Thymic Studies Investigations into the effects of childhood thymectomy, and characterization of thymic B cells and Hassall's corpuscles

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Curiosity killed the cat

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ABSTRACT

This thesis focuses on the human thymus, a primary lymphoid organ responsible for the maturation of T cells. Progenitors arrive from the bone marrow and start to randomly assemble their T cell receptor (TCR) followed by a thorough selection process in which the TCR is tested for functionality and autoreactivity. The selection process is carried out with the help of different types of antigen presenting cells to ensure that only functional mature T cells that do not react towards the body's own structures are released into the periphery. In the selection process, also T regulatory cells that can maintain tolerance by acting immunosuppressive are generated from subset of the autoreactive T cells. Only around 3% of the progenitors that enter the thymus leave as mature T cells two-three weeks later and the net output is approximated to 1.7 $x10^7$ cells/day. The thymus is most active during childhood. Starting at puberty the thymus gradually involutes, but even though only a fraction of its original capacity eventually remains it is functional throughout life.

In paper I we investigated the effect of early thymectomy on the diversity of the TCR in the peripheral T cell pool. We followed up on thymectomized children 18 years after thymectomy by analyzing peripheral blood samples. In these children, more than 90% of the thymus had been removed during heart surgery before the age of six months. T and B cells were sorted out from peripheral blood, DNA encoding TCR was sequenced, and the results were compared with age and gender matched controls. Thymectomized children showed reduced diversity of the T cell receptor repertoire in the periphery compared with controls, which may lead to reduced infection control and

blunted regulatory functions of the T cell pool. The B cell receptor diversity was unaffected.

Paper II focuses on thymic B cells, a small population that while consisting of less than 1% of the total cell count in the thymus, covers a relatively large area of the medulla. We discovered that a significant fraction of these B cells underwent immunoglobulin class switching, a process that usually takes place in germinal centers after the body encounters an infection, which should be a rare event in a newborn infant. The thymic B cells displayed a mature phenotype and expressed high levels of co-receptors for T cell communication along with the transcription factor AIRE, which would imply a role as an antigen presenting cell (APC) that may aid in the T cell selection process.

Paper III aims to characterize a prominent structure in the human thymic medulla, the Hassall's corpuscles. Since the medullary epithelial cells (mTEC) in and surrounding the structure are difficult to digest into a single cell suspension, they were cut out using laser microdissection for further studies. Analyses of the retrieved sections using RNA sequencing and proteomics showed an increasing similarity with skin epidermis the more differentiated and closer to the Hassall core the cells were located. The center, devoid of nuclei, also contained bacterial defense proteins, further emphasizing similarity to the skin. The mTEC differentiation is thought to be influenced by the expression of the *AIRE* gene. Comparisons between Down syndrome thymus (three copies of *AIRE*) and control thymus showed larger corpuscles in the former, perhaps due to a higher turn-over and differentiation of mTECs than in control tissue. In mouse models in which the *Aire* gene is knocked out, the corpuscle like structures in the thymus were fewer and smaller, and the skin was thinner.

Keywords: thymus, thymectomy, TCR, B cells, APC, Hassall's corpuscles, AIRE

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SAMMANFATTNING PÅ SVENSKA

Avhandlingens titel är Thymusstudier, undersökningar av effekten av thymektomi i barndomen, och karakterisering av B-celler och Hassallska korpuskler i thymus. Den beskriver funktioner hos human thymus, vad som händer med immunsystemet om thymus tas bort och beskriver olika cellers funktion och utveckling i thymus.

Thymus är ett viktigt organ i immunsystemet. Dit färdas stamceller från benmärgen för att utvecklas till mogna T-celler, en sorts vita blodkroppar som reglerar många immunsvar. Organet är som störst och mest aktivt under barnåren och börjar tillbakabildas och ersättas av bind- och fettväv under puberteten. Man behåller en viss produktion av T-celler livet ut.

De blivande T-cellerna måste utbildas i thymus för att kunna fungera i den genetiskt unika individen och för att hindra att de angriper kroppens egna vävnader. De har en T-cells receptor vars struktur slumpas fram genom olika kombinationer av gener och som används för att känna igen proteiner. I teorin skulle det kunna finnas 10^{20} möjliga kombinationer, och detta leder till att varje T-cell har en unik receptor. Bland dessa kloner, som de också kallas, finns en andel som skulle kunna känna igen och attackera våra egna vävnader och ge upphov till autoimmuna sjukdomar. I thymus finns en speciell celltyp, thymusepitelceller, som med hjälp av en transkriptionsfaktor, AIRE, kan uttrycka olika protein från hela kroppen. T-celler som binder in för starkt till dessa elimineras i thymus. Genom detta system tillåts inte celler som är potentiellt autoimmuna lämna thymus, vilket annars hade riskerat autoimmunitet ute i kroppens vävnader. Thymus alstrar även T-regulatoriska celler vilka dämpar immunförsvaret och motverkar felaktig aktivering av immunsystemet i periferin.

Avhandlingens första arbete undersöker effekterna av thymektomi i tidig ålder. Thymus är proportionellt mycket stort hos små barn och under hjärtkirurgi tas hela eller delar av organet bort, vilket är nödvändigt för att kunna komma åt hjärtat. I Sverige genomförs det drygt 200 hjärtoperationer varje år där thymus tas bort. Vi analyserade förekomsten av olika kloner av T-celler i blodprover hos thymektomerade barn 18 år efter operationen, och dessa jämfördes med kontroller som ej genomgått thymektomi. Resultaten visar en minskning av antalet T-celler med unika receptorer hos de som genomgått thymektomi. Detta skulle kunna ge problem senare i livet genom en bristfällig respons mot olika patogener eller oönskad respons mot kroppsegna strukturer.

Det andra arbetet karakteriserar de B-celler som återfinns i thymus i relativt lågt antal. De utvecklas ur samma stamceller i benmärgen som T-celler men stannar i benmärgen under den första mognadsfasen och är inte beroende av thymus för sin fortsatta utveckling. Vi upptäckte att en betydande del av Bceller i thymus hos nyfödda barn hade en mogen fenotyp som annars inte förekommer innan kroppen genomgått upprepade infektioner, något som spädbarn normalt sett inte haft. Dessa celler hade även högre nivåer av receptorer som används för att kommunicera med T-celler, vilket gör att vi tror att deras funktion i thymus är att hjälpa epitelceller att utbilda T-celler.

Tredje arbetet undersöker en struktur i human thymus som består av thymusepitelceller som heter Hassallska korpuskler. Förekomsten av dessa har varit känd en lång tid, men det är ännu okänt vilken deras funktion är. För att kartlägga dessa strukturer grundligt skar vi ut dem med ett mikroskop i kombination med en UV-laser och proverna analyserades avseende genuttryck och proteininnehåll. Resultaten visade på en keratinisering av korpusklerna som liknar den som pågår i hudens yttersta lager. Detta bekräftades även av studier med mikroskop. Jämförelser gjordes mellan thymus från barn med Downs syndrom och kontroller eftersom personer med Downs syndrom har en extra kopia av genen *AIRE*. *AIRE* tros driva utveckling av epitelceller mot hudlika strukturer. Thymus från barn med Downs syndrom har mycket större Hassallska korpuskler än kontroller. Vi studerade även möss med genen *Aire* borttagen, och dessa uppvisade mindre Hassallska korpuskler.

LIST OF PAPERS

This thesis is based on the following studies, referred to in the text by their Roman numerals.

- I. Gudmundsdottir J*, **Lundqvist C***, Ijspeert H, van der Slik E, Óskarsdóttir S, Lindgren S, Lundberg V, Berglund M, Lingman-Framme J, Telemo E, van der Burg M, Ekwall O**.** T-cell receptor sequencing reveals decreased diversity 18 years after early thymectomy*. J Allergy Clin Immunol. 2017 Dec;140(6):1743-1746.e7. doi: 10.1016/j.jaci.2017.08.002. Epub 2017 Sep 1.* * These authors contributed equally to this work.
- II. **Lundqvist C***, Camponeschi A*, Visentini M, Telemo E, Ekwall O[‡], Mårtensson IL[‡]. Switched CD21-/low B cells with an antigen-presenting phenotype in the infant thymus. *J Allergy Clin Immunol. 2018 Nov 30. pii: S0091- 6749(18)31721-4. doi: 10.1016/j.jaci.2018.11.019.* * These authors contributed equally to this work. ‡ These authors contributed equally to this work.
- III. **Lundqvist C**, Lindgren S, Cheuk S, Lundberg V, Berglund M, Thörn K, Telemo E, Ekwall O. Characterization of Hassall's corpuscles in the human thymus. *Manuscript*

PUBLICATIONS NOT INCLUDED IN THE THESIS

Rentzos G, Lundberg V, **Lundqvist C**, Rodrigues R, van Odijk J, Lundell AC, Pullerits T, Telemo E. Use of a basophil activation test as a complementary diagnostic tool in the diagnosis of severe peanut allergy in adults. *Clinical and translational allergy*. 2015;5:22.

Lundberg V, Berglund M, Skogberg G, Lindgren S, **Lundqvist C**, Gudmundsdottir J, Thörn K, Telemo E, Ekwall O. Thymic exosomes promote the final maturation of thymocytes. *Scientific reports*. 2016;6:36479.

Lundell AC, Nordstrom I, Andersson K, **Lundqvist C**, Telemo E, Nava S, Kaipe H, Rudin A. IFN type I and II induce BAFF secretion from human decidual stromal cells. *Scientific reports*. 2017;7:39904.

Raposo B, Merky P, **Lundqvist C**, Yamada H, Urbonaviciute V, Niaudet C, Viljanen J, Kihlberg J, Kyewski B, Ekwall O, Holmdahl R, Bäcklund J. T cells specific for post-translational modifications escape intrathymic tolerance induction. *Nat Commun*. 2018 Jan 24;9(1):353

Statello L, Maugeri M, Garre E, Nawaz M, Wahlgren J, Papadimitriou A, **Lundqvist C**, Lindfors L, Collén A, Sunnerhagen P, Ragusa M, Purello M, Di Pietro C, Tigue N, Valadi H. Identification of RNA-binding proteins in exosomes capable of interacting with different types of RNA: RBP-facilitated transport of RNAs into exosomes. *PLoS One*. 2018 Apr 24;13(4)e0195969

Lloyd KA, Wigerblad G, Sahlström P, Garimella MG, Chemin K, Steen J, Titcombe PJ, Marklein B, Zhou D, Stålesen R, Ossipova E, **Lundqvist C**, Ekwall O, Rönnelid J, Mueller DL, Karlsson MCI, Kaplan MJ, Skriner K, Klareskog L, Wermeling F, Malmström V, Grönwall C. Differential ACPA Bindning to Nuclear Antigens Reveals a PAD-Independent Pathway and a Distinct Subset of Acetylation Cross-Reactive Autoantibodies in Rheumatoid Arthritis. *Front Immunol*. 2019 Jan 4;9:3033.

Eriksson D, Bacchetta R, Gunnarsson H I, Chan A, Barzaghi F, Ehl S, Hallgren Å, van Gool F, Sardh F, **Lundqvist C**, Laakso SM, Rönnblom A, Ekwall O, Mäkitie O, Bensing S, Husebye ES, Anderson M, Kämpe O and Landegren N. The autoimmune targets in IPEX are dominated by gut epithelial proteins (2019) *J Allergy Clin Immunol*, in press (JACI-D-18- 01617R2, accepted Feb 27, 2019)

CONTENT

ABBREVIATIONS

- TBE Tick-borne encephalitis
- TCR T cell receptor
- TRA Tissue restricted antigen
- TREC T cell receptor excision circle
- Tx Thymectomy

1 INTRODUCTION

The body needs to balance the need of having a well-functioning immune response to pathogens against not reacting with self-structures causing autoimmunity. Part of this balance is exacted in the thymus, a primary lymphoid organ situated on top of the heart in the thoracic cavity (Figure 1). Here the developing T cells form a functioning adaptive immune system that does not react to self. The works included in this thesis are focused mainly on the human thymus.

Figure 1. Thymus in a child, located on top of the heart in the thoracic cavity. Anatomy of the Human Body, 20th ed. Gray, Henry. 1918.

1.1 THYMUS IN THE PAST

The earliest mention of the thymus gland in medical literature is from the first century AD by Rufus of Ephesus in Greece who described the thymus anatomically as a gland located over the heart. (Rufus Med. *De corporis humani appellationibus* $168.1 - 169.1$ ². An interesting theory about the origin of the word thymus has been put forward by Konstantinos Laois. Thymus might originate from Indo-European with the meaning of "vapor" or "fume". Since the involution of the organ was difficult to investigate at that time the disappearance of the organ could have been linked to vapor, or going up in smoke². Thymus has also been attributed to a Greek word for heart or soul. The interpretation being that the proportionally big thymus seated above the heart in young animals must be the base of the soul^{3, 4}.

In the beginning of the $20th$ century a large thymus was seen as a condition of sickness in young children. The organ was thought to put pressure on the lungs and impede breathing, treatment with irradiation was sometimes recommended. This belief might have risen due to the many autopsies performed on children diseased from serious illnesses such as diphtheria. The shrunken thymus seen in these children might have become the norm⁴.

In "The Anatomy of the Thymus Gland" from 1832 a detailed description of the human thymus is recorded. The author, Sir Astley Cooper, dissected and uncovered that the two thymic lobes are divided into smaller lobes that can be unraveled in a serpentine manner, comparing the organ to a necklace of beads (Figure 2).

Figure 2. 1. The serpentine form of the lobes. 2. The lobes partially unraveled. From "The Anatomy of the Thymus Gland" by Sir Astley Cooper. 1832.

Veins, arteries and mucous membranes needed to be removed for the thymus to unravel in this fashion. The different lobules were connected allowing communication between them with a spiral cavity in the center of the gland. He demonstrated the connection between the lobes by injecting mercury into one lobe and followed the diffusion into the adjacent lobe. He also described the thick fluid coming out from the organ as filled with particles, and described

it as the same particles found in blood⁵. These particles, or blood lymphocytes, and the function of the thymus was not generally accepted until 1960s and were long considered without a role in immunity until Jacques Miller showed dramatic effects on the immune system in mice thymectomized at birth 6 .

