, Greenwich, Ct, USA). The impedance waves, ECG and phonocardiogram were presented on a Mingograph (Siemens-Elema) ink chart recorder. The ejection time (LVET) and dZ/dt were manually derived from the recorded tracings. The recordings were made during relaxed apnoea and each stroke volume was calculated from mean values of 6 consecutive heart beats. Cardiac output was calculated from the product of stroke volume and heart rate. Although impedance cardiography has shown a high agreement with cardiac output measured with dye dilution<sup>83</sup> and thermodilution<sup>84</sup> techniques, the results in our study (I) are presented as relative changes from an initial control period before start of anaesthesia.

#### Laser Doppler flowmetry

Laser Doppler flowmetry measures the flux of erythrocytes, that is the product of the number of erythrocytes and their velocity. The use of this method for blood flow measurements was first

presented in 1972 by Riva 85 , The first studies concerned blood flow in retinal vessels. In 1975 measurements of skin blood flow were presented and in 1977 the first equipment for clinical use was available<sup>86</sup>. Further development and evaluation of the technique was done by Tenland and Nilsson $87-89$ . The technique uses laser light with a wavelength of 632.8 nm in vacuum (red light) to illuminate the examined tissue. Light beams scattered in moving erythrocytes undergo a frequency shift according to the Doppler effect. These frequency fluctuations in reflected light are measured by a photodetection unit. The output signal has been shown to be directly proportional to the flux of erythrocytes in an experimental model. A decrease in oxygen tension from 15 to 5 kPa has been shown to decrease the flowmetry output by 5%. The measuring depth in skin is assumed to be about 1 mm. Since large local variations in skin blood flow have been observed, it is important that repeated recordings are made from the same site.

In our studies, we have used PeriFlux Id (IV) and 2B (V), Perimed AB, Stockholm, Sweden. The model 2B has improved linearity to overcome the disadvantage of a slight underestimation at high flow rates. The laser Doppler probe was placed on the plantar side of the right big toe and not moved during the experiment. Before the initial control period, the zero level was checked and recorded on tape for subsequent quantification. In some experiments the gain was changed when the output signal exceeded the maximal range. Since these values are relative and not absolute measurements of the blood flow, data were presented as percentage changes of the awake control period value  $(= 100\%)$ .

Laser Doppler flowmetry has been compared with heat and isotope washout techniques, dynamic capillary

microscopy, occlusion and photopulse plethysmography with good qualitative correlation<sup>87,90,91</sup>.

#### Photoelectric pulse plethysmography

The use of photoelectric plethysmography for blood volume measurements started in the 1930s. Pioneer methodological work was done by Hertzman<sup>92</sup>. The principle of the technique is illumination of, for example, a skin area and photodetection of the light modulated by blood volume changes. The light source and the photodetector can be arranged on either side on, for example, a finger (transillumination) or positioned side by side on virtually any skin area. With both arrangements, the light undergoes scattering, absorption, reflection and refraction as it passes through the skin. Nowadays, both the light source and the photodetector are made with semiconductor technology, as the modified van Gough type ILP/7A used by us (III,IV,V), and can therefore be placed side by side in a small-sized probe head<sup>93</sup>. The light emitting diodes (LED) generate light in the IR-wave range (800-900 nm) without heating the skin. Use of IR-light has decreased errors due to influence of ambient light and temperature changes. The potential error in relation to haemoglobin oxygenation which occurs at lower light waves (below 800 nm) is negligible in the IR-wave range used.

The pulse amplitude is used as an index of blood flow. Although the pulsations mainly represent changes in blood volume, it has also has been proposed that orientation and packing of erythrocytes contribute to the pulse amplitude. This was shown by Challoner and recently also by Lindberg in experiments with peristaltic blood flow through a system of closed, glass tubes<sup>93,94</sup>. Although the glass tubes did not allow volume changes, a photoelectric pulse Plethysmograph recorded a pulsative flow in the tubes. The photoelectric method has been used for blood flow measurements on the surfaces of the kidneys, liver, brain, mesenterium and nasal septum, but the most common use is for skin blood flow measurements. Comparative studies between photoelectric pulse plethysmography and occlusion plethysmography<sup>95</sup>, impedance plethysmography<sup>96</sup>, piezoelectric plethysmography<sup>97</sup> and laser Doppler  $f$ lowmetry $98$  have shown good qualitative correlations.

Unfortunately, the depth of measurement in the skin is unknown. Blood flow in all vessels contributes to the output signal (i.e. both flow through arteriovenous anastomoses and through the capillary network). Since it is difficult to obtain reliable absolute flow values with the photoelectric pulse plethysmographie method, all values in our studies (III,IV,V) are presented as relative values related to the awake control period.

#### Catecholamine, propofol and methohexitone concentration measurements

For catecholamine analysis (I) 5 ml of arterial blood was sampled in iced tubes containing 1.4 mg heparin and 50  $\mu$ l of a 0.4 M glutathione solution. The samples were stored in ice-water and immediately after terminating the anaesthesia they were centrifuged at 4°C and 3000 rpm for 10 minutes. The plasma was removed and stored at -70°C until analysed. The levels of adrenaline, noradrenaline and dopamine were determined by electrochemical detection after high-performance liquid chromatography (HPLC)<sup>99,100</sup>. The minimum detection level was  $0.03$  ng $\cdot$ ml<sup>-1</sup>.

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Blood samples for measurements of propofol concentrations (I, II) were stored in heparin tubes at 4°C before analysis. Samples for analysis of methohexitone (I, II) were centrifuged and plasma were stored at -70°C. Both concentrations of propofol in blood 101 and methohexitone in plasma<sup>102</sup> were analysed by HPLC-technique.

#### **Statistics**

In general, parametric statistical tests were used. Analysis of variance (ANOVA) was used in all studies to detect overall statistical significance before further multiple comparisons were made (I-V). Statistical comparisons between groups were performed with one- or two-factor ANOVA with the Tukey Compromise post-hoc test (I,II,V). Within group comparisons were made with either one-factor ANOVA with Fisher's protected least significant difference (PLSD) comparative test (III,IV) or multiple paired Student's T-tests with Bonferroni correction for multiple comparisons (I,V). In some cases nonparametric tests were used (I,II,III), for example in the analysis of recovery data which were not assumed to be normally distributed. Then Kruskal-Wallis test was used for overall statistical analysis and Mann-Whitney U with Bonferroni correction was used for statistical comparisons (I). P-values <0.05 were considered to be significant. The computer software used for statistical analysis was StatView and SuperANOVA (Abacus Concepts Inc, Berkeley, CA, USA).

### 4. Résumé of **papers**

#### Paper I:

**Anesthetic modulation of the cardiovascular response to microlaryngoscopy.**  comparison of propofol and methohexital with special reference to leg blood flow, catecholamines and recovery.

Thirty-five (ASA physical status 1-2) patients (22-68 years) scheduled for microlaryngoscopy were studied. One patient participated twice but in different groups. Invasive arterial blood pressure, heart rate, stroke volume (impedance cardiography), leg blood flow (LBF; occlusion plethysmography) and catecholamine concentrations were measured. The patients were randomized with Pocock and Simons method for sequential allocation<sup>103</sup>, according to age and sex, to four groups receiving either a low or a high maintenance dose of propofol or methohexitone (9 patients in each group). The infusion rates of propofol were 6 or 12 mg·kg<sup>-1.</sup>h<sup>-1</sup> and of methohexitone  $5$  or  $10$  mg $\text{kg-1}\cdot\text{h-1}$ . These infusion rates were assumed to be equianaesthetic, based on studies defining ED50 and ED95 of the two drugs 104,105 . After a preanaesthetic dose of atropine  $(5 \mu g \cdot kg^{-1})$  and fentanyl  $(2 \mu g \cdot kg^{-1})$  $\mu$ g·kg<sup>-1</sup>), the anaesthesia was induced with either propofol  $(2.0 \text{ mg} \cdot \text{kg}^{-1})$  or methohexitone  $(1.5 \text{ mg} \cdot \text{kg}^{-1})$  depending on to which group of anaesthetic infusion during maintenance the patient was randomized. The patients were muscle relaxed with suxamethonium (bolus followed by an infusion), intubated with a naso-tracheal insufflation catheter and ventilated artificially with high frequency positive

pressure ventilation (HFPPV). With the laryngoscope fixed to a frame mounted on the operating table, the microlaryngoscopy started exactly 4 minutes after the initial injection of propofol or methohexitone. Measurements were made before injection of atropine (control), before intubation with the naso-tracheal ventilation catheter (-2 min) and 1, 5, 10 and 15 min after the start of microlaryngoscopy. Blood was also sampled for analysis of arterial catecholamines and drug concentrations, and arterial blood-gases. During the postoperative phase, various recovery parameters were noted.

#### *Results*

The groups were similar concerning haemodynamic data in the control state before anaesthesia. The low methohexitone infusion dose was insufficient to control MAP, which increased 41% (p<0.05) during the first 15 minutes of the microlaryngoscopy compared with the awake state (fig. 11). In the other groups, the corresponding increases were 11-22% (n.s.). The HR increased in all groups but the increase was most prominent in the low-dose methohexitone group.

