Diagnostics and pathophysiology

Anna Jeppsson

Department of Clinical Neuroscience Institute of Neuroscience and Physiology Sahlgrenska Academy, University of Gothenburg



UNIVERSITY OF GOTHENBURG

Gothenburg 2019

Cover illustration by Kerstin Lieberath

CSF biomarkers in idiopathic normal pressure hydrocephalus Diagnostics and pathophysiology © Anna Jeppsson 2019 anna.jeppsson@gu.se

ISBN 978-91-7833-426-1 (PRINT) ISBN 978-91-7833-427-8 (PDF)

Printed in Gothenburg, Sweden 2019 Printed by BrandFactory For my mother. Because diagnosis makes a difference.

"The only reason for time is so that everything doesn't happen at once"

Albert Einstein

ABSTRACT

Idiopathic normal pressure hydrocephalus (iNPH) is a disease of the elderly with enlarged ventricles despite a normal CSF pressure. Clinically, iNPH presents with gait- and balance disturbances, cognitive decline and incontinence. As the symptoms are reversed by shunt surgery, precise diagnostics is of essence. As of today, the etiology of the disease is largely unknown and specific diagnostic and prognostic tests are lacking.

The overall aim of this thesis project was to explore the diagnostic and prognostic potential of CSF biomarkers in iNPH. By measuring markers reflecting different pathophysiological aspects, we also wanted to elucidate underlying pathophysiologic mechanisms of iNPH.

In *paper I*, we showed that NFL was elevated and amyloid precursor protein (APP)-derived proteins and tau proteins were lower in patients with iNPH than in healthy individuals (HI). Post-surgery, there was an increase of NFL, APP-derived proteins, p-tau, and albumin in ventricular CSF, whereas levels of MBP and T-tau decreased. In *paper II* the concentrations of all soluble forms of APP, all Aβ isoforms and APL1β28 were lower, whilst APL1β25 and APL1\u00b327 were higher in CSF of iNPH patients compared to HI. No difference could be seen in biomarker concentrations between patients who improved after surgery and those who did not. In paper III, iNPH patients had lower concentrations of tau and APP-derived proteins in combination with elevated MCP-1 compared to HI and the most important differential diagnostic disorders. A prediction algorithm consisting of T-tau, Aβ40 and MCP-1 was designed as a diagnostic tool showing high discriminating ability. In *paper IV* all soluble forms of APP and all $A\beta$ isoforms were lower in both subcortical small vessel disease (SSVD) and iNPH in comparison to HI, albeit with a more pronounced reduction in iNPH. INPH and SSVD had elevated concentrations of NFL, MBP and GFAP compared to HI.

Our findings indicate that patients with iNPH have a CSF biomarker profile that distinguishes them from HI of the same age as well as from their mimics. The profile is characterized by a downregulation of APP-proteins, CSF biomarkers reflecting destruction to the white matter and astrocyte activation but no substantial cortical damage. Analysis of CSF biomarkers may provide an important tool for diagnosing patients with iNPH.

Keywords: Idiopathic normal pressure hydrocephalus, cerebrospinal fluid, biomarkers

ISBN 978-91-7833-426-1 (PRINT) ISBN 978-91-7833-427-8 (PDF)

POPULÄRVETENSKAPLIG SAMMANFATTNING

Demenssjukdomar är ett växande problem såväl inom hälso- och sjukvården som för samhället i stort. De flesta demenssjukdomar är idag obotliga eller har mycket begränsad möjlighet till behandling. Normaltryckshydrocephalus (NPH), som ger drabbade patienter gång- och balanssvårigheter, kognitiv nedsättning och inkontinens, kan betraktas som ett demenstillstånd hos äldre där förloppet är potentiellt reversibelt. Patienterna har ökad mängd ryggvätska (hydrocephalus = vattenskalle) och kan behandlas genom insättandet av en shuntslang från hjärnans vätskefyllda hålrum till (vanligtvis) bukhålan, där överskottsvätskan kan tas upp av kroppen. NPH kan ibland förklaras av patientens sjukdomshistoria men en stor del uppkommer utan någon känd orsak, och benämns då idiopatisk NPH (iNPH). Hos den äldre befolkningen är iNPH vanligare än vad statistiken antyder och andelen som kommer till diagnos och får behandling med shunt är låg.

I denna avhandling har vi undersökt proteiner (= äggviteämnen) i ryggvätskan hos patienter med iNPH. Genom att studera dessa ville vi öka precisionen i diagnostiken och öka kunskapen om sjukdomsmekanismer för iNPH.

Vi har funnit att proteinerna i ryggvätskan karaktäriseras av ett specifikt mönster bestående av lägre halter av amyloid- och tauproteiner och ökning av vissa proteiner som speglar påverkan på hjärnans vita substans. Vi tror att detta kan förklaras av att den ökande mängden vätska bidrar till en försämrad cirkulation i hjärnvävnaden och som en följd av detta till en minskning av dessa proteiner. Det vita substansen och hjärnans stödceller är påverkade men hjärnbarken är enligt våra resultat inte påverkad i någon större grad. Vi tror att påverkan på hjärnans små kärl till viss del liknar den vid andra så kallade "sub-kortikala" sjukdomar och detta pekar mot att det kanske finns fler individer som skulle kunna hjälpas av en shuntoperation än de som opereras idag. Proteinmönstret hjälper oss att skilja iNPH patienter från friska äldre och även från de viktigaste sjukdomarna som kan likna symptombilden vid iNPH och försvåra diagnostiken.

Det är vår förhoppning att resultaten kommer att bidra med nya pusselbitar för att förstå sjukdomsprocesserna vid iNPH och att denna kunskap kan hjälpa fler patienter till en säkrare diagnos, liksom till potentiell symptomlindring genom kirurgi.