Before the role of the thymus was revealed, it became famous in Swedish media. In 1952 a Swedish newspaper published a story about veterinarian Elias Sandberg and how he had discovered a new medicine for cancer. He defended his thesis about the calf thymus a decade earlier and believed that the key to immunological resistance laid in the thymus. He had started to treat people suffering from terminal cancer with injections of THX, a calf thymus extract, which became national news. This was the start of a prolonged conflict between Sandberg, medical doctors and the state, which lasted until his death in 1989⁷. Until 2009 there was still a registered alternative medicine, Enzythym, based on Elias Sandberg's theories8.

Figure 3. Human thymus section stained with Hoechst to show the nuclei. Cortex (C) is the dense area and the medulla (M) is the sparse area. Scale bar 200µm.

1.2 THYMUS TODAY

Huge progress has been made in the field of immunology and thymus research since its function was first described by Miller⁶.

The lobules of the human thymus consist of two distinct areas; medulla and cortex. The cortex consists mainly of immature thymocytes, heavily branched cortical epithelial cells (cTECs) and macrophages with the main function to clear apoptotic thymocytes. The medulla is much sparser and mainly consists

of single positive thymocytes, medullary epithelial cells (mTEC), macrophages, dendritic cells (DC), and B cells.

Other cell types have also been reported to inhabit the thymus, such as neutrophils⁹, eosinophils^{10, 11} and mast cells¹². One of the most unexpected cells found in the thymic medulla was the myoid cell, containing myofibrils¹³, and from these cells a cell line was established that expressed a functional acetyl choline receptor¹⁴. The latest cell type to be uncovered in the human thymus was the tuft cell, usually seen in the gastrointestinal tract^{15, 16}.

1.2.1 THYMOCYTE DEVELOPMENT

The T cell progenitors from the bone marrow enter the thymus in the corticomedullary junction. The capillaries extending into the cortex are impermeable, but venules in the corticomedullary junction are fenestrated, allowing progenitors to enter the thymus. The so-called blood thymus barrier prevents antigens from reaching the developing thymocytes in the cortex^{17, 18}, but is incomplete in the medulla, allowing antigens through from the blood.

When the thymocytes enter the cortex, they are double negative (DN), expressing neither of the T cell markers CD4 or CD8. At the third double negative stage the thymocytes begin to re-arrange their T cell receptor (TCR), starting with the β -chain, and if successful they receive signaling through their pre-TCR. The pre-TCR consist of the rearranged β -chain and a pre-alpha chain. The thymocyte then rearranges the α -chain until it results in a productive $\alpha\beta$ -TCR. The theoretical TCR diversity has been calculated up to 10²⁰ possible clones¹⁹. At this stage the thymocytes have a short lifespan and are destined to apoptosis, and if they are not rescued by a survival signal from binding to MHC molecules on cTECs they die by neglect²⁰⁻²². cTECS have constitutive autophagy degrading their intracellular proteins to be presented on both MHC class I and II to the developing thymocytes²³. There is also growing evidence for a negative selection process in the cortex, which seems to be dependent on presentation of self-antigens by dendritic cells²⁰.

The surviving thymocytes migrate into the medulla as single positive, either for CD8 or CD4 depending on if the survival signal came from binding MHC class I or II. In the medulla, self-antigens are presented to the thymocytes by mTECs or DCs which results in one of three main outcomes depending on the affinity for the antigens presented; negative selection (by activation induced apoptosis), diversion into the regulatory T cell lineage or egress from the thymus as an effector T cell. The mTECs express a vast number of tissue restricted antigens (TRAs) under the influence of AIRE, and a high constitutive

autophagy activity for the generation of numerous self-peptides. When these are presented to the thymocytes, autoreactive clones will effectively be removed or be directed into the regulatory T cell lineage²⁴⁻²⁶. The TRAs produced by the mTECs have also been shown to be transferred to DCs to enlist them in the negative selection process²⁷. This transfer has been suggested to be partly mediated via exosomes carrying MHC-peptide complexes emanating from the mTECs²⁸. Eventually, approximately 3% of the thymocytes exit the thymus as mature T cells²⁹.

The importance of the generation of a regulatory T cell population expressing FoxP3 for preventing autoimmunity is illustrated by the disease immune dysregulation, polyendocrinopathy, enteropathy, X-linked (IPEX) caused by mutations in the *FOXP3* gene. It is a rare, severe, autoimmune disease with bowel and skin inflammation, autoimmune diabetes and other autoimmune manifestations presenting already in the neonatal period 30 . It was recently shown that regulatory T cells can arise from two different development programs, where one path develops through agonist selection similar to negative selection with high affinity to self and the other path shows more similarities with positive selection and display a broader repertoire³¹.

B cells have also been suggested to be of importance for the development of regulatory T cells, having MHC class II, and costimulatory molecules such as CD80, CD86 and CD40. A mouse strain lacking B cells shows no difference in $CD4^+$ and $CD8^+$ thymocytes but has lower numbers of regulatory T cells in the thymus 32 .

1.2.2 THYMIC INVOLUTION

The thymus grows in size until puberty when the involution starts, this process continues throughout life and if extrapolated it has been estimated that the thymus would be completely absent at 120 years of age³³.

Signs of involution, such as widening of trabeculae and of the perivascular space, has been attributed as early as after the first year of life³⁴. The impact of puberty on thymic involution has been debated³⁵, and peak cellularity has been proposed to occur as early as at 6 months of age³⁶. In an effort to better quantify involution and thymus senescence a labeling technique with a modified form of Sudan black (binding lipofuscin) was developed by Barbouti and coworkers. They demonstrated that infant and young thymi showed no cellular senescence but during adolescence senescence seems to be activated 3^7 . Involution does not seem to be due to intrinsic aging of the lymphohematopoietic stem cells and early T cell progenitors, but rather

changes in the thymic environment³⁸. For example, FoxN1, which is of vital importance for mTEC development and function, is shown to gradually decrease with age in mTECs³⁹.

2 PAPER I: THYMECTOMY

2.1 THYMECTOMY

Thymectomy (Tx) for a non-medical reason is performed on children undergoing cardiac surgery to correct congenital heart defects. The thymus blocks the surgeon's access to the heart and is removed routinely. This type of surgeries started to become more common after 1970 when surgical techniques, as the cardiopulmonary bypass, allowed more lifesaving interventions⁴⁰. Heart defects affect approximately 1 % of all children of which 1/4 to 1/3 undergo open surgery including thymectomy. Roughly 200 Txs are performed each year in Sweden (Figure 4). Individuals that have undergone Tx are increasing in number and age, which makes it important to study the immunological and clinical consequences of thymectomy thoroughly.

Figure 4. Thymectomies performed in Sweden over time. Adapted from Gudmundsdottir et al1.

2.1.1 THYMIC OUTPUT

2.1.2 TRACING THYMIC OUTPUT

When T cell progenitors enter the cortex, they start to rearrange the T cell receptor (TCR), beginning with the b-chain during the DN3 stage. After successful rearrangement of the β -chain the thymocyte undergoes proliferation and progresses into the DN4 stage, and the TCR α -chain rearranges²¹. TCR α chain can make multiple rearrangements, until the recombination is halted by positive selection, or the cell dies^{41}. Thymic nurse cells are believed to help in the multiple rearrangements of the α -chain^{20, 42}.

In the rearrangement process of the TCR genes, TCR rearrangement excision circles (TREC) are generated. The most commonly measured variant is the signal joint TREC (sjTREC), circular DNA strands created during recombination of the α -chain⁴³. The rings are stable and not duplicated in mitosis, and are therefore diluted when the cells expand in the periphery to reconstitute the T cell pool. Recent thymic emigrants (RTE) have a higher level of TRECs than memory T cells, due to that less cell divisions have occurred in RTEs. A drop of 1-1.5 log_{10} is expected during a lifetime. TRECs are still detectable in elderly people, while no TREC can be measured in patients with complete Di George syndrome, that lack a thymus^{44, 43}.

TRECs represent a useful way to quantify thymic output, however, it can be misleading since naïve T cells are long-lived and TRECs can remain in nondividing cells the whole lifetime. Thus, a TREC containing naïve T cell is not necessarily recently produced by the thymus. Adult thymectomy, when the individual has an established repertoire, does not lead to a rapid decline in TREC levels 43 .

2.1.3 THYMIC OUTPUT WITH AGE AND THYMECTOMY

Thymic involution is a process were the active lymphoid tissue is replaced by fat and connective tissue. This process takes place slowly over a long period of time, with an increase at puberty and periods of fast involution with following rebound, such as after pregnancy and corticosteroid treatment $45-48$. The pregnancy studies were mainly performed in mice although the same pattern should be expected in humans. A newly released study in humans showed no difference in TREC levels in naïve T cells during pregnancy compared to non-pregnant controls, arguing that the thymic output is maintained in humans. However, due to the longevity of naïve cells and the limited time of a pregnancy, it is difficult to draw any firm conclusions 49 . An

argument against a long-lasting impact of sex hormones on the thymus is that the observed castration-induced involution in mice is short lived⁵⁰.

Involution normally starts 10-15 years later than a childhood Tx and proceed at a slow pace, with TRECs still detectable up in high ages since adult thymus contains areas of active tissue⁵¹. Even though the decrease in TREC levels between 25 and 60 years of age has been shown to be more than 95%, the TCR diversity at 60-65 years did not differ too much from young adults, with a clone diversity comprising 20 million different ß-chains. After 70 years of age the repertoire diversity decreased drastically to a clone diversity of $200,000^{52}$. An aging immune system, with involution of the thymus, correlates with an increase of infections and autoimmune diseases, and is referred to as $immunosenescence⁵³$.

Thymectomy at a young age would be expected to affect the peripheral T cell pool in a similar but accelerated way as seen in the process of aging. Disruption of the T cell compartment after thymectomy was shown already in $1970s^{54}$. Although some studies have shown no apparent effects on the immune system $55-57$, the majority of studies performed have found that early thymectomy leads to an impairment of the T cell compartment. Lower T cells numbers, lower TRECs and fewer RTEs have also been reported⁵⁸⁻⁶⁰ together with alterations in the CD4 and CD8 ratio^{58, 61}. Reduction of naïve T cells in thymectomized individuals together with an increase in Ki67 indicate that an expansion of T cells in the periphery compensate for absent thymic output⁶². A recent study shows lower CD4 and CD8 naïve cell counts, but a preserved regulatory T cell compartment, in Tx individuals 63 . Earlier the same group suggested that homeostatic proliferation of peripheral regulatory T cells explained their increased numbers⁶⁴. These observations regarding increased numbers of regulatory T cells after thymectomy is confirmed in a separate study in which an increase of T regulatory cells and their cytokine production was detected during the first years after thymectomy⁶⁵. Peripheral proliferation of T regulatory cells could potentially play an important role in limiting the amount of autoimmune diseases after Tx.

Thymectomized individuals have also been demonstrated to have increased frequencies of autoantibodies, for example autoantibodies associated with autoimmune liver disease and SLE $^{63, 66}$.

2.2 THYMECTOMY FOLLOW UP

2.2.1 THYMECTOMY FOLLOW UP STUDY

Paper I is part of a study that was started in 1993 by Solveig Oskarsdottir and Anders Fasth. Children under the age of 6 months that got more than 90% of their thymus removed during the cardiac surgery at the Queen Silvia Children´s Hospital in Gothenburg were included in the study. Blood samples were taken preoperatively, at 18 months and 18 years of age and compared to matched controls.

Childhood thymectomy resulted in immunological changes resembling premature aging. The thymectomy resulted in lower absolute numbers of naïve $CD4^+$ cells, $CD31^+$ cells and T regulatory cells, although the proportions were mainly unaffected. TREC levels among the thymectomized patients were low to non-detectable. The telomeres were shorter among $CD8⁺$ cells, indicating peripheral expansion. Signs of repertoire oligoclonality were discovered using flow cytometry analysis for TCR variable β -chain⁶⁷, which prompted us to follow up with immunorepertoire sequencing (**Paper I**), to enable a more detailed repertoire analysis.

DNA from sorted $CD4^+$, $CD8^+$ and $CD19^+$ cells was sequenced and analyzed for T cell receptor β chain (TCR β) and immunoglobulin heavy chain (IGH) usage. This allowed a more detailed investigation than possible with flow cytometry. It did not only give information about the genes used but also deletions, insertions and CDR3 length and composition.

The method used to quantify the clonality is based on the occurrence of coincidences⁶⁸. The sample was divided into six reactions that were amplified and sequenced individually. If the same clone appeared in more than one reaction it was termed a coincidence. Based on the coincidences it is possible to calculate a clonality score. Our main result from **Paper I** was the significantly increased clonality among CD4⁺ and CD8⁺ T cells in the thymectomized patients. As an internal control we could, as expected, not detect any difference in the clonality of $CD19⁺$ B cells between thymectomized individuals and controls. The clonality score among T cells were negatively correlated with the number of CD4+ and CD8+ T cells in peripheral blood, further strengthening the results.

2.2.2 THYMECTOMY LONG TERM EFFECTS

Responses to vaccines obtained previous to Tx, e.g. MMR seem relatively unaltered, with similar MMR-specific IgG concentration as controls. Responses to vaccinations after thymectomy, e.g. tick-borne encephalitis, was delayed, with a normal response first after the third vaccination⁶⁹. Age at Tx correlated with TBE-specific IgG antibody levels, with higher levels the later the Tx was performed, which is supported by the observation that thymectomized children show significantly lower total counts and percentages of naïve T cells, which correlated with the time passed since Tx, compared to controls⁷⁰. Hepatitis B vaccination in individuals with no thymic activity revealed undetectable or low levels of Hepatitis B-specific IgG^{7I} .

Ageing mice have an impaired immune response against influenza virus. Their aged immune system suffers from a restricted diversity of $CD8⁺$ T cells, resulting in holes in the repertoire, which hampers the immune response. The same effect was seen in thymectomized mice, consistent with the decreased repertoire, where absolute number of $CD8⁺$ T cells was unchanged, but a reduced response in influenza specific $CD8⁺$ T cells was observed. These results strengthen the arguments for links between decreased diversity, age and less responsiveness to infections⁷². In a larger register study, Gudmundsdottir et al reported an increased risk for autoimmune diseases such as hypothyroidism and type 1 diabetes and infections in thymectomized patients compared to surgery controls. The study included 5664 thymectomized individuals, but due to the relatively low average age of the patients (mean 14 years) the follow up time was still short. The amount of thymus tissue removed during surgery was not reported, but far from all subjects had undergone total thymectomy, which might lead to an underestimation of the differences between the compared groups¹.