There were no significant changes in cardiac output (CO) during microlaryngoscopy. Nor were significant differences in CO or systemic vascular resistance (SVR) found between the groups. However, compared with the control period, significant increases in SVR by 25-39% were noted at some points of measurements in the low-dose



*Figure. 11. Mean arterial blood pressure (MAP) and heart rate (HR) (mean ± SEM) at awake control ( C), after induction of anaesthesia (A) and during endoscopy (starts at 0) with infusion of*  propofol 6 or 12  $mg \cdot kg^{-1} \cdot h^{-1}$  (represented by P6 and P12) or methohexitone 5 or 10  $mg \cdot kg^{-1} \cdot h^{-1}$ *(M5, M10). Significant changes (p<0.05) compared to awake control are depicted by Significant differences (p<0.05) between groups are shown below each graph.* 

methohexitone and the low-dose propofol groups. LBF increased consistently in all groups, except in patients anaesthetized with the low dose of methohexitone (fig. 12). In the high-dose methohexitone group, on the other hand, LBF increased 300%, which was more than in the other groups. In contrast to methohexitone, the increases in LBF were similar with the low and high doses of propofol (110% above control values).' During endoscopy,

changes in LVR were essentially reciprocal to those in LBF (fig. 12). The methohexitone low-dose group was the only group which did not show a decrease in LVR. The methohexitone low-dose group showed increases in noradrenaline levels compared with awake values and in adrenaline levels compared with the other groups.



Figure. 12. Leg blood flow (LBF) and leg vascular *resistance (LVR) (mean ± SEM in percent of control values - 100) at awake control (C) and during endoscopy (starts at 0) with infusion of propofol 6 or 12 mg-kg' 1 -h~ <sup>1</sup>(P6, P12) or methohexitone* 5 or 10  $mg \cdot kg^{-1} \cdot h^{-1}$  (M5, M10). *Induction of anaesthesia is illustrated by A. Significant changes (p<0.05) compared to awake control are depicted by \*. Significant differences (p<0.05) between groups are shown below each graph.* 

		Eyes open on command		Breathing easily	Orientation in time and location		Recall of telephone nr		Leaving operating room		Leaving recovery room	
	mean	rance	mean	range	mean	range	mean	range	mean	range	mean	range
Propofol 6 mg·kg <sup>-1</sup> ·h <sup>-1</sup>	$3.6$ $*$	$(2-7)$		$4.1 * (2 - 10)$	4.4''	$(3-6)$	$4.4$ *	$(3-6)$		$9.1$ (7-14)	13.3	$(6-22)$
Propofol 12 mg·kg·1-h-1	7.4	$(2-13)$	7.4	$(2-15)$	8.1	$(3-13)$	8,2	$(3-13)$	12.7	$(9-18)$		$26.0$ $(10-50)$
Methohexital 5 mg·kg <sup>-1</sup> ·h <sup>-1</sup>	3.8''	$(0-8)$		$4.3$ $*(1-10)$	9.9	$(2-40)$	$5.2*$	$(2-9)$		$9.7 * (6-14)$	31.7	$(12-114)$
Methohexital 10 mg·kg <sup>-1</sup> ·h <sup>-1</sup>	11.3	$(4 - 18)$	13.6	$(3-30)$	18.6	$(5-40)$	23,3	$(5 - 60)$		$18.2$ (10-25)	47.4	$(16-71)$

*Table 2. Recovery parameters (min). Significant differences compared to the methohexitone high dose group* (10  $mg \cdot kg^{-1} \cdot h^{-1}$ ) are depicted by \*.

Recovery was significantly prolonged after high-dose methohexitone anaesthesia compared with the other groups (table 2).

#### Paper II:

#### The effects of propofol, methohexitone and isoflurane on the baroreceptor reflex in the cat.

In an experimental open-loop model in the cat (n=14), the effects on the baroreceptor reflex were studied during administration of either isoflurane (n=8), methohexitone (n=8) or propofol (n=8). Data during basal chloralose anaesthesia alone, obtained before administration of each additional anaesthetic, were used as control values. The order of administration of the three anaesthetics - isoflurane, propofol and methohexitone - was randomized. The anaesthetics were administered in a low and a high dose (table 3) corresponding to 0.5 and 1.0 MAC (minimum alveolar concentration) or assumed equipotent intravenous infusion rates. The baroreflex was tested by artificially changing the carotid sinus pressure from 50 to 200 mm Hg in steps of 25 mm Hg. This

	Low dose	High dose			
Propofol	4.5 mg $kg^{-1} h^{-1}$	9.0 mg $kg^{-1} \cdot h^{-1}$			
Methohexitone	$3.75$ mg kg <sup>-1</sup> h <sup>-1</sup>	7.5 mg $kg^{-1} \cdot h^{-1}$			
Isoflurane	0.80%	$1.6\%$			

*Table* 3. *Maintenance doses of propofol, methohexitone and isoflurane.* 

procedure was performed before the administration of each anaesthetic and repeated after 20-30 minutes at each dose level. An elimination period of 2 hours was allowed after the intravenous anaesthetics and 1 hour after isoflurane. A normalized baroreceptor reflex response was a prerequisite before introducing the next anaesthetic. Because of this requirement, many experiments had to be discontinued after one or two anaesthetics. Two cats had a maintained baroreflex reactivity before the administration of the third anaesthetic.

#### *Results*

Increases in carotid sinus pressure from 50 to 200 mm Hg were associated with decreases in MAP and HR. In the control state, the average baroreflex sensitivity, in terms of MAP response, was 74.9±6.2 mm Hg/mm Hg (mean±SEM), which corresponds to a gain of 3.0. The effects of the baroreceptor reflex tests on systemic MAP and HR before and during maintenance with low and high doses of the three anaesthetics are shown in figure 13. In the isoflurane and methohexitone groups, reflex sensitivity expressed either as MAP or HR responses was significantly depressed at the high dose level of the anaesthetic (fig. 14). In the isoflurane group, reflex sensitivity in terms of MAP response was also depressed with the low isoflurane dose. In the propofol group, there were no significant differences compared with the control state. With the high dose of the



*Figure. 13. Effects on systemic MAP and HR (means and SEM) of artificial unloading and overloading of the carotid sinuses (baroreceptor reflex test) before (=control) and during low and high doses of propofoi, methohexitone and isoflurane.* 

anaesthetics, reflex sensitivity in terms of either MAP or HR responses was significantly lower during isoflurane and methohexitone than during propofol anaesthesia. With the low dose, only reflex sensitivity in terms of MAP was significantly lower during isoflurane compared with propofoi anaesthesia. During isoflurane and methohexitone anaesthesia, the capacity of the baroreflex was attenuated in a similar way as the baroreflex sensitivity. The baroreflex set point for MAP was significantly higher during methohexitone than during propofoi anaesthesia and the set point for HR was significantly decreased for isoflurane compared with control values. In a subgroup of the animals, baroreflex sensitivity concerning MAP and HR substantially recovered within 10 minutes after discontinuation of the anaesthetics.



*Figure. 14. Baroreceptor reflex sensitivity - in systemic MAP and HR before (=control) and during low and high doses of propofoi ( P), methohexitone (M) and isoflurane (I). Means and SEM. Significant changes compared to control: \**   $p < 0.05$ .

#### Paper III:

#### Characteristics of muscle nerve sympathetic activity during general anaesthesia in humans.

Eighteen patients (ASA physical status 1- 2), aged 30-70 (mean 51) years, scheduled for elective otorhinolaryngeal surgery participated in this descriptive study investigating the effects of various anaesthetics, baroreflex stimuli and surgical stress on muscle nerve sympathetic activity (MSA), arterial blood pressure, heart rate and skin blood flows (laser Doppler flowmetry and finger pulse plethysmography).

#### *Results*

In the 6 patients who received a bolus injection of fentanyl  $(3 \mu g/kg)$  before the start of anaesthesia, mean arterial pressure decreased gradually from 112±6 (mean $\pm$ SEM) to 106 $\pm$ 9 mm Hg (p<0.05)

during the first 90 sec after the injection. Total MSA (defined as the product of mean burst amplitude and bursts/min) was unaltered apart from a transient increase in 5 of 6 patients one minute after the fentanyl injection.

During induction of anaesthesia with propofol (n=16) or methohexitone (n=2) MSA and blood pressure decreased in all patients (fig. 15). In a subgroup of 7 propofol patients the MSA burst frequency decreased to about 50% and total MSA to one-third of the control values, but the pulse-synchronous burst character was always maintained. An initial transient increase in MSA before the decrease started was confirmed in two of five patients by the cumulative sum (CUSUM) technique, which visualized the time relationship between changes of MSA and MAP.

Laryngoscopy and tracheal intubation initiated an immediate strong increase



*Figure. 15. Examples of changes of MSA and arterial blood pressure during induction of anaesthesia with bolus doses of propofol (upper panel, A) and methohexitone (lower panel, B). Note the druginduced progressive decreases of MSA and arterial blood pressure and the sudden increases of nerve traffic with tracheal intubation.* 



*Figure. 16. Change of pattern of MSA during intubation. MSA was pulse-synchronous before anaesthesia and immediately before intubation (although with low burst amplitudes). Intubation caused an instant increase in MSA and the activity became continuous (pulse-synchrony was lost and baseline elevated). Two minutes after intubation arterial blood pressure had increased to a stable level and the pulse* $synchrony$  of MSA had returned.

in MSA, followed by a slower gradual increase of blood pressure (fig. 16). In 9 of 13 patients the normal pulse-synchronous burst pattern was lost and replaced by an irregular burst pattern and in 12 of 13 patients the activity became continuous and the neurogram baseline was elevated. These changes gradually disappeared and normal pulse-synchrony and baseline levels re-occurred in all patients within 45 s after intubation. External manipulation of the tracheal tube or pharyngeal suction evoked effects similar to those evoked by the intubation procedure, but weaker and without loss of cardiac rhythmicity.