LIST OF PAPERS

This thesis is based on the following studies, referred to in the text by their Roman numerals.

I. Jeppsson, A, Zetterberg, H, Blennow, K, Wikkelsø, C.

Idiopathic normal-pressure hydrocephalus- Pathophysiology and diagnosis by CSF biomarkers.

Neurology 2013;80:1385-1392.

II. Jeppsson A, Holtta M, Zetterberg H, Blennow K, Wikkelsø C, Tullberg M.

Amyloid mis-metabolism in idiopathic normal pressure hydrocephalus. Fluids Barriers CNS 2016;13:13.

III. Jeppsson, A, Wikkelsø, C, Blennow, K, Zetterberg, H, Constantinescu, R, Remes A M, Herukka, S-K, Rauramaa, T Nägga, K, Leinonen, V, Tullberg, M.

CSF biomarkers distinguish idiopathic normal pressure hydrocephalus from its mimics.

Accepted for publication in Journal of Neurology, Neurosurgery & Psychiatry.

IV. Jeppsson, A, Bjerke, M, Hellström, P, Blennow, K, Zetterberg, H, Kettunen, P, Wikkelsø, C, Wallin, A, Tullberg, M.

> CSF biomarkers highlight pathophysiological similarities and differences in idiopathic normal pressure hydrocephalus and subcortical small vessel disease.

Manuscript.

CONTENT

| A | BB | REVIA | .TIONS | XI |
|---|------------|----------------------|---|----------------|
| 1 | I | NTROI | DUCTION | 1 |
| 2 | I | DIOPA | THIC NORMAL PRESSURE HYDROCEPHALUS | 3 |
| | 2.1 | Diag | gnosis | 3 |
| | | 2.1.1 | Gait | 5 |
| | | 2.1.2 | Cognition | 6 |
| | | 2.1.3 | Incontinence | 8 |
| | | 2.1.4 | Other symptoms associated with iNPH | 8 |
| | 2.2 | Prec | diction | 9 |
| 3 | C | CSF in | HEALTH AND INPH | 11 |
| | 3.1 | Nev | w views on CSF and its circulation | 14 |
| 4 | C | SF BI | OMARKERS | 16 |
| | 4.1 | Am | yloid precursor protein-derived proteins and their homologues | 17 |
| | 4.2 | Tau | -proteins | 20 |
| | 4.3 | Bior | markers of white matter damage | 21 |
| | 4.4 | Infl | ammation and activation | |
| 5 | A | AIMS | | 25 |
| 6 | Ν | (IETHO | DDS AND STUDY DESIGN | 27 |
| | 6.1 | INP | PH patient cohort | 27 |
| | | 6.1.1 | Staging of severity; the iNPH scale | |
| | 6.2 | Bio | chemical analysis | 31 |
| | | 6.2.1 | CSF sampling | |
| | | | | |
| | | 6.2.2 | Analytical methods | |
| | 6.3 | 6.2.2 Rad | Analytical methods liological evaluation | 32 35 |
| | 6.3 6.4 | 6.2.2 Rad Stat | Analytical methods liological evaluation istical analysis | 32 35 36 |

| | 6.5.1 | Study I | 39 | |
|------------------|--------|---|----|--|
| | 6.5.2 | Study II | 42 | |
| | 6.5.3 | Study III | 45 | |
| | 6.5.4 | Study IV | 49 | |
| | 6.5.5 | Overlap | 50 | |
| 7 Results | | | | |
| 7.1 | l Sepa | arating iNPH from healthy individuals with CSF biomarkers | 53 | |
| 7.2 | 2 The | differential diagnostic capacity of CSF biomarkers | 60 | |
| 7.3 | 3 CSF | biomarkers in ventricular CSF | 65 | |
| 7.4 | 4 Prec | licting shunt response by CSF biomarkers | 69 | |
| 7.5 | 5 Rad | iological white matter changes | 70 | |
| 8 I | Discus | SION | 73 | |
| 8.1 | l Am | yloids in iNPH | 73 | |
| 8.2 | 2 Cor | tical pathology in iNPH? | 77 | |
| 8.3 | 3 Dan | nage to white matter and glia activation | 79 | |
| 8.4 | 4 Vas | cular changes in iNPH | 81 | |
| 8.5 | 5 Prec | licting outcome? | 83 | |
| 8.0 | 6 Can | we use CSF biomarkers to diagnose iNPH? | 84 | |
| 8.7 | 7 Gen | eral methodological considerations | 85 | |
| 9 (| Concl | USIONS AND FUTURE PERSPECTIVES | 87 | |
| ACKNOWLEDGEMENTS | | | | |
| Ref | ERENC | CES | 95 | |

ABBREVIATIONS

| Ab | Antibody |
|--------|--|
| ACG | Anterior cingulate gyrus |
| AD | Alzheimer's disease |
| AED | Astheno-emotional disorder |
| ANOVA | Analysis of variance |
| APLP1 | Amyloid precursor like protein 1 |
| APP | Amyloid precursor protein |
| AQP-4 | Aquaporin-4 |
| ARWMC | Age related white matter changes |
| AUC | Area under the curve |
| Аβ | Amyloid beta |
| BACE1 | β-site APP cleaving enzyme |
| BBB | Blood-brain-barrier |
| BD | Binswangers disease |
| BPH | Benign prostatic hyperplasia |
| CNS | Central nervous system |
| CV | Coefficient of variation |
| CSF | Cerebrospinal fluid |
| CSF TT | CSF tap test |
| CSF-OP | CSF opening pressure |
| CT | Computed tomography |
| DSI | Disease state index |
| ECF | Extracellular fluid |
| ECM | Extracellular matrix |
| ECS | Extracellular space of the brain |
| EI | Evans index |
| ELD | External lumbar drainage |
| ELISA | Enzyme-linked immunosorbent assay |
| EMD | Emotional-motivational blunting disorder |
| FLAIR | Fluid-attenuated inversion recovery |
| | |