Two studies that studied atopy in thymectomized patients reported different findings. In the first study, heart surgery was associated with increased frequencies of atopic disorders, possibly due to an altered T cell repertoire. They showed that thymectomy significantly increased the development or worsening of atopic symptoms, mainly asthma. The patients had undergone heart transplantation and were treated with immunosuppression, which may have affected the results⁷³. A second Danish study of risk for atopic dermatitis among thymectomized infants showed that the risk for atopic dermatitis was reduced in the surgery group compared to controls⁷⁴. This was also shown by Gudmundsdottir et al in the register study mentioned above¹ and may be explained by the decreased T cell efflux following thymectomy.

2.2.3 PERIPHERAL EXPANSION

Studies of the effects of human thymectomy generally show relatively mild clinical outcome. This supports the notion that homeostatic proliferation of naive T cells in the periphery is effective and can compensate for a decreased thymic output. This regulation is particularly active in lymphopenic hosts, such as elderly individuals and thymectomized patients⁷⁵.

Patients thymectomized during their first 30 days of life that were followed up showed lower TREC levels and higher levels of IL-7 in serum. The levels of IL-7 correlated negatively with absolute $CD4^+$ T cell counts two years postthymectomy61. Another article also reported significantly elevated levels of IL-7 the first years after thymectomy⁷⁶. Further findings supported the idea that peripheral expansion counteract the decrease in thymic output to maintain T cell homeostasis. The altered equilibrium has also been illustrated by higher levels of Ki67 in naïve T cells after thymectomy, which did not normalize until ten years post-thymectomy⁶².

Most centenarians have undetectable TRECs and lower levels of $CD4^+$ and $CD8⁺$ T cells than both young controls and middle-aged individuals. An important factor for the thymic T cell production and the maintenance and survival of the peripheral T cell pool is IL-7, and interestingly plasma levels of IL-7 were higher in women, which have been speculated to be a factor involved in the higher number of female centenarians⁷⁷. Furthermore, IL-7 given to aged macaques increased the thymic output measured by TRECs and resulted in an increase of central memory cells⁷⁸. Thus, the higher IL-7 among female centenarians is possibly resulting in a better conservation of the lymphocyte pool.

In mice the maintenance of the peripheral naïve T cell pool is sustained by thymic output throughout their lifetime, and almost all naïve T cells originate from thymic output in mice, even at old age. The T cells have a short life span of approximately 7 weeks for $CD4^+$ and 11 weeks for $CD8^{+29}$. In contrast, the human T cell pool is more dependent on peripheral T cell division⁷⁹, which makes comparisons between human and mouse less relevant and can probably account for the relatively mild clinical manifestations of childhood thymectomy observed in the clinical follow-ups so far.

A diverse repertoire can have an impact on health later in life. In a study on glioblastoma multiforme, where advanced age is a predictor for poor clinical outcome, a favorable prognosis correlated better with CD8+ RTE levels measures, as measured by TRECs, than with age⁸⁰. The age dependent decreased thymic output of $CD8⁺$ T cells could possibly influence the agerelated cancer mortality. An immune model was used to show the association with cancer and thymic involution rather than with age, although it normally accompanies each other. An interesting speculation was that the reduced cancer risk observed in certain shark species could be due to the thymus not involuting⁸¹.

Thymic output is thought to be vital during T cell repertoire establishment, but not essential for repertoire maintenance during adulthood, at least for a limited time. The relative diversity seen in thymectomized individuals and the proportions between naïve and memory T cells are often reported to be sustained during a long time. Due to that the peripheral expansion is so efficient in humans, it may take a long time before the full effects of thymectomy are shown as clinical manifestations. With an emerging group of thymectomized patients, and a population growing older, treating diseases of aging by targeting the thymus, the thymic output or the peripheral expansion represents interesting therapeutic possibilities.

3 PAPER II: THYMIC B CELLS

3.1 THYMIC B CELLS

B cells constitute about 1 % of the total cell number in both human and murine thymus^{82, 83, 84}. They were first discovered in the human thymus in 1987 by immunohistochemistry, which revealed the presence of these cells almost exclusively in the medulla 85 .

3.1.1 MOUSE THYMUS

The B cells in the mouse thymus have been reported to emanate from progenitor cells within the thymus, with the recruitment from the periphery playing only a minor part 86 . The progenitors are located in the cortex area while more mature B cells reside in the medulla⁸⁷. However, other studies have reported that peripheral immigration contribute substantially to the establishment and maintenance of the thymic B cell population 82 .

Thymic B cells are characterized by the expression of Aire, CD80, CD86 and high levels of MHC class II and CD40^{82, 86}. These specific features of the thymic B cells are acquired in the thymic environment, which was shown by Yamano et al by injecting IgM⁺IgD⁺MHCII^{int}CD80⁻Aire⁻ B cells and later finding them in the thymus with higher levels of MHCII and positive for CD80 and $Aire^{82}$.

Interestingly, even though the percentage of B cells in the thymus increases with age, the absolute number of B cells goes down. The expression of *Aire* and self-antigens appear to diminish with age, and if aged B cells are injected intra-thymically in young mice, this expression is not restored. These results suggest that the inability to express *Aire* and self-antigens due to aging is an intrinsic feature of the B cells⁸⁸.

The Ig switching of the thymic B cells in mice is thought to take place intrathymically, and is dependent on the B-T cell interaction where the CD40- CD40L interaction plays an important role. This interaction is also crucial for the maintenance and proliferation of the thymic B cells⁸⁹. The repertoire of the thymic B cells is distinct, with a high degree of autoreactivity, making the B cells capable of acting as effective APCs for self-antigens during T cell selection, which suggests an important role in shaping the $CD4^+$ T cell repertoire $83, 86$. Similarly to dendritic cells, the thymic B cells are reported to be able to aid in the negative but not the positive selection⁹⁰.

Thymic B cells have also been proposed to play a role both in the induction of T regulatory cells^{91, 92} and in the deletion of autoreactive thymocytes in an experimental murine system using myelin oligodendrocyte (MOG) reactive thymocytes and B cells expressing MOG on MHC-class $II^{93, 94}$.

A specific thymic B cell population in the mouse, expressing sialidase, was discovered in 2004⁹⁵. It has been proposed that these B cells, together with mTECs, remove sialic acid on thymocytes to aid interaction with APCs in the negative selection process. SP thymocytes have higher levels of sialic acid covering D-galactose residues. This can be shown by staining with peanut agglutinin (PNA), which binds the galactose residues in the DP thymocytes in the cortex whereas staining is impaired in the SP thymocytes with higher level of sialic acid. It has been proposed that in order to allow tight interactions between maturing thymocytes and APCs this sialic acid needs to be removed $96-$ 98.

We have seen a similar staining pattern as it has been described in mouse thymus when staining with PNA in human thymus tissue. (Figure 5)

Figure 5. PNA (green) and nuclear stain Hoechst (gray) staining of the same area, showing PNA staining in the immature thymocytes in the cortex. Scale bar 200µm.

A study in non-obese diabetic (NOD) mice showed an increased activity of the thymic B cells in the prediabetic phase. The thymic tissue showed an accumulation of thymic B cells in the cortico-medullary junction and formation of germinal centers. Autoantibodies binding cytokeratin 5^+ epithelial cells were found in the NOD mice together with a higher level of apoptosis among these cells. The antibodies, presumably produced by the accumulated B cells, could be inducing apoptosis in mTECs, including insulin expressing mTECs. This was thought to impair the thymic negative selection of insulin reactive T cells driving the development of diabetes in the NOD mice⁹⁹.

3.1.2 HUMAN

Less work has been done concerning human thymic B cells. Human thymic B cells are located in the medulla or in the perivascular spaces, similarly to the distribution in mice 100 . The B cells in the perivascular area are thought to be plasma cells, secreting antibodies towards viral proteins. These cells are maintained throughout aging and are assumed to protect the thymus from infections 101 .

The B cells located in the medulla are suggested to take part in the negative selection of thymocytes. Thymic B cells show a prominent reactivity towards peptide autoantigens¹⁰², and by cloning and expressing antibodies from thymic B cells they appear to be more autoreactive than B cells in the bone marrow. ¹⁰² It has also been shown that thymic B cells have a strong bias towards $V(H)4$, a gene segment family frequently encountered in autoimmunity 103 .

According to a recent study, about half of the thymic B cells in humans are naïve B cells 84 . As shown in this thesis and by others, the thymic B cells express AIRE and high levels of CD86, MHC class II and CD40^{84, 88}. Moreover, human thymic B cells express tissue restricted antigens (TRAs) that are different from those expressed by $mTECs⁸⁴$. Together with their location in the medulla and their activated phenotype, the expression of AIRE and TRAs in human thymic B cells supports their possible involvement in negative selection and generation of regulatory T cells.

As in mice, the percentage of B cells in the thymus has been reported to rise in older children³⁶, but the levels of AIRE declines with age⁸⁸. Autoimmunity has been linked to abnormal B cell numbers in the thymus and germinal center formation has for example been observed in $SLE¹⁰⁴$ and myasthenia gravis $(MG)^{105}$. In addition, CCL21 is overexpressed in MG thymus, attracting both T cells and naïve B cells from the periphery¹⁰⁶. Furthermore, an increase in T follicular helper cells has been reported in thymic tissue of MG patients. Similarly to T follicular helper cells in mice they might drive the B cell development by expressing $IL21^{107}$.

3.2 CD21^{-1 LOW} B CFLLS

Over the last decade a population of mature B cells with low expression of the complement receptor 2, $(CD21^{-/low})$, has been described in tonsils and peripheral blood of healthy individuals^{108, 109}. This B cell population has also been found to expand with age and is more abundant in patients with chronic infections and autoimmune diseases 110 .

In peripheral blood from healthy controls the CD21^{-/low} B cells are mainly memory cells, and account for approximately 5% of all B cells¹⁰⁸. The population is absent in cord blood, suggesting that they are antigenexperienced cells. In tonsils they have been defined by their expression of the Fc-receptor-like protein 4 (FcRL4) and lack of CD27, a key marker for memory B cells. They are mainly isotype-switched and defined as tissue-based memory B cells $^{109, 111}$.

CD21-/low B cells are expanded in conditions with chronic infection such as human immunodeficiency virus $(HIV)^{112}$, hepatitis C virus $(HCV)^{113}$ and malaria¹¹⁴, but also in immunological disorders and autoimmune conditions such as common variable immunodeficiency $(CVID)^{115}$, rheumatoid arthritis $(RA)^{116}$ and systemic lupus erythematosus $(SLE)^{117}$. The persistent immune activation observed in these disorders makes the CD21^{-/low} B cells hyporesponsive to stimulation via the B-cell antigen receptor $(BCR)^{110}$. The role of CD21^{-/low} B cells in health and disease is not fully understood.

3.3 CD21^{-/LOW} B CELLS IN THE THYMUS

In **Paper II** we show that half of the B cells residing in the human thymus early in life display a unique phenotype characterized by the lack of or low surface expression of CD21, $(CD21^{-/low})$.

The lack of, or low, surface expression of CD21 is typical for immature B cells, such as early transitional B cells in peripheral blood of infants¹¹⁸, although these cells also express CD10, separating them from mature cells. However, when comparing the thymic $CD21^{-/low}$ B cells with cells from the same infants' peripheral blood, we found that the vast majority of the B cells in the thymus were mature cells, being CD10⁻CD34⁻.

Despite the thymic $CD21^{-/low}$ cells being negative for the memory B-cell marker CD27, almost half of them were Ig class switched cells. This was unexpected considering that switched B cells were almost absent in the peripheral blood from the same infants. Ig class switching occurs for example after active immunization or an infection, and switched B cells are not generally found in peripheral blood from healthy neonates. The origin of the switched B cells in the thymus of newborns could potentially be maternal due to cell microchimerism, however we found that the thymic B cells originated from the child, disproving the hypothesis of a maternal origin. As in mice, class switching could be facilitated by cognate interaction with the thymocytes 83

where the interaction between CD40 on the B cells with CD40L on the T cells seems crucial.

The thymic $CD21^{-/low}$ B cells were large in size and expressed high levels of the typical activation markers CD69 and CD95. Their high levels of CD86, a costimulatory molecule highly expressed on professional APCs, together with the high levels of HLA-DR and CD40 and their localization in the medulla, suggest a role of the thymic B cells, and in particular the $CD21^{-/low}$, in T-cell selection. This is supported by studies in mice where switched B cells play an important role in driving T-cell tolerance⁸³.

Thymic $CD21^{-/low}$ B cells expressed significant levels of AIRE, which was higher than in the $CD21^+$ cells. In mice, thymic B cells express TRAs induced by AIRE, and are efficient $APCs^{86, 82}$. Also, human thymic B cells express TRAs, which differ from the TRAs expressed by mTECs, suggesting a nonredundant contribution of thymic B cells to central T-cell tolerance⁸⁴. Our results propose that the major contributor to this selection is the CD21^{-/low} B cells, since they have an activated phenotype and express high levels of AIRE.

As mentioned above, CD21^{-/low} B cells are found in both peripheral blood and tonsils from healthy individuals, and are expanded under conditions of chronic immune stimulation. In most of these conditions, the CD21^{-/low} B cells express high levels of activation markers, with subsets that co-express T-bet, CD11c, FcRL4 and/or CXCR3. A CD21^{-/low} population termed age associated B cells (ABCs), that express T-bet and/or CD11c, has also been described in both wild type and autoimmune prone mouse strains^{119, 120, 121, 122}. CD11c expression, which also is a hallmark of dendritic cells, potentiates the ability of ABCs to present antigen to T cells¹²³. A subset of the CD21^{-/low} B cells in the thymus expresses CD11c supporting their role as APC. The inhibitory receptor FcRL4 has been found to dampen BCR-signaling¹²⁴ which would be consistent with most thymic $CD21^+$ cells being FCRL4 and respond to BCR agonists whereas some CD21^{-/low} cells were FCRL4⁺ and showed a bi-modal response.