The anaesthetics in this study were used alone or in combinations. Table 4 summarizes the effects on MSA of

#### these anaesthetics.

Surgical stimulation as such (n=13) was always associated with an increase in MSA followed by increases in arterial blood pressure and heart rate. Ventilator-induced sighs induced a transient increase of MSA (fig. 17). Repeated every 5th minute during maintenance of anaesthesia, the sighs were used as an index of baroreflex/lung receptor reflex response. This response was most clear-cut during administration of 70% nitrous oxide alone, whereas the addition of 0.3 and  $0.6\%$  isoflurane (n=8) dose dependently seemed to reduce all components of the response. During undisturbed anaesthesia with 1.0 MAC isoflurane (1.2% end-tidal concentration) sigh-



*Table 4. The effects on MS^4 of different anaesthetics used in study III.* 

*Figur e. 17. Effects of ventilator induced sighs during 70% nitrous oxide alone, with addition of 0.6% (end-tidal conc) isoflurane and 1.2% (endtidal conc) isoflurane alone. The MSA response was successively reduced but reappeared during surgery with 1.2% (end tidal conc) isoflurane anaesthesia. The panels shown were obtained after at least ten minutes administration of each anaesthetic.* 



induced changes were minimal or abolished, whereas surgery at this isoflurane dose caused an increased basal MSA and a restored reflex response.

#### Paper IV:

Percutaneous recording of muscle nerve sympathetic activity during propofol, nitrous oxide and isoflurane anesthesia in humans.

Muscle nerve sympathetic activity (MSA), arterial blood pressure, heart rate (HR) and skin blood flows (laser Doppler flowmetry and finger pulse plethysmography, respectively) were recorded before and during anaesthesia in 8 ASA physical status 1 patients (31-70 yrs) scheduled for otorhinolaryngeal surgery. After preanaesthetic medication with rectal diazepam (0.25 mg/kg) the patients

arrived at the operating room. Recordings of all parameters were established before the start of anaesthesia. After induction with propofol and vecuronium, tracheal intubation and mechanical ventilation  $(F<sub>i</sub>O<sub>2</sub> 0.30)$  were performed. Anaesthesia was maintained according to the following protocol:

- 10 min with  $N_2O/O_2$
- 10 min with  $N_2O/O_2 + 0.3\%$  isoflurane (end-tidal concentration)
- 10 min with  $N_2O/O_2 + 0.6\%$  isoflurane (end-tidal concentration)
- 10 min with  $O<sub>2</sub>/air + 0.6%$  isoflurane (end-tidal concentration)
- 10 min with  $O_2 / air + 1.2%$  isoflurane (end-tidal concentration)

Results are presented as means ± SEM from a 5-min control period before induction of anaesthesia, during the minute before and the minute after intubation, and during the last 3 min of each anaesthetic period.

#### *Results:*

During the control period before the start of anaesthesia, MSA showed its usual character with irregular sequences of pulse-synchronous bursts (range 18-63 bursts/min). Bolus injection of propofol (2.6 mg/kg) was associated with a decrease in both MSA burst frequency and burst amplitude, although pulsesynchronous rhythmicity was maintained (fig. 18). The total MSA (calculated from the product of the number of bursts/min and mean burst amplitude) was reduced to  $34\pm2\%$  (p<0.05) of the awake control level (fig. 19). Laryngoscopy and tracheal intubation initiated an immediate increase in MSA (total MSA 151±23% of control value), which in 4 of 7 patients was associated

with a temporary loss of pulsesynchrony (during 13-37 s). Mean arterial blood pressure (MAP), which decreased from 99±16 mm Hg during awake control to  $73\pm10$  mm Hg ( $p<0.05$ ) after induction of anaesthesia, increased to 109±15 mm Hg after intubation. Heart rate (HR) showed gradual increases from 68±8 to 78±8 beats/min before and 86±10 beats/min after intubation. Skin blood flows increased after induction of anaesthesia to 700-1000% of control values (p<0.05), whereas intubation did not evoke additional changes. After 10 min ventilation with nitrous oxide alone, total MSA was increased to the same level as during the first minute after intubation, while MAP was decreased from 109±5 to 84±4 mm Hg  $(p<0.05)$ .



*Figure. 18. Experimental records from one patient during induction of anaesthesia with propofol. The upper part of the figure shows a condensed recording (time scale 60 sec). In the lower part, selected periods have been displayed at a faster speed (time scale 5 sec).* 



*Figure. 19. Mean values ± SEM of mean arterial blood pressure (MAP), heart rate (HR) and total MSA (bursts/minute x mean burst amplitude) of the whole material. Total* MSA *expressed in % of the awake control value (=100). Symbols \*, t and A indicate statistical significance (p<0.05) for comparisons with the awake control period (\*), with 70% nitrous oxide (t) and with 0.6% isoflurane + 70% nitrous oxide (A), respectively. MSA data immediately before and after intubation include measurements from 7 patients.* 

Supplementation of 70% of increasing doses of isoflurane (0.3% and 0.6% endtidal concentration) to nitrous oxide was associated with gradual decreases in total MSA and MAP (p<0.05 at 0.6%)(fig. 19). Withdrawal of nitrous oxide, which implied a decrease in depth of anaesthesia from 1.0 MAC to 0.5 MAC, was associated with a substantial decrease in total MSA (from 107±56 to  $61±42$  % of control level;  $p<0.05$ ) while MAP increased from 71±6 to 78±5 mm Hg (ns). Increase of isoflurane from 0.6% to 1.2% did not significantly change

MSA. Skin blood flows were stable during nitrous oxide and/or isoflurane anaesthesia, with laser Doppler blood flows of 1100-1300% and finger pulse amplitudes of 1600-1900% of the respective control values.

#### Paper V:

#### Sympathetic muscle nerve activity, peripheral blood flows and baroreceptor reflexes in humans during propofol anesthesia and surgery.

Nine (ASA physical status 1-2) patients, aged 26-59 yrs, scheduled for elective microlaryngoscopy were studied. Muscle nerve sympathetic activity, arterial blood pressure, heart rate, leg blood flow (occlusion plethysmography) and skin blood flows (laser Doppler flowmetry and finger pulse plethysmography) were recorded before and during undisturbed propofol anaesthesia  $(0.1 \text{ mg} \cdot \text{kg}^{-1} \cdot \text{min}^{-1})$ and during microlaryngoscopy when the propofol infusion dose was individually adjusted so that mean arterial blood pressure was kept at the awake control level. The patients were muscle relaxed with atracurium and artificially ventilated through a tracheal tube. During the microlaryngoscopy the laryngoscope was fixed to a frame mounted on the operating table. The sensitivity of the muscle sympathetic and cardiac limbs of the baroreflex were tested by measuring the compensatory responses to the blood pressure reduction induced by bolus injections of sodium nitroprusside (SNP). The baroreflex sensitivity was represented by the linear regression slope when blood pressure values were plotted against MSA-area (MSAA) or cardiac RRintervals.

#### *Results*

Induction of anaesthesia with propofol (on average 2.4 mg/kg including supplementary doses) reduced both the number of MSA bursts and their mean areas in all subjects. Four minutes after induction, total MSA (MSAA/min) was 39% of the awake control value ( $p<0.05$ ). This reduction was similar to the decrease in MSA noted during undisturbed steady-state propofol infusion 15-20 minutes later (fig. 20). However, during microlaryngoscopy when MAP was controlled close to the awake control level MSA was restituted. HR increased both during anaesthesia and

during microlaryngoscopy compared with the awake control state (fig. 21). Leg vascular resistance (derived from the ratio of MAP and LBF) decreased in parallel with MSA during anaesthesia, but in contrast to MSA the resistance remained decreased during microlaryngoscopy (fig. 20). This was reflected in part by a further increase in LBF during endoscopy when the propofol infusion dose was increased more than three-fold. The increase in blood flow also seen in laser Doppler skin blood flow and finger pulse amplitude during anaesthesia remained unchanged during microlaryngoscopy.





*Figure. 20. Mean values ± SEM (in percent of control = 100) of "total MSA" and leg vascular resistance (LVR) in the awake state (control), during undisturbed anaesthesia and microlaryngoscopy (surgery). Significant changes (p<0.05) are depicted by \*.* 

Figure. 21. Mean values  $\pm$  SEM of HR and MAP in *the awake state (control), during undisturbed anaesthesia and microlaryngoscopy (surgery). Significant changes (p<0.05) are depicted by \*.* 



*Figure. 22. Mean values ± SEM of slope values (i.e. regression coefficients) from RR-interval vs. MAPslopes (upper panels) and MSAA vs. DAP-slopes (lower panels) calculated from individual baroslopes with correlation coefficient exceeding 0.5. Slope values from depressor test periods only including the first 20 s are shown in the left panels and tests including the whole period are shown in the right panels. MSAA is expressed as percent of basal MSAA value during the awake control period. Significant changes*   $(p<0.05)$  are depicted by  $*$ .