| FTLD | Fronto-temporal lobar degeneration |
|-------|------------------------------------|
| GFAP | Glial fibrillary acidic protein |
| HI | Healthy individuals |
| IL | Interleukin |
| iNPH | Idiopathic NPH |
| ISF | Interstitial fluid |
| IQR | Interquartile range |
| LBD | Lewy-body dementia |
| LCSF | Lumbar CSF |
| LLOQ | Lower limit of quantification |
| LRG | Leucine-rich a2-glycoprotein |
| LTP | Long-term potentiation |
| MAb | Monoclonal antibody |
| MBP | Myelin basic protein |
| MCI | Mild cognitive impairment |
| MCP-1 | Monocyte chemoattractant protein 1 |
| MMP | Matrix metalloproteinase |
| MMSE | Mini mental state examination |
| MRI | Magnetic resonance imaging |
| MSA | Multiple systems atrophy |
| NFH | Neurofilament heavy chain |
| NFL | Neurofilament light chain |
| NFM | Neurofilament medium chain |
| NPH | Normal pressure hydrocephalus |
| OAB | Overactive bladder |
| PAG | Periaqueductal grey |
| PD | Parkinson´s disease |
| PD | Proton density |
| PDD | Parkinson's disease with dementia |
| PET | Positron emission tomography |
| PFC | Prefrontal cortex |
| PMC | Pontine micturition centre |
| PSP | Progressive supranuclear palsy |
| RAS | Reticular activation system |
| | |

| RAVLT | Rey auditory verbal learning test |
|-------|---|
| RCG | Rostro-caudal gradient |
| SAE | Subcortical arteriosclerotic encephalopathy |
| sAPP | Soluble amyloid precursor protein |
| SAS | Subarachnoid space |
| sNPH | Secondary NPH |
| SSCD | Somnolence-sopor-coma disorder |
| SSVD | Subcortical small vessel disease |
| SVD | Subcortical vascular disease |
| TIMP | Tissue inhibitor of metalloproteinases |
| ULOQ | Upper limit of quantification |
| VA | Ventriculo-atrial |
| VAD | Vascular dementia |
| VCSF | Ventricular CSF |
| WMC | White matter changes |
| VRS | Virchow-Robin spaces |
| VP | Ventriculo-peritoneal |

1 INTRODUCTION

In 2015, it was estimated that 47.5 million people suffered from dementia worldwide, and numbers are thought to double every 20 years ¹. Regardless of numbers, each case of dementia is a burden not only to society, but a psychological and social burden to families, friends and not least to the afflicted person.

For adequate prognosis, planning and exploration of treatment options, an exact diagnostic method of the diseases causing dementia is of essence. Additionally, long term planning in terms of assistance from caregivers and society to the individual suffering from neurodegenerative disorders could improve the daily life of patients and caregivers.

Among the dementias, there are a few that are regarded as "reversible", including normal pressure hydrocephalus (NPH)². NPH is a condition of the elderly with enlarged ventricles despite a normal CSF pressure. Clinically, the characteristic symptoms of NPH consist of gait disturbances, impaired balance, cognitive deterioration and incontinence, sometimes referred to as Hakims triad ²⁻⁴.

NPH has been known as a clinical syndrome since the neurosurgeon Salomón Hakim identified it in 1957 at Hospital San Juan de Dios in Bogotá, Colombia ⁵. Not surprisingly, the finding was regarded with initial scepticism when Hakim showed that the symptoms of dementia could be reversed in hydrocephalic patients with normal CSF pressure, a phenomenon that was

1

previously thought restricted to dementia secondary to vitamin deficiencies and to endocrine disorders ²³.

Hydrocephalus is divided into communicating and non-communicating where in the latter there is a blockage of CSF flow and in general a high pressure in the CSF. Herein I will focus on communicating hydrocephalus with a "normal" pressure (NPH).

NPH is classified as either secondary (sNPH) if there is a known cause, or idiopathic (iNPH). The secondary forms are seen following various kinds of brain trauma, subarachnoid haemorrhage, meningitis or stroke ⁶. The idiopathic form is more elusive, with no definite aetiology being found as of today.

The aetiology of iNPH remains an enigma. We know that it is underdiagnosed and under-treated ⁷ but that the vast majority of cases of iNPH are improved by shunt surgery ⁸. The focus of this thesis is to elaborate on how CSF biomarkers can aid in finding and diagnosing the patients that suffer from this disorder and by studying the biomarkers, helping us to understand a bit more of the pathophysiology at work.

2 IDIOPATHIC NORMAL PRESSURE HYDROCEPHALUS

The prevalence of iNPH has been difficult to assess accurately and thus the numbers have varied ⁹⁻¹², perhaps due to that only a small minority of the patients are thought to be diagnosed and even fewer are being treated by shunt insertion ⁷. Population based studies have estimated the prevalence of iNPH to be as high as 5,9 % in the population of 80 years and higher ¹². Probably, only about 20% of patients with the diagnosis are treated, possibly attributed to poor knowledge of the disorder and its treatment options.

The only method for managing the hydrocephalic state being used today is inserting a shunt, usually a ventriculo-peritoneal, or a ventriculo-atrial shunt ¹³. Shunt surgery improves around 80 % of the patients ⁸. If not treated, the patients condition will deteriorate. They will still improve after surgery, albeit to a lesser degree than if they had been operated early. The delay means loss-of-function that cannot be restored ^{14 15}.