These findings, together with the findings that about half of thymic $CD21^{-/low}$ B cells were apoptosis prone and half were $Ki67⁺$ proliferating cells provide evidence that thymic B cells are heterogeneous and in a highly dynamic state. Thymic B cells communicate actively with the thymocytes, shown in our coculture experiments were CD21^{-/low} B cells were able to induce CD25 upregulation in T cells more effectively than in the $CD21⁺$ counterparts.

4 PAPER III: HASSALL'S CORPUSCLES

4.1 THYMIC EPITHELIAL CELLS

The epithelial cells in the thymus are presumed to originate from a common progenitor¹²⁵. The common progenitor passes through a stage with expression of both cTEC and mTEC markers before differentiating into their respective lineages^{126, 127}. However, there is also evidence for lineage committed progenitors, where the mTECs have been shown to originate from one progenitor clone, forming islets 128 .

The mTECs differentiate from mTEC^{low} immature cells (Aire- MHC II^{lo}CD80-) to mTEC^{high} (Aire⁺ MHC II^{hi} CD80⁺) and subsequently return to a state of $mTEC^{low}$ (Aire⁻ MHC II^{lo} CD80⁻). Both differentiation into the mTEC lineage and maturation from mTEC^{low} to mTEC^{high} requires activation of the NF- κ B signaling pathway by members of the TNF-family, e.g. RANKL, which is produced by single positive thymocytes¹²⁹. Deficiency in RANKL leads to impaired medulla formation due to its importance for mTEC development¹²⁹. In adult mouse the halftime of $Aire^+$ mTEC^{high} is about 2 weeks¹³⁰. A normal mTEC life cycle, including intact kinetics, seem to be needed for the organization of the medulla to be successful.

The transition to post-*Aire* mTEC^{low} is less studied than the conversion to $mTEC^{high 131}$. In the post-Aire state the cells start to express late-stage keratins and later form the Hassall's corpuscles¹³², in a process which may be supported by the expression of keratinocyte growth factor by single positive CD4 and $CD8$ thymocytes 133 .

An important transcription factor in thymus ontogeny and development is *FOXN1*134. A lack of function mutation in *FOXN1* in humans is related to loss of hair, athymia and deficiencies in the T cell compartment¹³⁵. The mutation was first described in mice and gives rise to a nude phenotype, lacking hair and a functional T cell system¹³⁶. *Foxn1* is not completely non-redundant for all epidermal differentiation in the skin as nude mice do not have an altered skin histopathology, apart from absence of hair 137 .

4.1.1 AIRF

Aire is an mTEC defining gene that allows promiscuous gene expression, a process that aims to mirror the complete repertoire of peripheral self-antigens within the thymic medulla^{24, 25}. mTECs can express more than 18 000 genes, approximately 85% of the coding genome. In the absence of *Aire,*

approximately 15 000 genes were still expressed in the epithelial cells suggesting that *Aire* is responsible for inducing 3-4 000 genes. Only a fraction of all TRAs are expressed in one cell at a given time point^{138, 139}. The mTECs expressing a specific gene tend to localize in clusters in the medulla, about 1- $3%$ of all mTECs express a particular TRA 140 .

Aire interacts with unmethylated histone-3, found on inactive chromatin¹⁴¹. It induces histone modifications at a low frequency resulting in the low number of specific TRAs expressed by each $mTEC¹⁴²$. A recently described transcription factor, *Fezf2*, has been reported to control a set of Aire independent TRAs in mTECs, even though the function of *Fezf2* is not completely understood¹⁴³.

A deficiency of *Aire* in mice has been shown to cause failure in the thymic tolerance induction among single positive thymocytes. The thymus shows a defective removal of autoreactive thymocytes specific for *Aire* dependent antigens, and also an impaired generation of regulatory T cells, leading to multiple autoimmune manifestations. This indicates that the peripheral tolerance mechanisms are not enough to prevent autoimmunity, without negative selection taking place in the thymus²⁶.

Lack of functional AIRE in humans causes autoimmune polyendocrine syndrome type 1 (APS1) or autoimmune polyendocrinopathy-candidiasisectodermal dystrophy (APECED)¹⁴⁴, a syndrome characterized by multiple organ-specific autoimmune diseases such as hypoparathyroidism, primary adrenal insufficiency (Addison's disease) and mucocutaneous candidiasis¹⁴⁵. Although candidiasis in itself is an infection and not an autoimmune disease, the chronic mucocutaneous candidiasis in APS1 patients is thought to emerge as a result of neutralizing autoantibodies against IL-17 and IL-22, which hampers the defense against fungal infections^{146}. Other manifestations such as vitiligo, enamel hypoplasia, pernicious anemia, autoimmune hepatitis and type 1 diabetes are also common^{147}.

Even though AIRE is needed for a well-functioning negative selection, an overexpression of AIRE does not seem to improve the exposure of TRAs and result in a better central tolerance induction. Individuals with Down syndrome, or trisomy 21, have three copies of the *AIRE* gene, and it has been reported that higher levels of AIRE and also of insulin, a common AIRE dependent TRA is expressed in their thymus^{148, 149}. However, this overexpression of AIRE does not seem to be of benefit since both autoimmunity and infections are overrepresented in Down syndrome¹⁵⁰. In Down syndrome, signs of accelerated mTEC maturation kinetics is seen, which may be a result of the high AIRE

levels. A signature of premature involution including a small thymus size with a high ratio of medullary versus cortical areas together with larger cystic involutions in the medulla is also seen¹⁴⁸. Low levels of TREC is also confirming an impaired thymic function¹⁵¹.

The effects of increased AIRE expression have also been investigated in mice by creating a model in which extra copy of the human AIRE gene was added. This led to an altered maturation of the mTECs and a failure in depletion of autoreactive thymocytes 152 .

4.1.2 DISEASES OF THE THYMIC EPITHELIUM

The most common acquired disease of the thymic epithelium is thymoma. It can originate from either cTEC or mTEC. There are associations between thymomas and autoimmune diseases, the most common being myasthenia gravis. The maturation of mTEC in the thymomas can be disturbed, with an impaired expression of AIRE and TRA which in turn can result in a dysfunction of the negative selection¹⁵³.

Atrophy of the thymus is a common denominator in infectious diseases, mainly because of the depletion of thymocytes. This is mainly due to an increased apoptosis of DP thymocytes in the cortex and can be seen for example in AIDS and rabies¹⁵⁴. It has been theorized that this can be due to a rise in circulating glucocorticoid levels during infections¹⁵⁵. Thymocyte depletion in rabies can be prevented by adrenalectomy, showing that the thymic involution is depending on glucocorticoids¹⁵⁶. The measles virus can attack the thymic epithelium itself, and can cause an arrest in cell growth and induce terminal differentiation¹⁵⁷

4.1.3 LATE STAGE DIFFERENTIATION

The role of *Aire* in the organization of the thymic medulla is not completely clear. After a transient activated stage with expression of *Aire*, the mTECs continue into a post-*Aire* stage. The loss of *Aire* is accompanied by a decrease of MHC class II and CD80 expression¹⁵⁸.

Aire affects the differentiation of the thymic epithelium in mouse and the absence of *Aire* causes morphological changes with reduced numbers of terminally differentiated mTECs expressing involucrin¹⁵⁹. Deficiency of *Aire* also results in altered transcriptional profile, disruption of the thymic corticomedullary organization and a reduction of the medullary compartment. This indicates a role for Aire in the differentiation and composition of thymic epithelium 160 .

By lineage tracing experiments it was shown that the Hassall's corpuscles constitute the final developmental stage for the $Aire⁺ mTECs$, and that all involucrin positive cells have at one point also been positive for Aire ^{158, 161}.

4.2 HASSALL'S CORPUSCLES

The origin of the Hassall's corpuscles has been the subject of considerable controversy. They were long considered to be degenerated endothelial cells from small vessels in the thymus. Later it was proposed that they originate from remnants of omnipotent embryonal epithelial cells. They have also been said to contain reticuloendothelial cells and striated myoid cells in the core. Furthermore, the proposed function has varied over time, from a site to destroy `forbidden clones´ that are dangerous to the organism and recycling of materials, to storage of antibodies and antigens¹⁶²⁻¹⁶⁵.

An interesting study was made were it was tested if Hassall's corpuscles reacted to antigens. Tetanus toxin was injected to guinea pigs and the morphology of the thymus was investigated, showing proportionally more epithelial cells and larger Hassall's¹⁶⁶. This was probably due to stress and corticosteroid induction of apoptosis within the thymocyte population and not an active response from the epithelial cells.

The cell layers immediately surrounding the corpuscles stain positive for various late-stage keratins such as cytokeratin 10 and involucrin, also found in the upper layers of the skin. Although the Hassall's corpuscles origin from epithelial cells and their keratinization resembling skin was noted already in 1979^{167} , a reason for the many theories surrounding the structures could be the difficulty to study them microscopically due to the tendency of unspecific binding of histological stains and antibodies. The slow progress of deciphering the function of the Hassall's corpuscles could also be due to the limited size and appearance of the corpuscles in mice.

It is likely that the Hassall´s corpuscles represent the last stage of the mTEC development, where the epithelial cells lose their nuclei and form the core of the Hassall's corpuscles. The function, if any, of the corpuscles is still unknown and the ideas have varied over the years. The two most recent theories are the induction of regulatory T cells by Hassall's production of thymic stromal lymphopoietin $(TSLP)^{168}$ and a role as recruiters of IL-23 producing neutrophils by CXCR5 to induce plasmacytoid DCs to produce $TNF\alpha^9$.

An interesting sex difference has been reported, with a higher average number of Hassall's corpuscles in male than female fetuses¹⁶⁹. This could be explained by the differences in sex hormones that female and male fetuses are exposed to during pregnancy¹⁷⁰. A sex difference in AIRE expression has also been noted with a higher AIRE expression in males, which in turn could drive the mTEC differentiation and result in a higher turnover and more Hassall's corpuscles in males 171 .

4.2.1 HASSALL'S CORPUSCLES AND SKIN

In the manuscript included as **Paper III**, we aimed at characterizing the Hassall's corpuscles in the human thymus more extensively than had been done previously. It has been proven difficult to degrade the keratinized cell compartment of the Hassall's corpuscles and create a single cell suspension to be able to sort the different cell populations by FACS¹⁷². To overcome this problem and to acquire cells from the rim of the corpuscles as well as material from within the Hassall's corpuscles we chose to use a laser microdissection technique.

The RNA sequencing revealed more upregulated genes(3285) in the cell layers surrounding the HC than downregulated genes (656), compared to Hassall's corpuscle free thymic medulla. Gene ontology analysis revealed an enrichment of skin differentiation processes and a downregulation of T and B cell markers in the cells surrounding the corpuscles. This could be due to the enrichment of mTECs around the Hassall's corpuscles, resulting in a relatively lower frequency of lymphocytes. An analysis of the chromosomal enrichment gives the highest hit on Chromosome 1q21, where the epidermal differentiation complex is located. This consists of several genes with important functions in the terminal differentiation of the epidermis, such as S100As, involucrin, loricrin and $SPRRs^{173}$.

The results from the proteomic analysis of the cells surrounding the corpuscles and the material within the Hassall´s corpuscle core show a downregulation of chromatin and RNA processing systems in the latter, as the mTECs lose their nuclei and end up in the amorphous core of the corpuscle. An enrichment of late-stage keratins and antibacterial proteins is seen in the core, mimicking the keratinization process observed in the skin. The observed similarities regarding RNA and protein content between human thymus and skin was not entirely surprising, since similarities have been described in mice where immunohistochemistry has shown similar staining patterns in thymus and \sin^{158} .

As in mouse, the human mTEC^{low} stage is followed by mTEC^{high}, in which the cells express AIRE and TRAs to be presented to the developing thymocytes in the negative selection process. After the loss of AIRE, the cells transition into mTEC^{low}, expressing late-stage keratins and eventually forms the Hassall's corpuscles.

As reported earlier, we demonstrated a higher level of AIRE expression in Down syndrome thymus, probably caused by the extra copy of *AIRE*148, 149. The higher level of AIRE could be driving an increased turnover of the epithelial cells, resulting in the enlarged Hassall's corpuscles typical for Down syndrome thymus. Unfortunately, we have not managed to acquire skin samples from Down syndrome patients to investigate if the differentiation of the epidermis is comparably affected. However, it is interesting in this context that Down syndrome patients have been reported to have increased frequencies of keratinization defects of the skin, resulting in diseases such as keratosis pilaris and xerosis 174 .

In Aire \pm mice we can report a decrease of involucrin positive areas, the mouse equivalent of Hassall's corpuscles, in the thymic medulla. Interestingly, the skin of Aire-/- mice showed a reduced thickness of the involucrin positive cell layer, suggesting an involvement of Aire in the development of both thymic epithelium and skin in mouse.

In theory, the Hassall's size could correspond to the thickness of the skin of the animal species in question. The human skin is thicker than for example the mouse skin, which corresponds to the bigger Hassall's corpuscles seen in humans. (see Figure 6). However, this theory remains unproven, as articles describing the thymus histology from animals with thick skin, such as Nile crocodiles and rhinoceros, do not mention the particular size of the Hassall's corpuscles¹⁷⁵ ¹⁷⁶.

One potential function of the Hassall's corpuscles could simply be to provide skin specific antigens for T cell education. Interestingly, it has been shown that Hassall's corpuscles for example express the pemphigus vulgaris-related autoantigens Desmoglein-1 and -3158.

Figure 6. Thymus staining with Involucrin (green, upper row) and skin (lower row) from human (left) and mouse (right). nuclear stain (gray) is Hoechst. Scale bar 50µm.

As the end stage mTECs around the Hassall´s cannot desquamate after being keratinized, in the same way as the keratinized cells of the skin do, they create these large formations of keratin, which probably are challenging for the thymus to break down. A question, yet to be addressed, is how the corpuscles are degraded in the thymus, since there has been no report of them accumulating with age to the extent expected if not being possible to degrade.

5 METHODS AND LIMITATIONS

For a detailed description of the methods used in the thesis I refer to the original papers.

5.1.1 TISSUE HANDLING

The thymic tissue was placed in cold PBS immediately after surgery and until further processing. Blood from the patient was collected in a heparin tube and kept at room temperature.