During undisturbed anaesthesia, the sensitivity was depressed in both the cardiac and muscle sympathetic limb of the baroreflex (fig. 22). During microlaryngoscopy the cardiac baroreflex sensitivity was further depressed, while the muscle sympathetic limb was unchanged. These baroreflex changes were also obvious when the analysis of the depressor tests was restricted to the first 20 sec of fall in blood pressure, although the cardiac baroreflex sensitivity in general was weaker (fig. 22).



### 5. Discussion

#### Induction of anaesthesia and tracheal intubation

Induction of anaesthesia is usually associated with a decrease in arterial blood pressure. All patients in the present studies had anaesthesia induced with propofol or methohexitone. Decreases in blood pressure induced by these agents are usually ascribed to both decreased cardiac output and vasodilation<sup>106,107</sup>. The decrease in blood pressure is often more pronounced during induction of anaesthesia with propofol than with methohexitone. The heart rate usually increases during methohexitone anaesthesia<sup>106</sup> whereas propofol is associated with varying heart rate changes<sup>107</sup>. With both drugs, we found that induction of anaesthesia is followed by a decrease in postganglionic sympathetic nerve activity to the skeletal muscle vasculature. This decrease was closely associated with the concomitant decrease in arterial blood pressure. The reductions in total MSA found during propofol induction (IV,V) are similar to those recently observed by Ebert after induction with propofol<sup>108</sup> and thiopentone<sup>109</sup> . In contrast, etomidate did not seem to affect either MSA or blood pressure<sup>108</sup>.

The fact that the onset of the decrease in MSA is normally followed by a blood pressure decrease (III) illustrates the role of the sympathetic activity in maintenance of vascular tone. However, with the aid of the CUSUM technique<sup>71</sup>, we found that before the marked reductions in MSA and MAP occurred, some patients showed an initial transient

increase in MSA. This initial augmentation of MSA may be caused by a central excitation during the induction of anaesthesia or a baroreflex response to a blood pressure reduction evoked by the direct vasodilatory effects of methohexitone or propofol before anaesthesia had caused depression of central sympathetic outflow and baroreflex sensitivity. The occurrence of transient increases of MSA during induction of anaesthesia has also been illustrated by unpublished data in two out or four pilot patients who received a subanaesthetic bolus dose of propofol (1 mg/kg) followed by infusion of 10 mg $\text{kg-1}\cdot h^{-1}$  during the first 10 minutes (fig. 23).

Although the decrease in MSA after induction of anaesthesia with propofol or methohexitone was pronounced, MSA retained its pulse-synchronous pattern, indicating that baroreceptor reflex control of MSA was still operative.

The fact that stimulation of the pharynx and larynx increases sympathetic activity was demonstrated in the cat by Tomori and Widdicombe 110 . We found that laryngoscopy and tracheal intubation not only increased MSA but also induced short periods of continuous sympathetic activity (i.e. elevated mean voltage baseline) with or without loss of pulse synchrony in the neurogram. Pulse synchrony is caused by recurrent inhibitory influence from arterial baroreceptors and normally the inhibitory effect of each systolic pulse wave is strong enough to cause a short pause in the sympathetic neurogram<sup>65</sup>. In theory, continuous sympathetic activity and



*Figure. 23. Induction of anaesthesia with a subanaesthetic bolus dose of propofol (1 mgfkg) followed by an infusion (10 mg-kg<sup>-1</sup> h<sup>-1</sup>). Note the increase of MSA during the first part of the induction sequence and the gradual decrease in sympathetic outflow when anaesthesia is established. Due to technical reasons, the blood pressure amplitude fades during the late part of the recorded sequence but is re-established before the sequence is ended.* 

lack of pulse synchrony should occur if baroreceptor inhibition is too weak in relation to the central sympathetic drive. The effects of temporary baroreceptor deafferentiation with bilateral blockade of the glossopharyngeal and vagal nerves by local anaesthetics support this view<sup>67</sup> . In the present studies, blood pressure was low after induction of anaesthesia and therefore the baroreceptor inhibitory influence was probably too weak to induce significant pulse rhythmicity in the strong sympathetic outflow evoked by intubation. This agrees with our finding that preintubation blood pressure was lower in subjects who lost MSA cardiac rhythmicity during intubation. We found that the increase in arterial blood pressure had a longer duration than the increase in MSA (fig. 24). The results suggest that the initial response to tracheal intubation is neurogenic and

that increases in blood pressure lasting more than 5 minutes after the start of intubation are likely to be maintained by humoral substances.

#### Maintenance of anaesthesia

#### *Propofol*

Compared with the awake control level, the decrease in arterial blood pressure during undisturbed steady-state propofol anaesthesia was associated with a decrease in MSA and increases in skin and skeletal muscle blood flows. The corresponding decrease in leg vascular resistance is consistent with a reduced afterload, which has been proposed as one of several possible mechanisms causing propofol-induced hypotension<sup>111,112</sup>. In our studies, measurements

*Figure. 24. Muscle nerve sympathetic activity (MSA), mean arterial blood pressure (MAP) and heart rate (HR) during laryngeal stimulation by external manipulation of the larynx and gentle movements of the tracheal tube for 10-15 sec in one patient anaesthetized with 70% nitrous oxide.* 



of cardiac output were performed only after the start of microlaryngoscopy. It is, however likely that undisturbed propofol anaesthesia induced a decrease in cardiac output<sup>41,113-115</sup>. It has been discussed whether this decrease in cardiac output is caused by myocardial depression 41,113 or whether venodilation and decreased preload are mainly responsible for the decrease in stroke volume<sup>112,114-119</sup>.

#### *Isoflurane*

The haemodynamic response to isoflurane anaesthesia is usually characterized by a reduction of total peripheral resistance<sup>120-122</sup>. The inhibitory effect of isoflurane on sympathetic nerve activity has been demonstrated indirectly in humans, for instance by low levels of plasma catecholamines<sup>123</sup>, and directly in animals by decreases in postganglionic activity in renal and cervical nerves<sup>22,124,125</sup>. Our findings show that MSA is inhibited already at relatively low doses of isoflurane and provide a contributory explanation for the decrease of vascular resistance.

#### *Halothane*

The effects of halothane on sympathetic outflow have been disputed since Millar and Biscoe<sup>126</sup> found an increase in sympathetic activity during halothane anaesthesia in the rabbit. Later, several studies in the cat led to the opposite conclusion<sup>27,30,127,128</sup>. However, large interindividual variations may occur<sup>30</sup>. Our observations are few and thus difficult to interpret. It is possible that in some patients administration of halothane causes an initial baroreflexmediated increase in MSA when direct cardiovascular depression by halothane precedes the depression of baroreceptor reflex sensitivity.

#### *Nitrous oxide*

Most anaesthetics cause various degrees of inhibition of the sympathetic outflow but nitrous oxide is an exception. The concept that nitrous oxide is a centrally acting stimulant of sympathetic activity $127,129-133$  is now supported by percutaneous recordings of increased MSA during nitrous oxide administration

both in studies by us (III,IV) and Ebert et  $al<sup>134,135</sup>$ . It can be argued that the increase in MSA demonstrated by Ebert during spontaneous ventilation with 40% nitrous oxide might be due to mental excitation and a changed respiratory pattern. However, our results were similar although the patients were anaesthetized, muscle relaxed and artificially ventilated. Withdrawal of nitrous oxide during stable isoflurane anaesthesia markedly decreased MSA in spite of a decreased MAC. Indeed, the interaction between nitrous oxide and isoflurane on MSA illustrates one aspect of so-called "balanced anaesthesia"<sup>136</sup>. It is also obvious that MSA may be a poor monitor of depth of anaesthesia during "balanced anaesthesia".

#### *Fentanyl*

Clinically, opiates have an important role in a balanced anaesthesia. We studied fentanyl, which as a sole agent is known to have only minor cardio-

vascular effects<sup>137</sup>. No significant change in MSA was observed after preanesthetic administration of small doses of fentanyl (3  $\mu$ g/kg). A lack of opiate interference with MSA during undisturbed conditions may seem consistent with the finding that the opiate antagonist naloxone at rest does not seem to affect haemodynamics or MSA<sup>24</sup>. However, during general anaesthesia and surgery the sympathetic effects of opiates may be more pronounced. This question remains to be studied.

#### Surgical stress

#### *Propofol and methohexitone*

We have shown that surgical stimuli increase MSA (fig.  $25$ )(III,V). This is not surprising since it is well known that arterial blood pressure and plasma catecholamines can increase promptly during surgery<sup>138</sup>. The restitution of



*Figure. 25. Effects of muscle nerve sympathetic activity (MSA) and arterial blood pressure ( MAP) of repeated reposition attempts of a*  zygomatic bone fracture in a *patient anaesthetized with methohexitone (4 mg* $kg^{-1} \cdot h^{-1}$ .