2.1 DIAGNOSIS

"The cardinal early features of normal-pressure hydrocephalus in our patients were a mild impairment of memory, slowness and paucity of thought and action, unsteadiness of gait and unwitting urinary continence. The symptomatology was unobtrusive, having no assignable date of onset, and evolved over a period of weeks or a few months"³.

To diagnose iNPH, evidence are collected from clinical history, physical examination, and brain imaging. There are two different set of guidelines for diagnosing iNPH and also the procedure to diagnose according to these guidelines varies between centres ¹⁶ ¹⁷. In this thesis, the International guidelines will be used.

The *clinical history* should focus on the mode of onset (insidious), its temporal course (progressive) and severity of symptoms. Diagnosing iNPH is further dependant on that no known factor, such as previous head-trauma, meningitis or intra-cerebral haemorrhage is explanatory of the condition (in that case, the term would be secondary NPH, sNPH). A close examination of other medical conditions is also of great importance since there are a number of diseases of the elderly that can easily be misinterpreted for iNPH. The presence of incontinence as well as its type and extent, should be explored.

In the *physical examination*, gait and balance should be tested. To diagnose iNPH, at least gait/balance disturbance should be present, accompanied by either impairment of cognition, or incontinence, or both. Retropulsion is often seen, either spontaneous or provoked. Cognitively, the patients are usually showing a slowing of thought, inattentiveness, apathy, and encoding and recall problems.

Using *brain imaging* (usually Magnetic resonance imaging, MRI), ventricular size can be measured. Evans index (EI) $\geq 0,3$ is used as a cutting point for an increase in ventricular size as compared to cerebral matter ⁹. EI is calculated by the maximum width of the frontal horns divided by the maximum inner width of the skull ⁹. Further, imaging is used to secure that the aqueduct is

open (to rule out a non-communicating hydrocephalus) and to estimate the level of cortical atrophy. Other radiological biomarkers have been put forth, such as disproportionately enlarged subarachnoid space hydrocephalus (DESH) but there is no consensus on the application in diagnosis and prediction ¹⁸.

The lumbar CSF opening pressure (CSF-OP) should be measured and be within 5-18 mm Hg or 70-245 mm H₂O.

Clinically, there are a number of potentially difficult differential diagnostic challenges. The gait pattern in iNPH can be misinterpreted for, or affected by, Parkinson's disease (PD) (including atypical parkinsonian syndromes), arthritis of the joints, and polyneuropathy of different aetiologies. The affected cognition can sometimes be misinterpreted for other forms of neurodegenerative diseases, such as Alzheimer's disease (AD), subcortical vascular dementia (SVD), Parkinson's disease with dementia (PDD), Lewy body disease (LBD), other dementias or depressive disorders. Urinary incontinence can also be present in other neurological conditions such as post stroke but also as manifestations of primary urological disorders

2.1.1 GAIT

"The mechanism that allows a 6 foot tall human to walk on his two hind legs is imperfect but the nature of the imperfection has yet to be identified"⁴.

The gait disturbance is usually the first symptom to become evident and it is often referred to as gait apraxia ⁴. The hydrocephalic gait is characterised by

hypokinetic movement which in turn is composed of reduced stride length (albeit with greater variation), reduction of foot-to-floor clearance (due to insufficient extension of the knee) with a tendency to strike the ground flat whilst walking, and a "*disturbance of the dynamic equilibrium*" ¹⁹. This latter component is evident by an enhanced step width and an outward rotation of the feet. The patients also lose balance whilst turning. The interstep variation is diminished, leading to inability to compensate for body sway. Slight reduction of the arm and trunk movement during walking is seen ^{19 20}. The gait has been described as being "glued to the floor" ^{3 20} or as "magnetic" ¹⁶ since the foot clearance is extremely low. Interestingly, these problems are restricted to the elevated patient. When in bed, normal limb movement is seen ⁴. The gait is worsened as the symptoms progress in time, leading to the need of a wheelchair and eventually to immobility as truncal apraxia develops.

The gait disturbances are thought to be partially explained by impaired balance. The inability to compensate for body sways, was in Fisher's view attributed to *"a slowness in correcting a potential instability"*⁴. A possible distortion of visual input (visual axis), leading to a fast movement backwards, as if the body compensates for a fall forward has been suggested ²¹⁻²³.

2.1.2 COGNITION

The patients show a slowing of the mind and are often seen as lacking initiative and as indifferent, what Fisher termed the *"abulic trait"*⁴. These traits are associated with as well subcortical as frontal types of dementia, suggesting a pattern of *"fronto-subcortical dementia"*²⁴. Some evidence points to that

the fronto-subcortical deficit is manifested early in the process and in time becomes more of a global cognitive impairment as the syndrome progresses, highlighting the importance of an early diagnose ²⁵. Hellström et al have reported that iNPH patients seem more impaired in the fields of mental speed and executive functioning than actual memory disturbances ²⁶. The cognitive symptoms are preferably examined using neuropsychological testing ²⁷.

Using organic psychiatry classification ²⁸, iNPH patients initially suffer from astheno-emotional disorder (AED) a condition that is characterised by difficulties regarding concentration and memory, fatigue, irritability and/or emotional lability. As the disease progresses, emotional-motivational blunting disorder (EMD) (with apathy, emotional indifference and a lack of drive) develops, and might lead to, or coexist with, somnolence-sopor-coma disorder (SSCD) with impaired wakefulness, general slowing and dampening of cognitive, emotional, conative and motor processes ²⁹. Following surgery, the inverse order of symptom recovery is seen, and the latter responding the most favourable to the procedure ^{30 31}. The symptoms of SSCD have been linked to the ascending reticular activation system (RAS) ³¹.