For analysis of lymphocytes, the tissue was cut finely and gently pressed over a 40µm cell strainer. The cells were passed through a cell strainer and the blood sample was centrifuged on a density gradient with Ficoll-Paque (GE Healthcare Life Sciences, Little Chalfont, Buckinghamshire, United Kingdom) to separate the lymphocytes and "clean" the sample.

For analysis of the epithelial cell compartment, mechanical and enzymatic digestion was needed. The tissue was cut into small pieces and incubated on rotation with RPMI media with Liberase TM (Roche) and DNase (Worthington) with intermittent mechanical degradation using a gentleMACS tissue dissasociator (Miltenyi).

Potential methodological problems could be that some lymphocytes might be left in the tissue and give a skewed cell population distribution, specifically cells close to the "sticky" keratinized part of the medulla could be more difficult to extract than cells in the cortex. The mechanical dissociation of the epithelial cells causes cell death, evident by the presence DNA and the need of DNase in the enzyme mixture. The enzymatical treatment might affect and strip away sensitive cell surface markers.

5.1.2 FLOW CYTOMETRY AND FACS

For Paper I peripheral blood mononuclear cells were sorted into CD4⁺, CD8⁺ T cells and CD19+ B cells with an i-Cyt Synergi cell sorter (Sony Biotechnology Inc, San Jose, California).

For **Paper II** CD19⁺ B cells were sorted into CD21⁺ and CD21^{-/low} and Ig switched and unswitched populations for FISH analysis and functional coculture experiments. Due to the small populations we did not manage to sort large samples, which was evident in the FISH analysis where the switched cell count did not reach 50. The flow cytometer used in the B cell paper was a

FACS lyric (BD Biosciences, San Diego, California) and the sorter a SH800Z (Sony Biotechnology Inc, San Jose, California).

All sorting experiments were performed with \geq 95% purity.

The image stream images in **paper II** were acquired with ImageStream X Mark II imaging flow cytometer (Amnis, Seattle, Wash). Thymic epithelial cells were run in parallel to have a positive control when analyzing AIRE.

5.1.3 IMMUNE REPERTOIRE SEQUENCING

The benefit of the DNA sequencing-based analysis of receptor repertoire, used in **paper I,** over Flow cytometry analysis of the V_B families is that it allows a detailed analysis of the repertoire and a resolution where it is possible to distinguish clones and not only chain usage. The samples came from DNA originally sorted for telomere length analysis⁶⁷. The sample populations were $CD4^+$ and $CD8^+$ T cells and $CD19^+$ B cells.

Multiplex PCR was performed according to BIOMED-2 guidelines¹⁷⁷ with specific primers to amplify $V\beta$ or IGH chains. A possible problem with this approach is the primer efficacy, if it differs it could give a skewing in the amplification step. However, this would affect the controls and the thymectomy samples to the same extent, and therefore it should not affect the comparison between the two.

The amplified reads were sequenced on an MiSeq sequencer (Illumina, San Diego, California). The clonality was analyzed by a method described by Boyd⁶⁸ were six parallel wells were amplified and sequenced and the occurrence of the same clone (determined by amino acid usage) in the separate wells is named coincidence. The coincidences are presumed to originate from the same T cell, meaning the fewer coincidences the more diverse repertoire.

We analyzed $CD19⁺$ B cells for somatic hypermutations (data not shown) to assess whether the lack for clonality among T cells affected the B cell compartment in Ig switching and affinity maturation in germinal centers. We could find no difference between the groups, although it would have increased the chances of finding differences if the sorted populations had been more specific and for example distinguished between naïve and memory B cells.

5.1.4 IMMUNOHISTOCHEMISTRY

Almost all sections used in both **paper II & III** were OCT embedded frozen samples. Although more time-consuming at the first tissue processing step the antigens are preserved without modification, which is an advantage compared to formaldehyde fixation that cause denaturation of proteins. Formaldehyde fixation gives a better morphology but risks that certain antigens are not recognized by the antibodies. The downstream steps of formaldehyde fixed samples require more time and the use of harmful chemicals, making frozen samples the preferred alternative.

All stains included in this thesis were well defined and the risk of detecting false positives as true staining were minimal. All images were acquired with an LSM700 confocal microscope (Carl Zeiss AG, Oberkochen, Germany).

5.1.5 LASER MICRODISSECTION

To analyze the Hassall's corpuscles in **paper III** there was a dilemma of how to acquire samples to analyze. Creating single cell suspensions of late stage epithelial cells would require tough degradation, without knowing if all cells are properly released in the suspension and represented in the analyses. This problem has been approached by comparing TEC numbers acquired by flow cytometry and microscopy, showing a clear bias were not all cells are represented in suspension¹⁷². To be able to acquire all cells from the area surrounding the Hassall's corpuscles, and also the very center of the corpuscle, we chose to use laser microdissection. The corpuscles are distinct and identifiable without specific staining, making the protocol straightforward. Unfortunately, all the cells surrounding the structure are collected simultaneously. The epithelial cells of interest are diluted by other cell types present in the vicinity. One way of approaching this problem could be to subtract e.g. thymocyte markers from the sequencing data.

5.1.6 QPCR

qPCR was performed in **Paper III** with TaqMan assay. Duplex PCR was used to save cDNA from samples. In duplex two different primers are used in the same well, with different fluorescent molecules attached to the probes. The two duplexed assays were diluted previously to limit the primer amount of the more expressed gene to prevent it from consuming all reagent. Samples were run separately and in same well simultaneously to compare the CT curves. The samples were analyzed with a Viia 7 real time PCR system. (Thermo Fisher, Waltham, Massachusetts)

6 PATIENT SAMPLES

Thymic samples were collected at the Queen Silvias Children's Hospital, Sahlgrenska University hospital, Gothenburg. Due to the location of the thymus in the upper chest cavity it blocks the surgeon's access to the heart and needs to be removed during corrective cardiac surgery.

The health of children undergoing surgery can be variable and may affect our studies, this needs to be kept in mind when analyzing human samples. Up to 10% of children with congenital heart disease have anomalies that fit into a syndrome. This is not always fully investigated at time of surgery and therefore not always registered for the samples in our biobank⁴⁰.

Some heart diagnoses are more subjected to cyanosis than others. The most common cyanotic congenital heart diseases are tetralogy of Fallot and transposition of the great arteries $(TGA)^{178, 179}$. It is not known to what extent cyanosis might affect the thymus, newborns suffering from this affliction are also discovered early and treated. A study performed on human thymic tissue could not find any effects on thymic subset distributions between cyanotic and well saturated children. However, it was not specified what subsets were investigated³⁶. A later study found a difference in the numbers of FoxP3 expressing cells in the thymic medulla in children with a cyanotic congenital heart disease who had greater numbers than non-cyanotic controls¹⁸⁰. Cyanosis can induce FoxP3 through hypoxia inducible factor- $1\alpha^{181}$ and cause an unintentional bias. One of the most common cyanotic heart defects, TGA, afflicts males more often than females¹⁸²⁻¹⁸⁴. This could introduce a gender bias if studying FoxP3.

The field of immunology had the fewest articles that specified sex out of ten research fields according to a review¹⁸⁵. This is surprising since there are large immunological differences reported between the sexes, for example many of the autoimmune diseases are more common in women¹⁸⁶. It has also been shown that males have higher levels of $AIRE¹⁷¹$ and that women express less AIRE after puberty. It was also shown that male castration decreases thymic AIRE expression. Cultures of human TEC showed a downregulation of AIRE upon addition of estrogen¹⁸⁷. The major differences arise in puberty when sex hormones are expressed in higher levels, but also more discrete changes of hormones, as in minipuberty, seems to affect the infant thymus during the first vears of life¹⁸⁸. The hypothalamic-pituitary-gonadal axis is active in mid gestation but silenced towards full term. At birth the axis reactivates which

leads to a testosterone rise in males that peaks at 1-3 months of age known as "minipuberty". In females it leads to an increase in oestradiol levels $170, 189$.

There are both limitations and benefits in working with human samples rather than mouse models. Some of the limitations includes less possibilities for *in vivo* manipulations since all experiments on human thymic tissue need to be performed *in vitro*. The sample access is more unreliable and difficult to plan. Despite all this it is a great opportunity to be able to study human tissue as animal models and cell lines are usually not as representative.

CONCLUDING REMARKS

In **paper I** we investigated the effects of thymectomy in early childhood on T and B cell receptor repertoires in peripheral blood. The T cell compartment showed a higher clonality, indicating an effect of the absent thymic output. The B cell clonality was not affected. The impact on the T cell compartment 18 years after thymectomy suggests an impending problem, which might also affect the B cell population with time.

Paper II reveals a new characteristic about the thymic B cells, half of them expressed low levels of CD21, associating them to an already know population of B cells usually found in diseases characterized by chronic immune stimulation. A surprising number of the thymic B cells were switched, higher among the CD21^{-/low} population than among the CD21⁺. They were also more effective in thymocyte crosstalk, probably due to their high levels of CD40 and MHC class II.

Paper III aims to characterize the last developmental stage of the thymic epithelial cells. The Hassall's corpuscles have since their discovery been an enigmatic structure. We showed with the help of laser microdissection that the cell layers surrounding the Hassall's have an enrichment of skin markers, and that the core of the structure itself contains late stage keratins and bacterial defensive proteins also found on the skin.

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REFERENCES

1. Gudmundsdottir J, Söderling J, Berggren H, et al. Long-term clinical effects of early thymectomy: Associations with autoimmune diseases, cancer, infections, and atopic diseases. *Journal of Allergy and Clinical Immunology* 2018; 141: 2294-2297.e2298. Article. DOI: 10.1016/j.jaci.2018.01.037.

2. Laios K. The thymus gland in ancient Greek medicine. *Hormones (Athens, Greece)* 2018; 17: 285-286. 2018/06/03. DOI: 10.1007/s42000-018-0026-4.

3. Ribatti D, Crivellato E and Vacca A. Miller's seminal studies on the role of thymus in immunity. *Clinical and experimental immunology* 2006; 144: 371-375. 2006/06/01. DOI: 10.1111/j.1365-2249.2006.03060.x.

4. Miller JFAP. The discovery of thymus function and of thymusderived lymphocytes. *Immunological reviews* 2002; 185: 7-14. DOI: 10.1034/j.1600-065X.2002.18502.x.

5. Cooper SA. *The Anatomy of the Thymus Gland*. London: Longman, Rees, Orme, Green, and Brown, 1832.

6. Miller JFAP. Immunological function of the thymus. *The Lancet* 1961; 278: 748-749. DOI: https://doi.org/10.1016/S0140- 6736(61)90693-6.

7. Björk M. *Kalvbrässkontroversen : Veterinärmedicinaren Elis Sandberg, cancermedlet THX och skolmedicinen i Sverige från 1952 till 1989.* . Lychnos, 2016.

8. Farmaceutiska specialiteter i Sverige, Enzythym,

https://www.fass.se/LIF/product?userType=0&nplId=19931118000058 (accessed 9th March 2019).

9. Wang J, Sekai M, Matsui T, et al. Hassall's corpuscles with cellular-senescence features maintain IFNalpha production through neutrophils and pDC activation in the thymus. *International immunology* 2018 2018/12/12. DOI: 10.1093/intimm/dxy073.

10. Muller E. Localization of eosinophils in the thymus by the peroxidase reaction. *Histochemistry* 1977; 52: 273-279. 1977/06/08.

11. Lee I, Yu E, Good RA, et al. Presence of eosinophilic precursors in the human thymus: evidence for intra-thymic differentiation of cells in eosinophilic lineage. *Pathology international* 1995; 45: 655-662. 1995/09/01.

12. Raica M, Cimpean AM, Encica S, et al. Increased mast cell density and microvessel density in the thymus of patients with myasthenia gravis. *Romanian journal of morphology and embryology = Revue roumaine de morphologie et embryologie* 2007; 48: 11-16. 2007/05/16.

13. Puchtler H, Meloan SN, Branch BW, et al. Myoepithelial cells in human thymus: staining, polarization and fluorescence microscopic studies. *Histochemistry* 1975; 45: 163-176. 1975/11/21.

14. Wakkach A, Poea S, Chastre E, et al. Establishment of a human thymic myoid cell line. Phenotypic and functional characteristics. *The American journal of pathology* 1999; 155: 1229-1240. 1999/10/09. DOI: 10.1016/s0002-9440(10)65225-x.

15. Miller CN, Proekt I, von Moltke J, et al. Thymic tuft cells promote an IL-4-enriched medulla and shape thymocyte development. *Nature* 2018; 559: 627-631. 2018/07/20. DOI: 10.1038/s41586-018-0345-2.

16. Bornstein C, Nevo S, Giladi A, et al. Single-cell mapping of the thymic stroma identifies IL-25-producing tuft epithelial cells. *Nature* 2018; 559: 622-626. 2018/07/20. DOI: 10.1038/s41586-018-0346-1.

17. Ribatti D. The discovery of the blood-thymus barrier. *Immunology letters* 2015; 168: 325-328. 2015/11/03. DOI: 10.1016/j.imlet.2015.10.014.

18. Irino S, Takasugi N and Murakami T. Vascular architecture of thymus and lymph nodes, blood vessels, transmural passage of lymphocytes, and cell-interactions. *Scanning electron microscopy* 1981: 89-98. 1981/01/01.

19. Zarnitsyna VI, Evavold BD, Schoettle LN, et al. Estimating the diversity, completeness, and cross-reactivity of the T cell repertoire. *Frontiers in Immunology* 2013; 4. Article. DOI: 10.3389/fimmu.2013.00485.

20. Klein L, Kyewski B, Allen PM, et al. Positive and negative selection of the T cell repertoire: what thymocytes see (and don't see). *Nature reviews Immunology* 2014; 14: 377-391. 2014/05/17. DOI: 10.1038/nri3667.

21. Huang CY, Sleckman BP and Kanagawa O. Revision of T cell receptor α chain genes is required for normal T lymphocyte development. *Proceedings of the National Academy of Sciences of the United States of America* 2005; 102: 14356-14361. Article. DOI: 10.1073/pnas.0505564102.

22. Naeher D, Daniels MA, Hausmann B, et al. A constant affinity threshold for T cell tolerance. *J Exp Med* 2007; 204: 2553-2559. 2007/10/17. DOI: 10.1084/jem.20070254.