RESP. MOVEMENTS MSA during microlaryngoscopy, in spite of a large increase of the propofol infusion rate  $(V)$ , illustrates the balance between the stimulatory effect of somatosensory afferents on sympathetic outflow and the inhibitory effect of anaesthetics. Since leg blood flow increased and leg vascular resistance decreased compared with awake control values, the most likely interpretation is that propofol also has marked direct vasodilatory effects which override the neural vasoconstriction during surgery. The vasorelaxant effects of propofol have been demonstrated both in isolated rat aorta 112,116 and in capacitance vessels<sup>116-118</sup>. The effects on the capacitance vessels are already evident at low propofol concentrations and a dose relationship has been established in vitro for human vessels. These vascular effects of propofol which mimic the vasodilating effects of nitroglycerine may explain why very high propofol doses are sometimes needed to block the pressor response to surgical stress (V).

When methohexitone was compared with propofol in assumed equianaesthetic doses, the differences in leg blood flow between the low and high doses were large in the methohexitone but none in the propofol groups. Our observations concerning methohexitone, a barbiturate, are supported by studies on thiopentone in an isolated cat skeletal muscle preparation by Grände et al<sup>139</sup>. These authors showed that thiopentone at low concentrations increased vascular resistance, probably due to a decreased metabolic demand and a reduced local concentration of vasodilating metabolites<sup>140,141</sup>, whereas higher concentrations decreased vascular resistance and myogenic vascular reactivity in a dose-dependent manner. The methohexitone effects on MSA may be similar to those of propofol, since two patients anaesthetized with methohexitone infusion

showed a dose-related depression of MSA during surgery (III).

Catecholamine concentrations have often been used as an index of increased sympathetic nervous activity during tracheal intubation and surgical stress<sup>138,142-144</sup>. MSA has not previously been used as an index of perioperative stress, but stress induced by isometric hand grip elicits parallel increases in MSA and venous plasma noradrenaline concentrations<sup>145</sup> . Our material does not allow such comparisons but, compared with the awake state, we observed that as long as the pressor response to stress was blocked, there were no increases in arterial noradrenaline concentrations (I) or in MSA (V).

#### Baroreceptor reflexes

During anaesthesia, pharmacological effects of the anaesthetic drugs on baroreflex sensitivity influence the ability of the cardiovascular system to buffer sudden changes in blood pressure. In our studies, the baroreceptor reflex was evaluated in an experimental baroreflex open-loop model (II) and in humans through either ventilatory induced circulatory changes (III) or SNPinduced hypotension (V). The baroreflex tests were performed either during steady-state anaesthesia at undisturbed conditions or during surgical stress. The studies focused on the baroreflex effects of propofol, methohexitone and isoflurane.

#### *Propofol*

Propofol maintained the baroreflex sensitivity better than methohexitone and isoflurane at assumed equianaesthetic doses when tested in vagotomized cats (II). These results may seem surprising since in the clinical setting both propofol and isoflurane are often

associated with marked decreases in blood pressure<sup>47,107</sup>. However, our findings are in agreement with previous studies which showed that the response in heart rate to a change in arterial blood pressure was not affected by propofol in humans 146,147 and was less affected by propofol than by other anaesthetics (althesin, pentobarbitone and ketamine) in the rabbit<sup>148</sup>. However, extrapolation of the sensitivity diagram in our study (II) also in dicates that propofol in higher doses than used in this study may cause significant depression of the baroreceptor reflex.

In the cat, the unchanged cardiac baroreflex sensitivity during propofol anaesthesia was assumed to be due to a relatively unaffected cardiac sympathetic response since the vagal influence, which is normally the most important heart rate modulating factor<sup>9</sup>, was intentionally abolished. The vagal influence is therefore likely to explain why cardiac baroreflex sensitivity in our human study (V) was markedly depressed by propofol.

Concerning the muscle sympathetic limb of the baroreflex during undisturbed propofol anaesthesia, the sensitivity was significantly depressed both in Ebert's<sup>108</sup> and in our study (V). During microlaryngoscopy, the baroreflex remained depressed. We suggest that this is explained by the pharmacological effects of the high dose of propofol during this procedure. For instance, it is possible that direct relaxant effects of the very high propofol doses on the baroreceptorharbouring vessels<sup>112,116</sup> change afferent baroreceptor nerve activity during baroreflex tests.

In contrast to muscle sympathetic baroreflex sensitivity, the sensitivity in the cardiac baroreflex limb was further depressed during surgery. This is consistent with observations of a selective

inhibition of the cardiac baroreflex during hypothalamic defence area stimulation<sup>13-15</sup> which could mimic the cardiovascular response to intense pain 12 .

#### *Methohexitone*

Barbiturates have previously been shown to depress the baroreceptor reflex<sup>25-27,149</sup>. This was confirmed in the cat study (II), in which the baroreflex sensitivity was significantly more depressed during methohexitone than during propofol anaesthesia. In a microneurographic study concerning the baroreflex sensitivity, MSA burst frequency was depressed by thiopentone 109 . Our MSA recordings in two patients with methohexitone infusion were not quantified but showed, at infusion rates of 4 mg $\cdot$ kg<sup>-1</sup> $\cdot$ h<sup>-1</sup>, an abolished baroreflex response to a valsalva-like manoeuvre or to 15 cm of positive end-expiratory pressure (PEEP).

#### *Isoflurane*

Experimentally, isoflurane anaesthesia was also associated with a depression of baroreflex sensitivity in terms of both heart rate and pressure responses (II). This is consistent with previous observations<sup>22,32,34,35,125</sup>. Isoflurane seems to act at multiple sites in the baroreceptor reflex arch. This includes both a dose-dependent sensitisation of the baroreceptors causing a decrease in efferent preganglionic sympathetic activity and a ganglionic depression causing a further decrease in efferent postganglionic sympathetic activity<sup>22</sup>. Our experimental findings in the cat are qualitatively consistent with our observations of MSA during isoflurane anaesthesia in patients (III). In patients, baroreflex responses evoked by ventilator-induced sighs were abolished at 1.2 % end-tidal concentration of isoflurane. With the influence of

surgical stress, however, baroreflex responses were restored.

#### *Summary of baroreflex effects*

When propofol is used as reference, human and animal data indicate that during undisturbed anaesthesia the sympathetic baroreflex will be less affected by etomidate<sup>108</sup> and nitrous oxide<sup>135</sup> and more depressed by methohexitone (II,III) and isoflurane (II,III). When the pressor response during microlaryngoscopy is controlled by high doses of propofol, the sympathetic baroreflex sensitivity shows about the same depression as during undisturbed anaesthesia. During surgical stress, the balance between different cardiovascular regulatory mechanisms in relation to anaesthetic interactions is complex and therefore not always recognized. Further investigations of these issues are thus of importance for anaesthetic management and care.



### 6. Conclusions

- Sympathetic activity to skeletal muscle (MSA) is depressed by propofol, methohexitone and isoflurane, whereas nitrous oxide is associated with an increase in MSA.
- Vasodilation during propofol anaesthesia may be caused both by an inhibition of sympathetic nerve traffic and by a direct vascular effect.
- In the cat, the arterial baroreceptor reflex is better maintained with propofol than with methohexitone or isoflurane in equianaesthetic doses.
- During anaesthesia, MSA retains its normal pulse-synchronous character, even when the activity is depressed. This indicates that arterial baroreceptor modulation is qualitatively operative.
- The cardiovascular response to microlaryngoscopy is better controlled by a low dose propofol infusion than by an equianaesthetic infusion of methohexitone. The recovery is more rapid after a high dose propofol infusion than after an equianaesthetic infusion of methohexitone.
- The depression of MSA during undisturbed propofol infusion is to a large extent restored during microlaryngoscopy in spite of a more than three-fold increase of the propofol infusion rate.
- Baroreflex sensitivities of both cardiac and muscle nerve sympathetic limbs are depressed by propofol. During surgery, a further depression of the cardiac baroreflex may be due to a central vagal inhibition similar to that found in animals during defence area stimulation.
- The baroreflex sensitivity is determined by a balance between an augmented central sympathetic outflow due to the intensity of surgical stimulation and inhibition due to the anaesthetic dose.