Cognitive improvements are seen following surgery, especially in the most severely demented group ²⁵, even though some evidence supports that iNPH patients still do not match healthy individuals of the same age ^{25 26 30}. Vascular comorbidity has been shown to worsen the cognitive performance ³² but the magnitude of improvement following surgery is not affected by vascular comorbidity ²⁶.

2.1.3 INCONTINENCE

The neurological mechanism for the incontinence in NPH patients is thought of as an "uninhibited neurogenic bladder" ³³. This means that the usual central descending inhibition of the primitive reflex of contraction of the detrusor muscle is inhibited, leading the muscle to contract prematurely, resulting in urgency and frequent voiding.

Sakakibara et al ³⁴, examining in detail the bladder dysfunction in iNPH found that storage symptoms were more prominent than voiding symptoms. More specifically, urinary urgency, nocturnal frequency, urgency incontinence, diurnal frequency, retardation when initiating urination, prolongation/poor flow, sensation of post-void residual, straining, and intermittency was seen. The authors argue that overactive bladder (OAB) is probably the initial manifestation of urinary dysfunction symptom in iNPH.

The incontinence is not always recognised by the patient, especially in advanced stages, which can be an indicator of a frontal executive dysfunction ¹⁶.

2.1.4 OTHER SYMPTOMS ASSOCIATED WITH INPH

Other symptoms frequently occur in iNPH patients. Among those are impaired wakefulness and an increased need of sleep (also a part of the symptoms in SSCD) which as previously reported responds well to shunt treatment ³¹. Moreover, paratonic rigidity, retropulsion, cerebellar signs and focal neurological signs are seen ^{21-23 35 36}.

2.2 PREDICTION

Up to 80 % of patients improve by shunt-surgery ⁸ but there are still patients that do not respond to shunt treatment for reasons unknown. Being able to predict which individual patient that would not benefit from shunt placement would also mean that these patients could be spared the risk of brain surgery. Therefore, finding ways to choose the right patients for shunt-placement for iNPH has been the Holy Grail of iNPH research.

So far, the quest is quite disappointing. We do know that comorbidities, including heavy vascular co-morbidity do not mean that patients would not respond to shunt-placement ^{37 38}. We also know that patients with a long-standing symptomatology still respond to treatment ¹⁵.

To date, the method with the best sensitivity as to predict favourable outcome of surgery is assessing the clinical response to removal of CSF ³⁹⁻⁴². This is performed by the Tap-test (TT) or External lumbar CSF drainage (ELD). However, even if these tests can aid in the inclusion of patients eligible for surgery, a negative test does not exclude the possibility that patients still can benefit from shunt surgery. The sensitivity of the test for successful outcome of shunt surgery is around 75-92 % for the TT and 80-100 % for the ELD. The specificity is however 26-61 % for the TT and 50-100 % for the ELD ⁴³.

Therefore, these supplementary tests can aid in the inclusion of patients for shunt surgery but cannot be used for exclusion.

There is a need for improvement of additional diagnostic tests for iNPH. The problem with the poor specificity in the tests used for prediction is that it will lead to under-diagnostics of patients that suffer from the disorder and that could benefit from shunt surgery.

3 CSF IN HEALTH AND INPH

Traditionally, CSF is thought to be mainly produced in the choroid plexus located in the ventricles, although some amount is thought to be produced in the brain parenchyma ⁴⁴. The choroid plexus is comprised of numerous villi protruding into the ventricles, lined with cuboidal epithelium. Beneath the epithelium, there are plentiful of arteries.

The CSF is said to derive from an ultrafiltrate of plasma and is (in healthy individuals) produced at a rate of approximately 0.34 ml/min or approximately 500 ml/day ⁴⁵. The formation rate of CSF has been shown to be relatively indifferent of CSF pressure ⁴⁶. In iNPH, the rate of CSF production is in the same range or slightly reduced in comparison to HI ⁴⁷. The total volume of the CSF in HI is reported at about 250 ml ⁴⁸ of which about 80 ml is held in the spinal canal ⁴⁹.

From its production site in the lateral ventricles, the CSF is said to flow through foramen Monroe and enters the third ventricle. From there, it enters the fourth ventricle via the aqueduct. From the fourth ventricle, the CSF enters the subarachnoid space (SAS) via the two lateral foramina of Luschka and the central foramen of Magendie ⁵⁰ (Fig 1).

Conventionally, CSF is said to have its primary absorption site in the superior sagittal sinus, through the arachnoid villi, but there are also other routes of absorption, such as spinal absorption ⁵¹ as well as along blood vessels and cranial nerves ⁵².

Hakim explained the nature of iNPH by referring to the law of Pascal which states that "the pressure applied to an enclosed fluid, is transmitted undiminished to every portion of the fluid and to the walls of the containing vessel". Applying this law, he argued that the fluid exerts a greater force on the ventricular walls despite the normal pressure in an enlarged ventricular system naming it the hydraulic press effect. Therefore, he argued, initially there probably had been a ventricular dilation secondary to an increased pressure but once the ventricles had been dilated, they were being held enlarged by the fact that a greater strain was being applied despite the pressure being normal ²³. As the production rate of CSF is within the same range as of HI⁴⁷, it is hypothesized that decreased CSF absorption could explain the excess amount of CSF in iNPH ⁵³. There seems to be a trans-capillary absorption presumably as a response to inadequate outflow 54. In secondary cases (sNPH) there is a possible explanation for the reduced absorption in terms of previous bleeding to the SAS, and immune activation with inflammation. In iNPH, if the cause is impairment in outflow, the aetiology of the blockage is unknown.