23. Nedjic J, Aichinger M, Emmerich J, et al. Autophagy in thymic epithelium shapes the T-cell repertoire and is essential for tolerance. *Nature* 2008; 455: 396-400. 2008/08/15. DOI: 10.1038/nature07208.

24. Derbinski J, Schulte A, Kyewski B, et al. Promiscuous gene expression in medullary thymic epithelial cells mirrors the peripheral self. *Nat Immunol* 2001; 2: 1032-1039. 2001/10/16. DOI: 10.1038/ni723.

25. Anderson MS, Venanzi ES, Klein L, et al. Projection of an immunological self shadow within the thymus by the aire protein. *Science (New York, NY)* 2002; 298: 1395-1401. 2002/10/12. DOI: 10.1126/science.1075958.

26. Liston A, Lesage S, Wilson J, et al. Aire regulates negative selection of organ-specific T cells. *Nat Immunol* 2003; 4: 350-354. 2003/03/04. DOI: 10.1038/ni906.

27. Koble C and Kyewski B. The thymic medulla: a unique microenvironment for intercellular self-antigen transfer. *J Exp Med* 2009; 206: 1505-1513. 2009/07/01. DOI: 10.1084/jem.20082449.

28. Skogberg G, Telemo E and Ekwall O. Exosomes in the Thymus: Antigen Transfer and Vesicles. *Frontiers in Immunology* 2015; 6: 366. 10.3389/fimmu.2015.00366.

29. den Braber I, Mugwagwa T, Vrisekoop N, et al. Maintenance of peripheral naive T cells is sustained by thymus output in mice but not humans. *Immunity* 2012; 36: 288-297. 2012/03/01. DOI: 10.1016/j.immuni.2012.02.006.

30. Allenspach E and Torgerson TR. Autoimmunity and Primary Immunodeficiency Disorders. *Journal of clinical immunology* 2016; 36 Suppl 1: 57-67. 2016/05/24. DOI: 10.1007/s10875-016-0294-1.

31. Owen DL, Mahmud SA, Sjaastad LE, et al. Thymic regulatory T cells arise via two distinct developmental programs. *Nat Immunol* 2019; 20: 195-205. 2019/01/16. DOI: 10.1038/s41590-018-0289-6.

32. Lu F-T, Yang W, Wang Y-H, et al. Thymic B cells promote thymus-derived regulatory T cell development and proliferation. *Journal of autoimmunity* 2015; 61: 62-72. DOI: https://doi.org/10.1016/j.jaut.2015.05.008.

33. Aspinall R and Andrew D. Thymic involution in aging. *Journal of clinical immunology* 2000; 20: 250-256. 2000/08/12.

34. Steinmann GG. Changes in the human thymus during aging. *Current topics in pathology* 1986; 75: 43-88. Review.

35. Steinmann GG, Klaus B and Muller-Hermelink HK. The involution of the ageing human thymic epithelium is independent of puberty. A morphometric study. *Scandinavian Journal of Immunology* 1985; 22: 563- 575.

36. Weerkamp F, de Haas EFE, Naber BAE, et al. Age-related changes in the cellular composition of the thymus in children. *Journal of Allergy and Clinical Immunology* 2005; 115: 834-840. DOI: 10.1016/j.jaci.2004.10.031.

37. Barbouti A, Evangelou K, Pateras IS, et al. In situ evidence of cellular senescence in Thymic Epithelial Cells (TECs) during human thymic involution. *Mechanisms of Ageing and Development* 2019; 177: 88-90. Article. DOI: 10.1016/j.mad.2018.02.005.

38. Zhu X, Gui J, Dohkan J, et al. Lymphohematopoietic progenitors do not have a synchronized defect with age-related thymic involution. *Aging cell* 2007; 6: 663-672. 2007/08/08. DOI: 10.1111/j.1474- 9726.2007.00325.x.

39. Rode I, Martins VC, Kublbeck G, et al. Foxn1 Protein Expression in the Developing, Aging, and Regenerating Thymus. *Journal of immunology (Baltimore, Md : 1950)* 2015; 195: 5678-5687. 2015/11/06. DOI: 10.4049/jimmunol.1502010.

40. Noonan JA. A history of pediatric specialties: The development of pediatric cardiology. *Pediatric Research* 2004; 56: 298-306. Review. DOI: 10.1203/01.PDR.0000132662.73362.96.

41. Petrie HT, Livak F, Schatz DG, et al. Multiple rearrangements in T cell receptor alpha chain genes maximize the production of useful thymocytes. *The Journal of Experimental Medicine* 1993; 178: 615. DOI: 10.1084/jem.178.2.615.

42. Nakagawa Y, Ohigashi I, Nitta T, et al. Thymic nurse cells provide microenvironment for secondary T cell receptor α rearrangement in cortical thymocytes. *Proceedings of the National Academy of Sciences of the United States of America* 2012; 109: 20572-20577. Article. DOI: 10.1073/pnas.1213069109.

43. Hazenberg MD, Borghans JAM, de Boer RJ, et al. Thymic output: A bad TREC record. *Nature Immunology* 2003; 4: 97-99. Note. DOI: 10.1038/ni0203-97.

44. Douek DC, McFarland RD, Keiser PH, et al. Changes in thymic function with age and during the treatment of HIV infection. *Nature* 1998; 396: 690-695. 1999/01/01. DOI: 10.1038/25374.

45. Gui J, Mustachio LM, Su D-M, et al. Thymus Size and Agerelated Thymic Involution: Early Programming, Sexual Dimorphism, Progenitors and Stroma. *Aging and disease* 2012; 3: 280-290.

46. Hince M, Sakkal S, Vlahos K, et al. The role of sex steroids and gonadectomy in the control of thymic involution. *Cellular immunology* 2008; 252: 122-138. DOI: https://doi.org/10.1016/j.cellimm.2007.10.007.

47. Kendall MD and Clarke AG. The thymus in the mouse changes its activity during pregnancy: a study of the microenvironment. *Journal of anatomy* 2000; 197: 393-411. 2000/11/23. DOI: undefined.

48. Bjelakovic G, Stojanovic I, Jevtovic-Stoimenov T, et al. Thymus as a target tissue of glucocorticoid action: what are the consequences of glucocorticoids thymectomy? *Journal of basic and clinical physiology and pharmacology* 2009; 20: 99-125. 2009/08/11.

49. Hellberg S, Mehta RB, Forsberg A, et al. Maintained thymic output of conventional and regulatory T cells during human pregnancy. *Journal of Allergy and Clinical Immunology* 2019; 143: 771-775.e777. DOI: 10.1016/j.jaci.2018.09.023.

50. Griffith AV, Fallahi M, Venables T, et al. Persistent degenerative changes in thymic organ function revealed by an inducible model of organ regrowth. *Aging cell* 2012; 11: 169-177. 2011/11/23. DOI: 10.1111/j.1474-9726.2011.00773.x.

51. Bertho JM, Demarquay C, Moulian N, et al. Phenotypic and immunohistological analyses of the human adult thymus: evidence for an active thymus during adult life. *Cellular immunology* 1997; 179: 30-40. 1997/07/10. DOI: 10.1006/cimm.1997.1148.

52. Naylor K, Li G, Vallelo AN, et al. The influence of age on T cell generation and TCR diversity. *Journal of Immunology* 2005; 174: 7446- 7452. Article. DOI: 10.4049/jimmunol.174.11.7446.

53. Palmer DB. The Effect of Age on Thymic Function. *Front Immunol* 2013; 4: 316. 2013/10/11. DOI: 10.3389/fimmu.2013.00316.

54. Moretta L, Mingari MC, Webb SR, et al. Imbalances in T cell subpopulations associated with immunodeficiency and autoimmune syndromes. *European journal of immunology* 1977; 7: 696-700. 1977/10/01. DOI: 10.1002/eji.1830071009.

55. Brearley S, Gentle TA, Baynham MI, et al. Immunodeficiency following neonatal thymectomy in man. *Clinical and experimental immunology* 1987; 70: 322-327. 1987/11/01.

56. Okoye AA, Rohankhedkar M, Konfe AL, et al. Effect of IL-7 therapy on naive and memory T cell homeostasis in aged rhesus macaques. *Journal of Immunology* 2015; 195: 4292-4305. Article. DOI: 10.4049/jimmunol.1500609.

57. Wells WJ, Parkman R, Smogorzewska E, et al. Neonatal thymectomy: does it affect immune function? *The Journal of thoracic and cardiovascular surgery* 1998; 115: 1041-1046. 1998/05/30. DOI: 10.1016/s0022-5223(98)70403-9.

58. Halnon NJ, Jamieson B, Plunkett M, et al. Thymic function and impaired maintenance of peripheral T cell populations in children with congenital heart disease and surgical thymectomy. *Pediatr Res* 2005; 57: 42- 48. 2004/11/09. DOI: 10.1203/01.Pdr.0000147735.19342.De.

59. Appay V, Sauce D and Prelog M. The role of the thymus in immunosenescence: lessons from the study of thymectomized individuals. *Aging* 2010; 2: 78-81. 2010/04/01. DOI: 10.18632/aging.100122.

60. Van Den Broek T, Delemarre EM, Janssen WJM, et al. Neonatal thymectomy reveals differentiation and plasticity within human naive T cells. *Journal of Clinical Investigation* 2016; 126: 1126-1136. Article. DOI: 10.1172/JCI84997.

61. Mancebo E, Clemente J, Sanchez J, et al. Longitudinal analysis of immune function in the first 3 years of life in thymectomized neonates during cardiac surgery. *Clinical & Experimental Immunology* 2008; 154: 375- 383. DOI: 10.1111/j.1365-2249.2008.03771.x.

62. Prelog M, Keller M, Geiger R, et al. Thymectomy in early childhood: Significant alterations of the CD4+CD45RA+CD62L+ T cell compartment in later life. *Clinical Immunology* 2009; 130: 123-132. Article. DOI: 10.1016/j.clim.2008.08.023.

63. Silva SL, Albuquerque A, Amaral AJ, et al. Autoimmunity and allergy control in adults submitted to complete thymectomy early in infancy. *PLoS ONE* 2017; 12. Article. DOI: 10.1371/journal.pone.0180385.

64. Silva SL, Albuquerque AS, Serra-Caetano A, et al. Human naïve regulatory T-cells feature high steady-state turnover and are maintained by IL-7. *Oncotarget* 2016; 7: 12163-12175. Article. DOI: 10.18632/oncotarget.7512.

65. van den Broek T, Madi A, Delemarre EM, et al. Human neonatal thymectomy induces altered B-cell responses and autoreactivity. *European journal of immunology* 2017; 47: 1970-1981. Article. DOI: 10.1002/eji.201746971.

66. Halnon NJ, Cooper P, Chen DY, et al. Immune dysregulation after cardiothoracic surgery and incidental thymectomy: maintenance of regulatory T cells despite impaired thymopoiesis. *Clinical & developmental immunology* 2011; 2011: 915864. 2011/07/22. DOI: 10.1155/2011/915864.

67. Gudmundsdottir J, Oskarsdottir S, Skogberg G, et al. Early thymectomy leads to premature immunologic ageing: An 18-year follow-up. *J Allergy Clin Immunol* 2016 2016/07/12. DOI: 10.1016/j.jaci.2016.05.014.

68. Boyd SD, Marshall EL, Merker JD, et al. Measurement and clinical monitoring of human lymphocyte clonality by massively parallel VDJ pyrosequencing. *Science translational medicine* 2009; 1: 12ra23. 2010/02/18.

69. Zlamy M, Würzner R, Holzmann H, et al. Antibody dynamics after tick-borne encephalitis and measles–mumps–rubella vaccination in children post early thymectomy. *Vaccine* 2010; 28: 8053-8060. DOI: https://doi.org/10.1016/j.vaccine.2010.10.002.

70. Prelog M, Wilk C, Keller M, et al. Diminished response to tickborne encephalitis vaccination in thymectomized children. *Vaccine* 2008; 26: 595-600. 2008/01/08. DOI: 10.1016/j.vaccine.2007.11.074.

71. Ogle BM, West LJ, Driscoll DJ, et al. Effacing of the T Cell Compartment by Cardiac Transplantation in Infancy. *The Journal of Immunology* 2006; 176: 1962. DOI: 10.4049/jimmunol.176.3.1962.

72. Yager EJ, Ahmed M, Lanzer K, et al. Age-associated decline in T cell repertoire diversity leads to holes in the repertoire and impaired immunity to influenza virus. *The Journal of Experimental Medicine* 2008; 205: 711. DOI: 10.1084/jem.20071140.

73. Avdimiretz N, Seitz S, Kim T, et al. Allergies and autoimmune disorders in children after heart transplantation. *Clinical Transplantation* 2018; 32. Article. DOI: 10.1111/ctr.13400.

74. Thyssen JP, Andersen YMF, Zhang H, et al. Incidence of pediatric atopic dermatitis following thymectomy: A Danish register study. *Allergy: European Journal of Allergy and Clinical Immunology* 2018; 73: 1741-1743. Letter. DOI: 10.1111/all.13457.

75. Sauce D, Larsen M, Fastenackels S, et al. Lymphopenia-driven homeostatic regulation of naive T cells in elderly and thymectomized young adults. *Journal of immunology (Baltimore, Md : 1950)* 2012; 189: 5541-5548. 2012/11/09. DOI: 10.4049/jimmunol.1201235.

76. van Gent R, Schadenberg AW, Otto SA, et al. Long-term restoration of the human T-cell compartment after thymectomy during infancy: a role for thymic regeneration? *Blood* 2011; 118: 627-634. 2011/06/02. DOI: 10.1182/blood-2011-03-341396.

77. Nasi M, Troiano L, Lugli E, et al. Thymic output and functionality of the IL-7/IL-7 receptor system in centenarians: implications for the neolymphogenesis at the limit of human life. *Aging cell* 2006; 5: 167-175. 2006/04/22. DOI: 10.1111/j.1474-9726.2006.00204.x.

78. Aspinall R, Pido-Lopez J, Imami N, et al. Old rhesus macaques treated with interleukin-7 show increased TREC levels and respond well to

influenza vaccination. *Rejuvenation research* 2007; 10: 5-17. 2007/03/24. DOI: 10.1089/rej.2006.9098.