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### 8. References

- 1. Shepherd J T, Abboud F M. Handbook of physiology, The cardiovascular system, vol 3, Peripheral circulation and organ blood flow, part 2. Bethesda: American Physiological Society, 1983.
- 2. Smith JJ, Kampine JP. Circulatory physiology - the essentials. (3 ed.) Baltimore: William & Wilkins, 1990
- 3. Folkow B. Intravascular pressure as a factor regulating the tone of the small vessels. Acta Physiol Scand 1949;17:289-310.
- 4. Grande P-O. Myogenic mechanisms in the skeletal muscle circulation. J Hypertension 1989;7 (suppl 4):S47-S53.
- 5. Meliander S, Johansson B. Control of resistance, exchange and capacitance functions in the peripheral circulation. Pharmacol Rev 1968;20:117-196.
- 6. Borgström P, Grande P-O, Meilander S. An evaluation of the metabolic interaction with myogenic vascular reactivity during blood flow autoregulation. Acta Physiol Scand 1984;122:275-284.
- 7. Shepherd JT. Circulation to skeletal muscle. In: Geiger S, ed. Handbook of physiology, The cardiovascular system, vol 3. Washington: American Physiological Society, 1983: 319-370.
- 8. Henriksen O, Amtorp O, Paris I, Agerskov K. Evidence for a local sympathetic venoarteriolar reflex in the dog hindleg. Circ Res 1983;52:534-542.
- 9. Koizumi K, Terui N, Kollai M. Effect of cardiac vagal and sympathetic nerve activity on heart rate in rhythmic fluctuations. J Autonom Nerv Syst 1985;12:251-259.
- 10. Somers VK, Mark AL, Zavala DC, Abboud FM. Contrasting effects of hypoxia and hypercapnia on ventilation and sympathetic activity in humans. J Appl Physiol 1989;67:2101-2106.
- 11. Somers VK, Mark AL, C ZD, Abboud FM. Influence of ventilation and hypocapnia on sympathetic nerve response to hypoxia in normal humans, J Appl Physiol 1989;67:2095-2100.
- 12. Johansson B. Circulatory responses to stimulation of somatic afferents. Acta Physiol Scand 1962;57 (Suppl. 198):1-91.
- 13. Djojosugito AM, Folkow B, Kylstra PH, Lisander B, Tuttle RS. Differentiated interaction between the hypothalamic defence reaction and baroreceptor reflexes. I. Effects on heart rate and regional flow resistance. Acta Physiol Scand 1970;78:376- 385.
- 14. Hilton SM. Inhibition of baroreceptor reflexes on hypothalamic stimulation. J Physiol (Lond) 1963;165:56-57.
- 15. Gebber GL, Snyder DW. Hypothalamic control of baroreceptor reflexes. Am J Physiol 1970;218:124-131.
- 16. Winsö O, Biber B, Martner J. Does dopamine suppress stress-induced intestinal and renal vasoconstriction? Acta Anaesthesiol Scand 1985;29:508-514.
- 17. Östman M, Biber B, Martner J, Reiz S. Influence of isoflurane on renal and intestinal vascular responses to stress. Br J Anaesth 1986;58:630-638.
- 18. Biber B, Martner J. Hemodynamic consequences of defence area stimulation and afferent somatic nerve stimulation during fentanyl-nitrous oxide anesthesia. Modifying effects of droperidol. Acta Anaestheiol Scand 1981;25:336-343.
- 19. Prys-Roberts C. The circulation in 30. anaesthesia.Oxford: Blackwell Scientific Publications, 1980
- 20. Martner J, Biber B. Anaesthesia and cardiovascular regulation. Acta Anaesthiol Scand 1982;26 (Suppl 76):20-31. 31.
- 21. Feldman S, van der Heide C, Porter R. Evoked potentials in the hypothalamus. Am J Physiol 1959;196:1163-1167.
- 22. Seagard JL, Elegbe EO, Hopp FA, et al. Effects of isoflurane on the baroreceptor reflex. Anesthesiology 1983;59:511-520.
- 23. Savege TM, Dubois M, Frank M, Holly JMP. 33. Preliminary investigation into a new method of assessing the quality of anaesthesia: The cardiovascular response to a measured noxious stimulus. Br J Anaesth 1978;50:481-488.
- 24. Farrell PA, Ebert TJ, Kampine JP. Naloxone augments muscle sympathetic nerve activity during isometric exercise in humans. Am J Physiol 1991;260 (Endocrinol. Metab. 23):E379-E388.
- 25. Carter JA, Clarke TNS, Prys-Roberts C, Spelina KR. Restoration of baroreflex control of heart rate during recovery from anaesthesia. Br J Anaesth 1986;58:415 -421.
- 26. Schumacher IG, Arndt JO. Der effekt von methohexital, fentanyl, dehydrobenzperidol sowie von chloralose auf die aktivität der barorezeptoren des aortenbogens decerebrierter katzen. Anaesthesist 37. 1978;27:10-20.
- 27. Skovsted P, Price ML, Price L. The effects of short-acting barbiturates on arterial pressure, preganglionic sympathetic activity and barostatic reflexes. 38. Anesthesiology 1970;33:10-18.
- 28. Biscoe TJ, Millar RA. The effect of cyclopropane, halothane and ether on sympathetic ganglionic transmission. Br J Anaesth 1966;38:3-12. 39.
- 29. Duke PC, Fownes D, Wade JG. Halothane depresses baroreflex control of heart rate in man. Anesthesiology 1977;46:184-187.
- Skovsted P, Price ML, Price HL. The effects of halothane on arterial pressure, preganglionic sympathetic activity and barostatic reflexes. Anesthesiology 1969;31:507-514.
- Seagard JL, Hopp FA, Donegan JH, Kalbfleisch JH, Kampine JP. Halothane and the carotid sinus reflex. Anesthesiology 1982;57:191-202.
- 32. Takeshima R, Dohi S. Comparison of arterial baroreflex function in humans anesthetized with enflurane or isoflurane. Anesth Analg 1989;69:284-290.
- Morton M, Duke PC, Ong B. Baroreflex control of heart rate in man awake and during enflurane and enflurane-nitrous oxide anesthesia. Anesthesiology 1980;52:221- 223.
- 34. Kotrly KJ, Ebert TJ, Vucins E, Igler FO, Barney JA, Kampine JP. Baroreceptor reflex control of heart rate during isoflurane anesthesia in humans. Anesthesiology 1984;60:173-179.
- 35. Bagshaw RJ, Cox RH. Baroreceptor control of central and regional hemodynamics with isoflurane in the dog. Acta Anaesthesiol Scand 1988;32:82-92.
- 36. Millar RA, Warden JC, Cooperman LH, Price HL. Further studies of sympathetic actions of anaesthetics in intact and spinal animals. Br J Anaesth 1970;42:366-377.
- Fukunaga AF, Epstein R. Effects of cyclopropane on the sympathetic nervous system and on neural regulation of circulation in the cat. Anesthesiology 1974;40:323-335.
- Adam HK, Briggs LP, Bahar M, Douglas EJ, Dundee JW. Pharmacokinetic evaluation of ICI 35868 in man. Single induction doses with different rates of injection. Br J Anaesth 1983;55:97-102.
- Heath PJ, Ogg TW, Gilks WR. Recovery after day-case anaesthesia. A 24-hour comparison of recovery after thiopentone or propofol anaesthesia. Anaesthesia 1990;45(11):911-915.
- 40. Sanders LD, Clyburn PA, Rosen M, Robinson JO. Propofol in short gynaecological procedures. Comparison of recovery over 2 days after anaesthesia with propofol or thiopentone as sole anaesthetic agent. Anaesthesia-1991;46(6):451-455.
- 41. Brüssel T, Theissen JL, Vigfusson G, Lunkenheimer PP, Van Aken H, Lawin P. Hemodynamic and cardiodynamic effects of propofol and etomidate: Negative inotropic properties of propofol. Anesth Analg 1989;69:35-40.
- 42. Henriksson B-Å, Carlsson P, Hallén B, Hägerdal M, Lundberg D, Pontén J. Propofol vs thiopentone as anaesthetic agents for short operative procedures. Acta Anaesthesiol Scand 1987;31:63-66.
- 43. Fahy LT, Van Mourik GA, Utting JE. A comparison of the induction characteristics of thiopentone and propofol (2, 6-diisopropyl phenol). Anaesthesia 1985;40:939-944.
- 44. Rolly G, Versichelen L. Comparison of propofol and thiopentone for induction of anaesthesia in premedicated patients. Anaesthesia 1985;40:945-948.
- 45. Magnusson H, Pontén J, Sonander HG. Methohexitone anaesthesia for microlaryngoscopy: Circulatory modulation with metroprolol and dihydralazine. Br J Anaesth 1986;58:976-982.
- 46. Wade JG, Stevens WC. Isoflurane: An anesthetic for the eighties? Anesth Analg 1981;60:666-682.
- 47. Eger E. Isoflurane: A review. Anesthesiology 1981;55:559-576.
- 48. Munson ES, Eger EI, Tham MK, Embro WJ. Increase in anesthetic uptake, excretion, and blood solubility in man after eating. Anesth Analg 1978;57:224-231.
- 49. Sellgren J, Pontén J, Wallin BG. Percutaneous recording of muscle nerve sympathetic activity during propofol, nitrous oxide and isoflurane anesthesia in man. Anesthesiology 1988;69:A639 (Abstract).
- 50. Henriksson B-Å, Biber B, Lundberg D, Martner J, Pontén J. Cardiovascular studies during controlled baroreflex activation in the dog: II. Effects of metoprolol and enflurane. Acta Anaesthesiol Scand 1985;29:95-100.
- 51. Henriksson B-Å, Biber B, Martner J, Pontén J, Werner O. Cardiovascular studies during controlled baroreflex activation in the dog: I. Effects of enflurane. Acta Anaesthesiol Scand 1985;29:90-94.
- 52. Biber B, Martner J, Werner O. Modification by baroreceptor feedback of circulatory responses to noxious stimuli during anaesthesia in cats. Acta Anaesthesiol Scand 1983;27:391-395.
- 53. Cox RH. Influence of chloralose anesthesia on cardiovascular function in trained dogs. Am J Physiol 1972;223:660-667,
- 54. Armstrong GG, Porter H, Langston JB. Alteration of carotid occlusion response by anesthesia. Am J Physiol 1961;201:897-900.
- 55. Hagbarth K-E, Vallbo ÅB. Pulse and respiratory grouping of sympathetic impulses in human muscle nerves. Acta Physiol Scand 1968;74:96-108.
- 56. Delius W, Hagbarth K-E, Hongell A, Wallin BG. Manoeuvres affecting sympathetic outflow in human muscle nerves. Acta Physiol Scand 1972;84:82-94.
- 57. Delius W, Hagbarth K-E, Hongell A, Wallin BG. General characteristics of sympathetic activity in human muscle nerves. Acta Physiol Scand 1972;84:65-81.
- 58. Hagbarth K-E, Hallin RG, Hongell A, Torebjörk HE, Wallin BG. General characteristics of sympathetic activity in human skin nerves. Acta Physiol Scand 1972;84:164-176.
- 59. Delius W, Hagbarth K-E, Hongell A, Wallin BG. Manoeuvres affecting sympathetic outflow in human skin nerves. Acta Physiol Scand 1972;84:177-186.
- 60. Vallbo ÅB, Hagbarth K-E, Torebjörk HE, Wallin BG. Somatosensory, proprioceptive and sympathetic activity in human peripheral nerves. J Physiol Reviews 1979;59(4):919-957.