Figure 1. An overview of the traditional view of the CSF circulatory system

The CSF is thought to represent the fluid microenvironment of the extracellular space of the brain (ECS) as CSF is said to lies in direct contact with the ECS ⁴⁴. The extracellular /interstitial fluid (ECF/ ISF) is traditionally said to communicate with the CSF via periventricular (Virchow-Robin) spaces and the exchange is mediated through gap-junctions in the Pia and ependyma. The spaces are said to be in dynamic equilibrium ^{45 46}. By its contact, CSF regulates the composition of the ECF, providing it with nutrients, serves to clear metabolic waste from the interstitium and serves as a medium for chemical signalling within the brain. The CSF's purpose is further to serve as a shock absorbent fluid, protecting the brain floating in CSF from rubbing against the cranium and contributes to the regulation of intracranial pressure.

3.1 NEW VIEWS ON CSF AND ITS CIRCULATION

In recent years, much interest has been given to re-evaluating many of the traditional views on CSF, its production, flow pattern and absorption ⁵⁵. These new thoughts have influenced how to think about the CSF biomarkers ability to truly reflect direct parenchymal processes.

The glymphatic system is thought to be the brain's version of a solution to waste clearance, an analogy to the lymphatic system in the rest of the body and its proposed role is to regulate CSF-ISF interchange ⁵⁶. Its name derives from *glia* and *lymphatic* to indicate the importance of the astroglia cells for this system.

In this model, subarachnoid CSF recirculates through the brain parenchyma via paravascular spaces. The CSF flows via the Virchow-Robin spaces (VRS) surrounding the penetrating arterioles (extensions of the pial arteries) and is transported by bulk flow through the parenchyma to peri-venous spaces ⁵⁶. Newly discovered lymphatic vessels seem to surround the dural sinuses and drain into deep cervical lymph nodes and is a possible missing link in the understanding of the brains immune-surveillance system ⁵⁷. These lymphatic vessels have also been visualized in meninges in humans in vivo ⁵⁸.

The water-transporting capability of the astrocytes is cardinal to the glymphatic system. The astrocytic foot processes cover the microvasculature ⁵⁹. Polarized Aquaporin-4 (Aqp4) channels in the astrocyte membrane are thought to facilitate water in- and outflow of the parenchyma and hence the

CSF-ISF interchange ⁶⁰ ⁶¹. This interchange is thought to decline with advancing age, possibly as a result of loss of Aqp4 polarization surrounding the penetrating arteriole secondary to reactive astrogliosis ⁶².

Sleep is thought to increase CSF-ISF turnover by expanding the extracellular space, thus allowing more CSF to enter the parenchyma ⁶³. This sleep-induced change in ECS is believed to be mediated by extracellular ion concentrations ⁶⁴. As the ISF/CSF recirculates and mixes in the parenchyma, it is proposed to clear metabolic waste, including A β by washing and hence "clean" the ECS ⁵⁶.

In this view, the interchange of ISF and CSF would be more tightly regulated than previously thought. Also, the directed flow from the choroid plexus and eventually to the arachnoid granulations is challenged and is now thought to involve a more to- and fro pattern directed by arterial pulsatility and respiratory rate which might also effect the direct ISF/CSF exchange in the peri-capillary spaces ⁵⁵. Further, ISF production site is coming into question, and is now thought to be, at least to a large extent, a product of capillary secretion ⁵².

Taken together, the view of concentrations of CSF biomarkers in lumbar CSF representing concentrations in ISF may be challenged and this discussion will continue. Nevertheless, there is a communication between ISF and CSF and as such, it is generally accepted that CSF biomarkers can be used as a way to study pathophysiological dynamics in the brains parenchyma.

4 CSF BIOMARKERS

"A biomarker does not substitute for a brain" Martin Möckel

A biomarker is by definition "a characteristic that is objectively measured and evaluated as an indicator of normal biological processes, pathogenic processes, or pharmacologic responses to a therapeutic intervention" ⁶⁵.

CSF biomarkers are in this thesis viewed as a form of "chemical footprints" of on-going procedures in the parenchyma. They are used to extrapolate theories on cerebral physiology and pathophysiology since the content of fluid is thought to represent the condition of the microenvironment of the brain via the CSF ⁶⁶.

CSF biomarkers offer a tool to be used in clinical practice in the aid of diagnosing different diseases leading to dementia as well as other neurological disorders. Today, this method is widely used in the diagnosis of AD ⁶⁷ and is now a part of the diagnostic criteria ⁶⁸.

For iNPH there are several areas of potential use for CSF biomarkers. Being one of the few disorders causing dementia that is to a certain extent reversible, the need for precise diagnostic methods for iNPH is of essence. Herein lies the need for biomarkers that are able to differentiate between iNPH and other types of dementia in the clinical setting and to provide a more solid foundation when planning for health care and social support for patients suffering from iNPH. There is also a need to understand the underlying pathophysiologic mechanism, as well as the dynamic, reversible nature of the syndrome and its relation to symptomatology. Furthermore, there is an ongoing search for biochemical markers that could predict outcome of shunting in iNPH. Some progress has been made, but no marker has so far showed enough sensitivity and specificity to be of practical use in selection of candidates eligible for operation ^{13 69-78}.

4.1 AMYLOID PRECURSOR PROTEIN-DERIVED PROTEINS AND THEIR HOMOLOGUES

In these studies, amyloid metabolism is studied using the derivates of Amyloid precursor protein (APP), soluble APP alfa and -beta (sAPP α , sAPP β) and Amyloid β (A β)-fragments of different lengths, (A β -38, A β -40 and A β -42).