79. Goronzy JJ, Fang F, Cavanagh MM, et al. Naive T Cell Maintenance and Function in Human Aging. *The Journal of Immunology* 2015; 194: 4073. DOI: 10.4049/jimmunol.1500046.

80. Wheeler CJ, Black KL, Liu G, et al. Thymic CD8+ T cell production strongly influences tumor antigen recognition and age-dependent glioma mortality. *Journal of immunology (Baltimore, Md : 1950)* 2003; 171: 4927-4933. 2003/10/22.

81. Palmer S, Albergante L, Blackburn CC, et al. Thymic involution and rising disease incidence with age. *Proceedings of the National Academy of Sciences* 2018; 115: 1883. DOI: 10.1073/pnas.1714478115.

82. Yamano T, Nedjic J, Hinterberger M, et al. Thymic B Cells Are Licensed to Present Self Antigens for Central T Cell Tolerance Induction. *Immunity* 2015; 42: 1048-1061. 2015/06/14. DOI: 10.1016/j.immuni.2015.05.013.

83. Perera J, Zheng Z, Li S, et al. Self-Antigen-Driven Thymic B Cell Class Switching Promotes T Cell Central Tolerance. *Cell reports* 2016; 17: 387-398. DOI: https://doi.org/10.1016/j.celrep.2016.09.011.

84. Gies V, Guffroy A, Danion F, et al. B cells differentiate in Human thymus and express AIRE. *J Allergy Clin Immunol* 2016 2016/11/20. DOI: 10.1016/j.jaci.2016.09.044.

85. Isaacson PG, Norton AJ and Addis BJ. The human thymus contains a novel population of B lymphocytes. *The Lancet* 1987; 330: 1488- 1491. DOI: 10.1016/S0140-6736(87)92622-5.

86. Perera J, Meng L, Meng F, et al. Autoreactive thymic B cells are efficient antigen-presenting cells of cognate self-antigens for T cell negative selection. *Proc Natl Acad Sci U S A* 2013; 110: 17011-17016. 2013/10/02. DOI: 10.1073/pnas.1313001110.

87. Akashi K, Richie LI, Miyamoto T, et al. B lymphopoiesis in the thymus. *Journal of immunology (Baltimore, Md : 1950)* 2000; 164: 5221-5226. 2000/05/09.

88. Cepeda S, Cantu C, Orozco S, et al. Age-Associated Decline in Thymic B Cell Expression of Aire and Aire-Dependent Self-Antigens. *Cell reports* 2018; 22: 1276-1287. DOI: https://doi.org/10.1016/j.celrep.2018.01.015.

89. Fujihara C, Williams JA, Watanabe M, et al. T Cell–B Cell Thymic Cross-Talk: Maintenance and Function of Thymic B Cells Requires Cognate CD40–CD40 Ligand Interaction. *The Journal of Immunology* 2014; 193: 5534. DOI: 10.4049/jimmunol.1401655.

90. Kleindienst P, Chretien I, Winkler T, et al. Functional comparison of thymic B cells and dendritic cells in vivo. *Blood* 2000; 95: 2610. 91. Walters SN, Webster KE, Daley S, et al. A role for intrathymic B cells in the generation of natural regulatory T cells. *Journal of immunology* *(Baltimore, Md : 1950)* 2014; 193: 170-176. 2014/05/30. DOI: 10.4049/jimmunol.1302519.

92. Mohammed Ali HH and Drela N. Role of thymic B cells in the development of thymus-derived regulatory T cell in vitro. *Immunology letters* 2017; 185: 56-63. 2017/03/14. DOI: 10.1016/j.imlet.2017.03.007.

93. Frommer F and Waisman A. B cells participate in thymic negative selection of murine auto-reactive CD4+ T cells. *PLoS One* 2010; 5: e15372. 2010/10/27. DOI: 10.1371/journal.pone.0015372.

94. Brocker T, Riedinger M and Karjalainen K. Targeted Expression of Major Histocompatibility Complex (MHC) Class II Molecules Demonstrates that Dendritic Cells Can Induce Negative but Not Positive Selection of Thymocytes In Vivo. *The Journal of Experimental Medicine* 1997; 185: 541. DOI: 10.1084/jem.185.3.541.

95. Kijimoto-Ochiai S, Doi N, Matsukawa H, et al. Localization of sialidase-positive cells expressing Mac-1 and immunoglobulin in the mouse thymus. *Glycoconjugate journal* 2004; 20: 375-384. 2004/07/09. DOI: 10.1023/B:GLYC.0000033994.99464.ce.

96. Reisner Y, Linker-Israeli M and Sharon N. Separation of mouse thymocytes into two subpopulations by the use of peanut agglutinin. *Cellular immunology* 1976; 25: 129-134. DOI: https://doi.org/10.1016/0008- 8749(76)90103-9.

97. Matsumoto-Mizuno T, Kijimoto-Ochiai S, Matsuoka I, et al. Existence of NEU1 sialidase on mouse thymocytes whose natural substrate is CD5. *Glycobiology* 2018; 28: 306-317. DOI: 10.1093/glycob/cwy009.

98. Kijimoto-Ochiai S, Kamimura K and Koda T. Neumedullocytes, sialidase-positive B cells in the thymus, express autoimmune regulator (AIRE). *Scientific reports* 2019; 9: 858. DOI: 10.1038/s41598-018- 37225-y.

99. Pinto AI, Smith J, Kissack MR, et al. Thymic B Cell-Mediated Attack of Thymic Stroma Precedes Type 1 Diabetes Development. *Front Immunol* 2018; 9: 1281. 2018/06/23. DOI: 10.3389/fimmu.2018.01281.

100. Flores KG, Li J and Hale LP. B cells in epithelial and perivascular compartments of human adult thymus. *Human pathology* 2001; 32: 926-934. DOI: https://doi.org/10.1053/hupa.2001.27106.

101. Nunez S, Moore C, Gao B, et al. The human thymus perivascular space is a functional niche for viral-specific plasma cells. *Science immunology* 2016; 1 2017/05/02. DOI: 10.1126/sciimmunol.aah4447.

102. Rother MB, Schreurs MW, Kroek R, et al. The Human Thymus Is Enriched for Autoreactive B Cells. *Journal of immunology (Baltimore, Md : 1950)* 2016; 197: 441-448. 2016/06/05. DOI: 10.4049/jimmunol.1501992.

103. Tonnelle C, D'Ercole C, Depraetere V, et al. Human thymic B cells largely overexpress the V(H)4 Ig gene family. A possible role in the control of tolerance in situ? *International immunology* 1997; 9: 407-414. Article. DOI: 10.1093/intimm/9.3.407.

104. Mackay IR, Masel M and Burnet FM. Thymic abnormality in systemic lupus erythematosus. *Australasian annals of medicine* 1964; 13: 5- 14. 1964/02/01.

105. Cron MA, Maillard S, Villegas J, et al. Thymus involvement in early-onset myasthenia gravis. *Annals of the New York Academy of Sciences* 2018; 1412: 137-145. Review. DOI: 10.1111/nyas.13519.

106. Berrih-Aknin S, Ruhlmann N, Bismuth J, et al. CCL21 overexpressed on lymphatic vessels drives thymic hyperplasia in myasthenia. *Annals of neurology* 2009; 66: 521-531. 2009/10/23. DOI: 10.1002/ana.21628. 107. Song Y, Zhou L, Miao F, et al. Increased frequency of thymic T follicular helper cells in myasthenia gravis patients with thymoma. *Journal of Thoracic Disease* 2016; 8: 314-322. Article. DOI: 10.21037/jtd.2016.03.03. 108. Thorarinsdottir K, Camponeschi A, Cavallini N, et al. CD21(- /low) B cells in human blood are memory cells. *Clinical and experimental immunology* 2016; 185: 252-262. 2016/03/25. DOI: 10.1111/cei.12795.

109. Ehrhardt GR, Hsu JT, Gartland L, et al. Expression of the immunoregulatory molecule FcRH4 defines a distinctive tissue-based population of memory B cells. *J Exp Med* 2005; 202: 783-791. 2005/09/15. DOI: 10.1084/jem.20050879.

110. Thorarinsdottir K, Camponeschi A, Gjertsson I, et al. CD21 - /low B cells: A Snapshot of a Unique B Cell Subset in Health and Disease. *Scand J Immunol* 2015; 82: 254-261. 2015/06/30. DOI: 10.1111/sji.12339.

111. Ehrhardt GR, Hijikata A, Kitamura H, et al. Discriminating gene expression profiles of memory B cell subpopulations. *J Exp Med* 2008; 205: 1807-1817. 2008/07/16. DOI: 10.1084/jem.20072682.

112. Moir S and Fauci AS. B cells in HIV infection and disease. *Nature reviews Immunology* 2009; 9: 235-245. 2009/03/26. DOI: 10.1038/nri2524.

113. Charles ED, Green RM, Marukian S, et al. Clonal expansion of immunoglobulin M+CD27+ B cells in HCV-associated mixed cryoglobulinemia. *Blood* 2008; 111: 1344-1356. 2007/10/19. DOI: 10.1182/blood-2007-07-101717.

114. Weiss GE, Crompton PD, Li S, et al. Atypical memory B cells are greatly expanded in individuals living in a malaria-endemic area. *Journal of immunology (Baltimore, Md : 1950)* 2009; 183: 2176-2182. 2009/07/14. DOI: 10.4049/jimmunol.0901297.

115. Agarwal S and Cunningham-Rundles C. Autoimmunity in common variable immunodeficiency. *Current allergy and asthma reports* 2009; 9: 347-352. 2009/08/13.

116. Isnardi I, Ng YS, Menard L, et al. Complement receptor 2/CD21- human naive B cells contain mostly autoreactive unresponsive clones. *Blood* 2010; 115: 5026-5036. 2010/03/17. DOI: 10.1182/blood-2009- 09-243071.

117. Odendahl M, Jacobi A, Hansen A, et al. Disturbed peripheral B lymphocyte homeostasis in systemic lupus erythematosus. *Journal of immunology (Baltimore, Md : 1950)* 2000; 165: 5970-5979. 2000/11/09.

118. Bemark M, Holmqvist J, Abrahamsson J, et al. Translational Mini-Review Series on B cell subsets in disease. Reconstitution after haematopoietic stem cell transplantation - revelation of B cell developmental pathways and lineage phenotypes. *Clinical and experimental immunology* 2012; 167: 15-25. 2011/12/03. DOI: 10.1111/j.1365-2249.2011.04469.x.

119. Rubtsov AV, Rubtsova K, Fischer A, et al. Toll-like receptor 7 (TLR7)-driven accumulation of a novel CD11 c (+) B-cell population is important for the development of autoimmunity. *Blood* 2011; 118: 1305-1315. 2011/05/06. DOI: 10.1182/blood-2011-01-331462.

120. Rubtsova K, Rubtsov AV, Thurman JM, et al. B cells expressing the transcription factor T-bet drive lupus-like autoimmunity. *The Journal of clinical investigation* 2017; 127: 1392-1404. 2017/02/28. DOI: 10.1172/jci91250.

121. Rubtsova K, Rubtsov AV, Cancro MP, et al. Age-Associated B Cells: A T-bet-Dependent Effector with Roles in Protective and Pathogenic Immunity. *Journal of immunology (Baltimore, Md : 1950)* 2015; 195: 1933- 1937. 2015/08/25. DOI: 10.4049/jimmunol.1501209.

122. Hao Y, O'Neill P, Naradikian MS, et al. A B-cell subset uniquely responsive to innate stimuli accumulates in aged mice. *Blood* 2011; 118: 1294-1304. 2011/05/13. DOI: 10.1182/blood-2011-01-330530.

123. Rubtsov AV, Rubtsova K, Kappler JW, et al. CD11c-Expressing B Cells Are Located at the T Cell/B Cell Border in Spleen and Are Potent APCs. *Journal of immunology (Baltimore, Md : 1950)* 2015; 195: 71- 79. 2015/06/03. DOI: 10.4049/jimmunol.1500055.

124. Sohn HW, Krueger PD, Davis RS, et al. FcRL4 acts as an adaptive to innate molecular switch dampening BCR signaling and enhancing TLR signaling. *Blood* 2011; 118: 6332-6341. 2011/09/13. DOI: 10.1182/blood-2011-05-353102.

125. Rossi SW, Jenkinson WE, Anderson G, et al. Clonal analysis reveals a common progenitor for thymic cortical and medullary epithelium. *Nature* 2006; 441: 988. DOI: 10.1038/nature04813.

126. Baik S, Jenkinson EJ, Lane PJL, et al. Generation of both cortical and Aire+ medullary thymic epithelial compartments from CD205+ progenitors. *European journal of immunology* 2013; 43: 589-594. DOI: 10.1002/eji.201243209.

127. Onder L, Nindl V, Scandella E, et al. Alternative NF-κB signaling regulates mTEC differentiation from podoplanin-expressing precursors in the cortico-medullary junction. *European journal of immunology* 2015; 45: 2218-2231. DOI: 10.1002/eji.201545677.

128. Rodewald H-R, Paul S, Haller C, et al. Thymus medulla consisting of epithelial islets each derived from a single progenitor. *Nature* 2001; 414: 763. DOI: 10.1038/414763a.

129. Hikosaka Y, Nitta T, Ohigashi I, et al. The cytokine RANKL produced by positively selected thymocytes fosters medullary thymic epithelial cells that express autoimmune regulator. *Immunity* 2008; 29: 438- 450. 2008/09/19. DOI: 10.1016/j.immuni.2008.06.018.

130. Gray D, Abramson J, Benoist C, et al. Proliferative arrest and rapid turnover of thymic epithelial cells expressing Aire. *J Exp Med* 2007; 204: 2521-2528. 2007/10/03. DOI: 10.1084/jem.20070795.

131. Abramson J and Anderson G. Thymic Epithelial Cells. *Annual review of immunology* 2017; 35: 85-118. 2017/02/23. DOI: 10.1146/annurevimmunol-051116-052320.

132. Takahama Y, Ohigashi I, Baik S, et al. Generation of diversity in thymic epithelial cells. *Nature Reviews Immunology* 2017; 17: 295. Review Article. DOI: 10.1038/nri.2017.12.