#### *8. References*

- 61. Walther O, Iriki M, Simon E. Antagonistic changes of blood flow and sympathetic activity in different vascular beds following central thermal stimulation. II. Cutaneous and visceral sympathetic activity during spinal heating and cooling in anaesthetized rabbits and cats. Pliigers Arch Physiol 1970;319:162-184.
- 62. Wallin GB, Sundlöf G, Delius W. The effect of carotid sinus nerve stimulation on muscle and skin nerve sympathetic activity in man. Pflügers Arch 1975;358:101-110.
- 63. Bini G, Hagbarth KE, Hynninen P, Wallin BG. Thermoregulatory and rhythmgenerating mechanisms governing the sudomotor and vasocontrictor outflow in human cutaneous nerves. J Physiol (Lond) 1980;306:537-552.
- 64. Wallin BG, König U. Changes of skin nerve sympathetic activity during induction of general anaesthesia with thiopentone in man. Brain Research 1976;103 suppl:157- 160.
- 65. Sundlöf G, Wallin BG. Human muscle nerve sympathetic activity at rest. Relationship to blood pressure and age. J Physiol (Lond) 1978;274:621-637.
- 66. Fagius J, Wallin BG. Sympathetic reflex latencies and conduction velocities in normal man. J of the Neurological Sciences 1980;47:433-448.
- 67. Fagius J, Wallin BG, Sundlöf G, Nerhed C, Engelsson S. Sympathetic outflow in man after anaesthesia of the glossopharyngeal and vagus nerves. Brain 1985;108:423-438.
- 68. Sundlöf G, Wallin BG. Effect of lower body negative pressure on human muscle nerve sympathetic activity. J Physiol 1978;278:525-532.
- 69. Sundlöf G, Wallin BG. The variability of muscle nerve sympathetic activity in resting recumbent man. ] Physiol (Lond) 1977;272:383-397.
- 70. Hornyak M, Cejnar M, Elam M, Matousek M, Wallin BG. Sympathetic muscle nerve activity during sleep in man. Brain 1991;114:1281-1295.
- 71. Ellaway PH. An application of cumulative sum technique (cusums) to neurophysiology. J Physiol (Lond) 1976;265:1P-2P.
- 72. Smyth HS, Phil D, Sleight P, Pickering GW. Reflex regulation of arterial pressure during sleep in man. Circ Res 1969;24:109- **121.**
- 73. Kotrly KJ, Ebert TJ, Vucins EJ, Roerig DL, Kampine JP. Baroreceptor reflex control of heart rate during morphine sulfate, diazepam, N20/02 anesthesia in humans. Anesthesiology 1984;61:558-563.
- 74. Sleight P. Methodology of baroreflex testing. G Ital Cardiol 1992;22:19-25.
- 75. Brodie TG, Russell AE. On the determination of the rate of blood-flow through an organ. J Physiol 1905;32:47-49.
- 76. Whitney RJ. The measurement of volume changes in human limbs. J Physiol 1953;121:1-27.
- 77. Pauca AL, Hopkins AM. Acute effects of halothane, nitrous oxide and thiopentone on the upper limb blood flow. Br J Anaesth 1971;43:326-334.
- 78. Lentner C, ed. Ceigy Scientific Tables; Heart and circulation. Basel: Ciba-Geigy Limited, 1990:278.
- 79. Greenfield ADM, Whitney RJ, Nowbray JF. Methods for the investigation of peripheral blood flow. Br Med Bull 1963;19:101-109.
- 80. Kubicek WG, Karnegis JN, Patterson RP, Witsoe DA, Mattson RH. Development and evaluation of an impedance cardiac output system. Aerospace Medicine 1966;37:1208- **1212.**
- 81. Geddes LA, Sadler C. The specific resistance of blood at body temperature. Med & Biol Eng 1973;11:336-339.
- 82. Porter JM, Swain ID. Measurement of cardiac output by electrical impedance plethysmography. J Biomed Eng 1987;9:222- 231.
- 83. Milsom I, Forssman L, Biber B, Dottori O, Sivertsson R. Measurement of cardiac stroke volume during cesarean section: A comparison cardiography and the dye dilution technique. Acta Anaesthesiol Scand 1983;27:421-426.
- 84. Ekman L-G, Milsom I, Arvidsson S, Biber B, Martineil S, Sjöqvist B-A. Clinical evaluation of an ensemble-averaging impedance cardiograph for monitoring stroke volume during spontaneous breathing. Acta Anaesthesiol Scand 1990;34:190-196.
- 85. Riva C, Ross B, Benedek GB. Laser-Doppler measurements of blood flow in capillary tubes and retinal arteries. Invest Ophtalmol 1972;11:936-944.
- 86. Holloway GA, Watkins DW. Laser-doppler measurement of cutaneous blood flow. J Invest Dermatol 1977;69:306-309.
- 87. Tenland T. On laser doppler flowmetry methods and microvascular applications. Thesis. Linköping University, Linköping, Sweden, 1982.
- 88. Nilsson GE, Tenland T, Öberg PÅ. Evaluation of a laser doppler flowmeter for measurement of tissue blood flow. IEEE Trans Biomed Eng 1980;27:597-604.
- 89. Nilsson GE, Tenland T, Öberg PÅ. A new instrument for continuous measurement of tissue blood flow by light beating spectroscopy. IEEE Trans Biomed Eng 1980;27:12-19.
- 90. Tooke JE, Östergren J, Fagrell B. Syncronous assessment of human skin micocirculation by laser doppler flowmetry and capillaroscopy. Int J Microcir Clin Exp 1983;2:277-284.
- 91. Johnson JM, Taylor WF, Shepherd AP, Park MK. Laser-doppler measurement of skin blood flow: comparison with plethysmography. J Appl Physiol 1984;56:798-803.
- 92. Hertzman AB. The blood supply of various skin areas as estimated by the photoelectric phethysmograph. Am J Physiol 1938;124:328-340.
- 93. Challoner AVJ. Photoelectric plethysmography for estimating cutaneous blood flow. In: Rolfe P, ed. Non invasive physiological measurements. Vol 1. London: Academic Press, 1979: 125-151. vol 1).
- 94. Lindberg L. Photoplethysmography. Methodological studies and applications. Linköping, 1991.
- 95. Zweifler AJ, Cushing G, Conway J. The relationship between pulse volume and blood flow in the finger. Angiology 1967;18:591-598.
- 96. Matsumura M. Comparative study on various methods of finger plethysmography, Tohoku J Exp Med 1968;94:337-346.
- 97. Thune P. Plethysmographie recordings of skin pulses. II. Piezoelectric and photoelectric measurements in psoriasis. Acta Dermatovenereol (Stockholm) 1970;50:263- 269.
- 98. Hales JRS, Stephens FRN, Fawcett AA, et al. Observations on a new non-invasive monitor of skin blood flow. Clin Exp Pharm Physiol 1989;16:403-415.
- 99. Hallman H, Farnebo L-O, Hamberger B, Jonsson G. A sensitive method for determination of plasma catecholamines using liquid chromatography with electrochemical detection. Life Sei 1978;23:1049-1052.
- 100. Hjelmdahl P. Catecholamine measurements by high-performance liquid chromatography. Am J Physiol 1984;247 (Endocrinol. Metab. 10):E13-E20.
- 101. Adam HK, Douglas EJ, Plummer GF, Cosgrove MB. Estimation of ICI 35,868 (Diprivan®) in blood by high-performance liquid chromatography, following coupling with Gibbs' reagent. J Chromatography 1981;223:232-237.
- 102. Redke F, Björkman S, Rosberg B. Pharmacokinetics and clinical experience of 20-h infusions of methohexitone in intensive care patients with postoperative pyrexia. Br J Anaesth 1991;66:53-59.
- 103. Pocock SJ, Simon R. Sequential treatment assignment with balancing for prognostic factors in the controlled clinical trial. Biometrics 1975;31:103-115.
- 104. Sear JW, Phillips KC, Andrews CJH, Prys-Roberts C. Dose-response relationship for infusions of althesin or methohexitone. Anaesthesia 1983;38:931-936.
- 105. Spelina KR, Coates DP, Monk CR, Prys-Roberts C, Norley I, Turtle MJ. Dose requirements of propofol by infusion during nitrous oxide anaesthesia in man. I: Patients premedicated with morphine sulphate. Br J Anaesth 1986;58:1080-1084.
- 106. Dundee JW, Wyant GM. Barbiturates: pharmacodynamics. In: Intravenous Anaesthesia. 2 ed. Edinburgh: Churchill Livingstone, 1988: 97-134.
- 107. Sebel PS, Lowdon JD. Propofol: A new intravenous anesthetic. Anesthesiology 1989;71:260-277.
- 108. Ebert TJ, Muzi M, Berens R, Goff D, Kampine JP. Sympathetic responses to induction of anesthesia in humans with propofol or etomidate. Anesthesiology 1992;76(5):725- 33.