APP is a large, transmembrane protein ^{79 80}. It has a large extracellular domain and a small cytoplasmic tail. Full-length APP is cleaved by α - (ADAM10) or β -secretase (BACE1) ⁸¹ generating sAPP α ("the non-amyloidogenic pathway") and sAPP β ("the amyloidogenic pathway") ^{82 83} respectively. Following α - and β -cleavage, intramembranous proteinolysis by γ -secretase generates A β -fragments of varying lengths from sAPP β (A β -X, with the number X corresponding to the number of amino acids in the fragment) and p3 from sAPP α ⁸³⁻⁸⁶.



Figure 2. Enzymatic cleavage of APP in the amylogenic pathway is initiated when β -secretase cleaves the ectodomain of the transmembrane protein. Subsequently, γ -secretase generates $A\beta$ of different lengths by intramembranous proteinolysis. Illustration by Jomi Jutlöv.

The amyloid hypothesis of AD states that an early, or initiating, event in AD is the alteration of A β metabolism ⁸⁷. An absolute or relative increase in the hydrophobic A β species A β 42 and -43 (by increased production or reduced clearance) leads to the formation of amyloid plaques that are being deposited in the brain parenchyma. The main component of the amyloid plaques in AD is A β 42 ⁸⁸. Lowered levels of A β 42, or a reduced A β 42/40 ratio in CSF are explained by the plaque deposits ^{87 89} which is supported from studies of APP-transgenic mice where there was an inverse relationship between plaque-burden and A β levels in ISF, not explained by a reduction in APP-production rate ⁹⁰.

The burden of amyloid deposits increases linearly with age and is primarily located to precuneus, temporal cortex and anterior- and posterior cingulate ⁹¹. A β oligomers inhibit Hippocampal long-term potentiation (LTP) in vivo and damage synaptic structures ⁸³ ⁹². It is thought that it is rather the soluble oligomers (released from the plaques), not the plaques per se, that are synaptotoxic ⁹². The cognitive decline in AD is not very well correlated to the amount of plaques but it is hypothesized that amyloid deposits may lead to downstream phenomena (such as activation of the innate immune system and the formation of tangles) leading to neuronal dysfunction in AD ⁹².

The physiological role of the evolutionary conserved APP-family is not yet fully understood. There are strong indications that processing of APP is important during brain development, synaptic functioning and dendritic formation. Both sAPP α , and $-\beta$ seem to be important to synapse formation and might act as signalling molecules regulating neuronal growth and interaction ⁹³. Accumulating evidence suggests that sAPP α has a neuroprotective role and is important for synaptic plasticity, learning and memory (although the main receptor target is not known). One possible mechanism is by regulating NMDA receptor function, and thus LTP ^{94.95}. α -secretase cleaving, and subsequent increased level of sAPP α is increased by neuronal activity ⁷⁹.

In rats with kaolin induced hydrocephalus, $A\beta$ has been shown to accumulate possibly as an effect of down regulation of LRP-1, the main efflux transporter of $A\beta$ over the blood-brain-barrier (BBB) ^{96 97}. The theory has been put forth that NPH and AD share a common pathophysiological aetiology in that

reduced clearance of A β would lead to AD-like pathology in iNPH-patients ^{97 98}.

The APP homologue APP- like protein 1 (APLP1) is cleaved by the same enzymatic machinery as APP resulting in the non-amyloidogenic APLP1 derivates APL1 β 25, -27, and -28. ⁹⁹⁻¹⁰². Being processed by the same enzymes although not aggregating in plaques, the ratio of APL1 β 28/ total APL1 β has been suggested as a marker for the relative production of A β 42 to total A β . ¹⁰⁰. The ratio APL1 β 28/ total APL1 β has been shown to increase in patients with AD, lending support to the notion of increased ratio of A β 42/total AB as an underlying mechanism of AD ¹⁰³.

4.2 TAU-PROTEINS

Tau binds to (mainly neuronal) microtubule, stabilising it and aiding its assembly of the protein ¹⁰⁴. In AD, tau becomes hyperphosphorylated, leading to microtubule instability and impaired axonal transport. Tangles, the other neuropathological hallmark of AD, has been shown to be made up mainly of fibrils containing aggregated hyperphosphorylated tau ⁸⁷. Tau is implicated in a large number of other neurodegenerative diseases ¹⁰⁵. In cortical neurodegenerative processes with axonal death, there is an outflow of tau into the CSF. Thus the level of tau in CSF reflects the extent of the damage to cortical structures ^{87 106}.

4.3 BIOMARKERS OF WHITE MATTER DAMAGE

Neurofilament light (NFL) reflects large-calibre myelinated axonal damage ⁶⁹. More specifically, it is said to mirror the loss of intermediate filament protein that leaks through injured cell membranes of large, myelinated axons ^{70 106}. NFL has been used as a cerebrospinal fluid (CSF) biomarker reflecting neuronal death and axonal degeneration in several neurological diseases ¹⁰⁷ but NFL is now regarded as a more general marker of neuronal degeneration ¹⁰⁸. Disorders with mainly cortical engagement do not typically exhibit high concentrations of NFL ¹⁰⁹. Higher concentrations have been associated with disease progression and NFL has been suggested as a disease-intensity marker, rather than a marker of a specific aetiology ¹¹⁰.

Myelin basic protein (MBP) is a membrane protein of oligodendroglia and comprises 30-40 % of the myelin in the CNS. Oligodendroglia cells wrap membrane processes around neural axons in the CNS which highly increases the speed of nerve conduction velocities ¹¹¹. Presumably, elevated levels of MBP in the CSF is due to leakage of MBP from the periventricular white matter ¹¹² and elevated MBP in the CSF is an indicator of demyelination ^{45 70}.