133. Erickson M, Morkowski S, Lehar S, et al. Regulation of thymic epithelium by keratinocyte growth factor. *Blood* 2002; 100: 3269. DOI: 10.1182/blood-2002-04-1036.

134. Rodewald H-R. Thymus Organogenesis. *Annual review of immunology* 2008; 26: 355-388. DOI: 10.1146/annurev.immunol.26.021607.090408.

135. Frank J, Pignata C, Panteleyev AA, et al. Exposing the human nude phenotype. *Nature* 1999; 398: 473-474. 1999/04/17. DOI: 10.1038/18997.

136. Pantelouris EM. Absence of Thymus in a Mouse Mutant. *Nature* 1968; 217: 370-371. DOI: 10.1038/217370a0.

137. Mecklenburg L, Nakamura M, Sundberg JP, et al. The nude mouse skin phenotype: the role of Foxn1 in hair follicle development and cycling. *Experimental and molecular pathology* 2001; 71: 171-178. 2001/10/16. DOI: 10.1006/exmp.2001.2386.

138. Brennecke P, Reyes A, Pinto S, et al. Single-cell transcriptome analysis reveals coordinated ectopic gene-expression patterns in medullary thymic epithelial cells. *Nat Immunol* 2015; 16: 933-941. 2015/08/04. DOI: 10.1038/ni.3246.

139. Sansom SN, Shikama-Dorn N, Zhanybekova S, et al. Population and single-cell genomics reveal the Aire dependency, relief from Polycomb silencing, and distribution of self-antigen expression in thymic epithelia. *Genome Res* 2014; 24: 1918-1931. 2014/09/17. DOI: 10.1101/gr.171645.113.

140. Derbinski J, Gäbler J, Brors B, et al. Promiscuous gene expression in thymic epithelial cells is regulated at multiple levels. *The Journal of Experimental Medicine* 2005; 202: 33. DOI: 10.1084/jem.20050471.

141. Koh AS, Kuo AJ, Park SY, et al. Aire employs a histonebinding module to mediate immunological tolerance, linking chromatin regulation with organ-specific autoimmunity. *Proc Natl Acad Sci U S A* 2008; 105: 15878-15883. 2008/10/09. DOI: 10.1073/pnas.0808470105.

142. Org T, Rebane A, Kisand K, et al. AIRE activated tissue specific genes have histone modifications associated with inactive chromatin. *Hum Mol Genet* 2009; 18: 4699-4710. 2009/09/12. DOI: 10.1093/hmg/ddp433.

143. Takaba H, Morishita Y, Tomofuji Y, et al. Fezf2 Orchestrates a Thymic Program of Self-Antigen Expression for Immune Tolerance. *Cell* 2015; 163: 975-987. 2015/11/07. DOI: 10.1016/j.cell.2015.10.013.

144. Aaltonen J, Björses P, Perheentupa J, et al. An autoimmune disease, APECED, caused by mutations in a novel gene featuring two PHDtype zinc-finger domains. *Nature genetics* 1997; 17: 399-403. DOI: 10.1038/ng1297-399.

145. Ahonen P, Myllarniemi S, Sipila I, et al. Clinical variation of autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED) in a series of 68 patients. *The New England journal of medicine* 1990; 322: 1829-1836. 1990/06/28. DOI: 10.1056/nejm199006283222601.

146. Puel A, Doffinger R, Natividad A, et al. Autoantibodies against IL-17A, IL-17F, and IL-22 in patients with chronic mucocutaneous candidiasis and autoimmune polyendocrine syndrome type I. *J Exp Med* 2010; 207: 291- 297. 2010/02/04. DOI: 10.1084/jem.20091983.

147. Michels AW and Gottlieb PA. Autoimmune polyglandular syndromes. *Nature reviews Endocrinology* 2010; 6: 270-277. 2010/03/24. DOI: 10.1038/nrendo.2010.40.

148. Marcovecchio GE, Bortolomai I, Ferrua F, et al. Thymic Epithelium Abnormalities in DiGeorge and Down Syndrome Patients Contribute to Dysregulation in T Cell Development. *Frontiers in Immunology* 2019; 10: 447. 10.3389/fimmu.2019.00447.

149. Skogberg G, Lundberg V, Lindgren S, et al. Altered expression of autoimmune regulator in infant down syndrome thymus, a possible contributor to an autoimmune phenotype. *Journal of immunology (Baltimore, Md : 1950)* 2014; 193: 2187-2195. 2014/07/20. DOI: 10.4049/jimmunol.1400742.

150. Ram G and Chinen J. Infections and immunodeficiency in Down syndrome. *Clinical and experimental immunology* 2011; 164: 9-16. Review. DOI: 10.1111/j.1365-2249.2011.04335.x.

151. Prada N, Nasi M, Troiano L, et al. Direct analysis of thymic function in children with Down's syndrome. *Immunity and Ageing* 2005; 2. Article. DOI: 10.1186/1742-4933-2-4.

152. Nishijima H, Kajimoto T, Matsuoka Y, et al. Paradoxical development of polymyositis-like autoimmunity through augmented expression of autoimmune regulator (AIRE). *Journal of autoimmunity* 2018; 86: 75-92. 2017/09/22. DOI: 10.1016/j.jaut.2017.09.006.

153. Kelleher P and Misbah SA. What is Good's syndrome? Immunological abnormalities in patients with thymoma. *Journal of clinical pathology* 2003; 56: 12-16. 2002/12/25.

154. Savino W. The thymus is a common target organ in infectious diseases. *PLoS pathogens* 2006; 2: e62. 2006/07/19. DOI: 10.1371/journal.ppat.0020062.

155. Herold MJ, McPherson KG and Reichardt HM. Glucocorticoids in T cell apoptosis and function. *Cellular and molecular life sciences : CMLS* 2006; 63: 60-72. 2005/11/30. DOI: 10.1007/s00018-005-5390-y.

156. Perry LL, Hotchkiss JD and Lodmell DL. Murine susceptibility to street rabies virus is unrelated to induction of host lymphoid depletion. *Journal of immunology (Baltimore, Md : 1950)* 1990; 144: 3552-3557. 1990/05/01.

157. Valentin H, Azocar O, Horvat B, et al. Measles Virus Infection Induces Terminal Differentiation of Human Thymic Epithelial Cells. *Journal of virology* 1999; 73: 2212.

158. Wang X, Laan M, Bichele R, et al. Post-Aire maturation of thymic medullary epithelial cells involves selective expression of keratinocytespecific autoantigens. *Front Immunol* 2012; 3: 19. 2012/03/27. DOI: 10.3389/fimmu.2012.00019.

159. Yano M, Kuroda N, Han H, et al. Aire controls the differentiation program of thymic epithelial cells in the medulla for the establishment of self-tolerance. *J Exp Med* 2008; 205: 2827-2838. 2008/11/19. DOI: 10.1084/jem.20080046.

160. Gillard GO, Dooley J, Erickson M, et al. Aire-dependent alterations in medullary thymic epithelium indicate a role for Aire in thymic epithelial differentiation. *Journal of immunology (Baltimore, Md : 1950)* 2007; 178: 3007-3015. 2007/02/22.

161. Metzger TC, Khan IS, Gardner JM, et al. Lineage tracing and cell ablation identify a post-Aire-expressing thymic epithelial cell population. *Cell reports* 2013; 5: 166-179. 2013/10/08. DOI: 10.1016/j.celrep.2013.08.038.

162. Jaroslow BN. Genesis of Hassall's corpuscles. *Nature* 1967; 215: 408-409. 1967/07/22.

163. Bodey B and Kaiser HE. Development of Hassall's bodies of the thymus in humans and other vertebrates (especially mammals) under physiological and pathological conditions: immunocytochemical, electronmicroscopic and in vitro observations. *In vivo (Athens, Greece)* 1997; 11: 61-85. 1997/01/01.

164. Bodey B, Bodey B, Jr., Siegel SE, et al. Novel insights into the function of the thymic Hassall's bodies. *In vivo (Athens, Greece)* 2000; 14: 407-418. 2000/07/25.

165. Kouvalainen K. Significance of Hassall's corpuscles in the light of their morphological and histochemical appearance. *Annales medicinae experimentalis et biologiae Fenniae* 1964; 42: 177-184. 1964/01/01.

166. Kater L. A Note On Hassall's Corpuscles. *Contemporary Topics In Immunobiology* 1973: 101-109.

167. Pierscinski A. Formation of Hassall's corpuscles owing to keratinization of thymic epithelial cells. *Folia biologica* 1979; 27: 255-262. 1979/01/01.

168. Watanabe N, Wang YH, Lee HK, et al. Hassall's corpuscles instruct dendritic cells to induce CD4+CD25+ regulatory T cells in human thymus. *Nature* 2005; 436: 1181-1185. 2005/08/27. DOI: 10.1038/nature03886.

169. Liberti EA, Konig B, Jr. and Adamo J. Contribution to the study of Hassall corpuscles in human fetuses. *Zeitschrift fur mikroskopischanatomische Forschung* 1986; 100: 253-261. 1986/01/01.

170. Kuiri-Hanninen T, Sankilampi U and Dunkel L. Activation of the hypothalamic-pituitary-gonadal axis in infancy: minipuberty. *Hormone research in paediatrics* 2014; 82: 73-80. 2014/07/12. DOI: 10.1159/000362414.

171. Zhu ML, Bakhru P, Conley B, et al. Sex bias in CNS autoimmune disease mediated by androgen control of autoimmune regulator. *Nat Commun* 2016; 7: 11350. 2016/04/14. DOI: 10.1038/ncomms11350.

172. Sakata M, Ohigashi I and Takahama Y. Cellularity of Thymic Epithelial Cells in the Postnatal Mouse. *Journal of immunology (Baltimore, Md : 1950)* 2018; 200: 1382-1388. 2018/01/05. DOI: 10.4049/jimmunol.1701235.

173. Mischke D, Korge BP, Marenholz I, et al. Genes encoding structural proteins of epidermal cornification and S100 calcium-binding proteins form a gene complex ("epidermal differentiation complex") on human chromosome 1q21. *The Journal of investigative dermatology* 1996; 106: 989- 992. 1996/05/01.

174. Folster-Holst R, Rohrer T and Jung AM. Dermatological aspects of the S2k guidelines on Down syndrome in childhood and adolescence. *Journal der Deutschen Dermatologischen Gesellschaft = Journal of the German Society of Dermatology : JDDG* 2018; 16: 1289-1295. 2018/10/10. DOI: 10.1111/ddg.13665.

175. Penrith ML and Huchzermeyer FW. Thymic necrosis in slaughtered Nile crocodiles. *Journal of the South African Veterinary Association* 1993; 64: 128-130. 1993/09/01.

176. Cave AJE. The thymus gland in three genera of rhinoceros. *Proceedings of the Zoological Society of London* 1964; 142: 73-84. DOI: 10.1111/j.1469-7998.1964.tb05155.x.

177. van Dongen JJ, Langerak AW, Bruggemann M, et al. Design and standardization of PCR primers and protocols for detection of clonal immunoglobulin and T-cell receptor gene recombinations in suspect lymphoproliferations: report of the BIOMED-2 Concerted Action BMH4- CT98-3936. *Leukemia* 2003; 17: 2257-2317. 2003/12/13. DOI: 10.1038/sj.leu.2403202.

178. Waldman JD and Wernly JA. Cyanotic congenital heart disease with decreased pulmonary blood flow in children. *Pediatric clinics of North America* 1999; 46: 385-404. 1999/04/28.

179. Ossa Galvis MM and Mendez MD. Cyanotic Heart Disease. *StatPearls*. Treasure Island (FL): StatPearls Publishing

StatPearls Publishing LLC., 2019.

180. Ceyran AB, Senol S, Guzelmeric F, et al. Effects of hypoxia and its relationship with apoptosis, stem cells, and angiogenesis on the thymus of children with congenital heart defects: a morphological and immunohistochemical study. *International journal of clinical and experimental pathology* 2015; 8: 8038-8047. 2015/09/05.

181. Clambey ET, McNamee EN, Westrich JA, et al. Hypoxiainducible factor-1 alpha-dependent induction of FoxP3 drives regulatory T-cell abundance and function during inflammatory hypoxia of the mucosa. *Proc Natl Acad Sci U S A* 2012; 109: E2784-2793. 2012/09/19. DOI: 10.1073/pnas.1202366109.

182. Pfitzer C, Helm PC, Ferentzi H, et al. Changing prevalence of severe congenital heart disease: Results from the National Register for Congenital Heart Defects in Germany. *Congenital Heart Disease* 2017; 12: 787-793. DOI: 10.1111/chd.12515.

183. Samanek M. Boy:girl ratio in children born with different forms of cardiac malformation: a population-based study. *Pediatric cardiology* 1994; 15: 53-57. 1994/03/01. DOI: 10.1007/bf00817606.

184. Bianca S and Ettore G. Sex ratio imbalance in transposition of the great arteries and possible agricultural environmental risk factors. *Images in paediatric cardiology* 2001; 3: 10-14.

185. Beery AK and Zucker I. Sex bias in neuroscience and biomedical research. *Neuroscience and biobehavioral reviews* 2011; 35: 565- 572. 2010/07/14. DOI: 10.1016/j.neubiorev.2010.07.002.

186. Ji J, Sundquist J and Sundquist K. Gender-specific incidence of autoimmune diseases from national registers. *Journal of autoimmunity* 2016; 69: 102-106. 2016/03/21. DOI: 10.1016/j.jaut.2016.03.003.

187. Dragin N, Bismuth J, Cizeron-Clairac G, et al. Estrogenmediated downregulation of AIRE influences sexual dimorphism in autoimmune diseases. *The Journal of clinical investigation* 2016; 126: 1525- 1537. 2016/03/22. DOI: 10.1172/jci81894.

188. Moreira-Filho CA, Bando SY, Bertonha FB, et al. Minipuberty and Sexual Dimorphism in the Infant Human Thymus. *Scientific reports* 2018; 8. Article. DOI: 10.1038/s41598-018-31583-3.

189. Tomlinson C, Macintyre H, Dorrian CA, et al. Testosterone measurements in early infancy. *Archives of disease in childhood Fetal and neonatal edition* 2004; 89: F558-559. 2004/10/23. DOI: 10.1136/adc.2003.034017.