- 109. Ebert TJ, Kanitz DD, Kampine JP. Inhibition of sympathetic neural outflow during thiopental anesthesia in humans. Anesth Analg 1990;71:319-326.
- 110. Tomori Z, Widdicombe JG. Muscular, bronchomotor and cardiovascular reflexes elicited by mechanical stimulation of the respiratory tract. J Physiol (Lond) 1969;200:25-49.
- 111. Claeys MA, Gepts E, Camu F. Haemodynamic changes during anaesthesia induced and maintained with propofol. Br J Anaesth 1988;60:3-9.
- 112. Park WK, Lynch C, Johns RA. Effects of propofol and thiopental in isolated rat aorta and pulmonary artery. Anesthesiology 1992;77:956-963.
- 113. Coetzee A, Fourie P, Coetzee J, et al. Effect of various propofol plasma concentration on regional myocardial contractility and left ventricular afterload. Anesth Analg 1989;69:473-483.
- 114. Lepage JYM, Pinaud ML, Hélias H, et al. Left ventricular function during propofol and fentanyl anesthesia in patients with coronary antery disease: Assessement with a radionuclide approach. Anesth Analg 1988;67:949-955.
- 115. Lepage JYM, Pinaud ML, Helias JH, Cozian AY, Le Normand Y, Souron J. Left ventricular performance during propofol or methohexital anesthesia: Isotopic and invasive cardiac monitoring. Anesth Analg 1991;73:3-9.
- 116. Bentley GN, Gent JP, Goodchild CS. Vascular effects of propofol: smooth muscle relaxation in isolated veins and arteries. J Pharm Pharmacol 1989;41:797-798.
- 117. Goodchild CS, Serrao JM. Cardiovascular effects of propofol in the anaesthetized dog. Br J Anaesth 1989;63:87-92.
- 118. Muzi M, Berens RA, Kampine JP, Ebert TJ. Venodilation contributes to propofolmediated hypotension in humans. Anesth Analg 1992;74(6):877-883.
- 119. Ismail EF, Kim S, Salem M, Crystal GJ. Direct effects of propofol on myocardial contractility in In situ canine hearts. Anesthesiology 1992;77:964-972.
- 120. Stevens WC, Cromwell TH, Halsey MJ, Bahlman SH. The cardiovascular effects of a new inhalation anesthetic, forane, in human volonteers at constant arterial carbon dioxide tension. Anesthesiology 1971;35:8-16.
- 121. Dolan WM, Stevens WC, Eger E, et al. The cardiovascular and respiratory effects of isoflurane-nitrous oxide anaesthesia. Can Anaesth Soc J 1974;21:557 -568.
- 122. Cromwell TH, Wendell C, Stevens WC, et al. The cardiovascular effects of compound 469 (Forane) during spontaneous ventilation and C02 challenge in man. Anesthesiology 1971;35:17-25.
- 123. Giesecke K, Hamberger B, Järnberg PO. Paravertebral block during cholecystectomy: effects on circulatory and hormonal responses. Br J Anaesth 1988;61:652-656.
- 124. Skovsted P, Sapthavichaikul S. The effects of isoflurane on arterial pressure, pulse rate, autonomic nervous activity, and barostatic reflexes. Can Anaesth Soc ] 1977;24(3):304- 314.
- 125. Seagard JL, Hopp FA, Bosnjak ZJ, Osborn JL, Kampine JP. Sympathetic efferent nerve activity in conscious and isofluraneanesthetized dogs. Anesthesiology 1984;61:266-270.
- 126. Millar RA, Biscoe TJ. Postganglionic sympathetic discharge and the effect of inhalation anaesthetics. Br J Anaesth 1966;38:92-114.
- 127. Millar RA, Warden JC, Cooperman LH, Price HL. Central sympathetic discharge and mean arterial pressure during halothane anaesthesia. Br J Anaesth 1969;41:918-928.
- 128. Skovsted P, Price HL. Sympathetic nervous system depression by halothane during prolonged exposure. Acta Anaesthesiol Scand 1972;16:65-68.
- 129. Smith NT, Corbascio AN. The cardiovascular effects of nitrous oxide during halothane anesthesia in the dog. Anesthesiology 1966;27:560-566.
- 130. Smith NT, Eger EI, Stoelting RK, Whayne TF, Cullen D, Kadis LB. The cardiovascular and sympathomimetic responses to the addition of nitrous oxide to halothane in man. Anesthesiology 1970;32:410-421.
- 131. Bahlman SH, Eger EI, Ty Smith N, et al. The cardiovascular effects of nitrous oxidehalothane anesthesia in man. Anesthesiology 1971;35:274-285.
- 132. Eisele JH, Smith NT. Cardiovascular effects of 40% nitrous oxide in man. Anesth Analg 1972;51:956-962.
- 133. Fukunaga AF, Epstein RM. Sympathetic excitation during nitrous oxide-halothane anesthesia in the cat. Anesthesiology 1973;39:23-36.
- 134. Ebert TJ, Kampine JP. Nitrous oxide augments sympathetic outflow: Direct evidence from human peroneal nerve recordings. Anesth Analg 1989;69:444-449.
- 135. Ebert TJ. Differential effects of nitrous oxide on baroreflex control of heart rate and peripheral sympathetic nerve activity in humans. Anesthesiology 1990;72:16-22.
- 136. Lundy JS. Balanced anesthesia. Minn Med 1926;9:399.
- 137. Tammisto T, Takki S, Toikka P. A comparison of the circulatory effects in man of the analgesics fentanyl, pentazocine and pethidine. Br J Anaesth 1970;42:317-324.
- 138. Derbyshire DR, Smith G. Sympathoadrenal responses to anaesthesia and surgery. Br J Anaesth 1984;56:725-739.
- 139. Grände P-O, Gustafsson D, Lindberg L. Effects of thiopental on resistance vessels in cat skeletal muscle. Intensive Care Med 1990;16:399-404.
- 140. Carlsson D, Harp JR, Siesjö BK. Metabolic changes in the cerebral cortex of the rat induced by intravenous pentothalsodium. Acta Anaesthiol Scand 1975;57:7-17.
- 141. Piatt JH, Schiff SJ. High dose barbiturate therapy in neurosurgery and intensive care. Neurosurgery 1984;15:427-444.
- 142. Derbyshire DR, Chmielewski A, Fell D, Vater M, Achola K, Smith G. Plasma catecholamine responses to tracheal intubation. Br J Anaesth 1983;55:855-860.
- 143. Zaloga GP. Catecholamines in anesthetic and surgical stress. In: International Anesthesiology Clinics. Little, Brown and Company, 1988: 187-198. vol 26).
- 144. Low JM, Harvey JT, Prys-Roberts C, Dagnino J. Studies of anaesthesia in relation to hypertension. VII: Adrenergic responses to laryngoscopy. Br J Anaesth 1986;58:471-477.
- 145. Wallin BG. Muscle sympathetic activity and plasma concentrations of noradrenaline. Acta Physiol Scand 1984;527 suppl:21 -24.
- 146. Cullen PM, Turle M, Prys-Roberts C, Way WL, Dye J. Effect of propofol anesthesia on baroreflex activity in humans. Anesth Analg 1987;66:1115-1120.
- 147. Samain E, Marty J, Gauzit R, et al. Effects of propofol on baroreflex control of heart rate and on plasma noradrenaline levels. Eur J Anaesthesiol 1989;6(5):321-326.
- 148. Blake DW, Jover B, McGrath BP. Haemodynamic and heart rate reflex responses to propofol in the rabbit. Br J Anaesth 1988;61:194-199.
- 149. Bristow JD, Prys-Roberts C, Fisher A, Pickering TG, Sleight P. Effects of anesthesia on baroreflex control of heart rate in man. Anesthesiology 1969;31:422- 428.

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V

### 9, Original **papers**

Anesthetic modulation of the cardiovascular response to microlaryngoscopy.

A comparison of propofol and methohexital with special reference to leg blood flow, catecholamines and recovery.

The effects of propofol, methohexitone and isoflurane on the baroreceptor reflex in the cat.

Characteristics of muscle nerve sympathetic activity during general anaesthesia in humans.

Percutaneous recording of muscle nerve sympathetic activity during propofol, nitrous oxide and isoflurane anesthesia in humans.

Sympathetic muscle nerve activity, peripheral blood flows and baroreceptor reflexes in humans during propofol anesthesia and surgery. På grund av upphovsrättsliga skäl kan vissa ingående delarbeten ej publiceras här. För en fullständig lista av ingående delarbeten, se avhandlingens början.

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