4.4 INFLAMMATION AND ACTIVATION

Chemokines and cytokines are regulators of the inflammatory system and are released by activated astro- and microglia cells in response to various

inflammatory threats to the CNS, including misfolded extracellular proteins and damaged synapses ¹¹³. Activated microglia is seen in relation to amyloid deposits ¹¹⁴ but if the macrophage activity is aiding the recovery, or worsening the condition, is a matter of debate ¹¹⁵.

IL-8 is a chemo attractant, acting on neutrophils, but is also thought to act on migrating monocytes, together with MCP-1 contributing to firm adhesion of monocytes to vascular endothelium under flow conditions ¹¹⁶. IL-8 binds to CXCR1 and CXCR2. Links have been established between IL-8 and the development of atherosclerosis ¹¹⁶.

IL-10 is an anti-inflammatory cytokine. In mice it has been shown that lack of IL-10 leads to more severe atherosclerosis, whereas increased levels of IL-10 show opposite effect, as well as decreased recruitment of monocytes ¹¹⁷.

Monocyte chemoattractant protein I (MCP-1) binds to the CCP2 receptor on migrating monocytes, and is involved in diapedes and migration of monocytes ¹¹⁵ ¹¹⁷ ¹¹⁸. MCP-1 acts as a chemoattractant of astroglia ¹¹⁹ and is present in amyloid plaques, probably of microglia origin ¹¹⁹. It is further a known marker of peripheral tissue macrophages ¹²⁰ and also released from astro- and microglia in the CSF, facilitating the migration of macrophages ¹²¹.

Glial acidic fibrillary acidic protein (GFAP) is a protein synthesized in fibrillary astrocytes and increased concentrations is an indicator of acute damage to astroglial cells or a marker of astrogliosis ^{122 123}. The protein is the main component of the astroglial filament and the CSF concentrations increase with age ¹²³. YKL-40 is, in vivo, mostly associated with astrocytes and
is elevated in particular in diseases with CNS inflammatory origin but also in the healthy elderly. It seems as if elevated GFAP and YKL40 are indicative of reactive gliosis but more of acute, than in chronic stages of gliosis ¹²⁴.



Figure 3. Schematic illustration of the origin of some of proteins measured in this thesis. NFL is located in myelinated axons, MBP in the myelin sheath of oligodendroglia cells, MCP-1, GFAP and YKL-40 are found in astroglial and tau in cortical neurons. APP and amyloid- β are located in axon terminals. Illustration by Jomi Jutlöv.

5 AIMS

The overall aim of this thesis project is to explore the diagnostic and prognostic potential of CSF biomarkers in iNPH. By measuring markers reflecting different pathophysiological aspects, we aim to elucidate underlying pathophysiologic mechanisms of iNPH.

The specific aims for the different papers were:

- To explore the pathophysiology of iNPH by examining a broad spectrum of CSF biomarkers and evaluate the diagnostic value of the biomarkers chosen.
- II. To examine CSF concentrations of APLP1-derived peptides in iNPH, especially if the APL1β28 form was increased, and to explore the prognostic value of amyloid-related CSF biomarkers.
- III. To validate the differential diagnostic significance of CSF biomarkers reflecting amyloid cascade function, AD-related amyloid β (A β) production and aggregation, cortical neuronal damage, tau pathology, damage to long myelinated axons and astrocyte activation. All of which hypothetically separates iNPH from other common neurodegenerative disorders.
- IV. To specifically expand the knowledge of pathophysiological similarities and differences between iNPH and SSVD, with healthy controls as a reference group, using a broad panel of CSF biomarkers reflecting amyloid pathology, subcortical neuronal degeneration, myelin damage, astrogliosis and markers of extracellular matrix remodeling.

6 METHODS AND STUDY DESIGN

6.1 INPH PATIENT COHORT

The iNPH-patients were included and diagnosed at the Hydrocephalus Unit at Sahlgrenska University Hospital, the referral unit for Västra Götaland region/western Sweden. The diagnosis was made according to international guidelines ¹⁶. Diagnosis included symptom duration > 2 months, with gait problems gradually developing and mental disturbances probably attributed to iNPH. Incontinence and balance difficulties could be present. The clinical diagnosis was complemented with MRI findings (i.e. EI > 0,3, an open aqueduct and no other known cause of ventriculomegaly). All patients were clinically assessed by a neurologist who reviewed the patients' clinical history and performed a neurological exam. A physiotherapist assessed gait and balance and a neuropsychologist tested the subjects for cognitive deficits. In addition, an MRI was performed and images were evaluated by an experienced neuroradiologist. Severity of the disorder was staged using the iNPH scale, see below 27. As a part of the evaluation, all patients were subjected to lumbar puncture, where opening pressure was measured and 10 mL of CSF was collected. Samples were collected in the morning with the patient in a recumbent position.

Peri-ventricular changes, deep white matter changes and lacunar infarcts seen on MRI were evaluated. No patients showing signs of acute hydrocephalus (i.e. symptom duration < 2 month), inability to perform the tests needed for the study, restricted life-expectancy due to other causes (e.g. malignancies),

showing other medical contra-indications to surgery or opposing inclusion despite earlier approval were included in the studies.

All patients diagnosed with iNPH and accepting shunt surgery were operated upon and given a shunt with a Rickham reservoir and an anti-siphon device. In most cases, a ventriculo-peritoneal (VP) shunt was placed but there were cases where this was not possible because of technical difficulties (e.g. prior operations in the peritoneal cavity) and in these cases, a ventriculo-atrial shunt (VA) was offered.

At six months after surgery, the patients were subjected to the same clinical examinations and an MRI scan. In patients who did not show significant improvement, shunts were checked for patency and all shunts were functional at the follow-up examination.

All patients and healthy individuals or their next of kin gave their oral and written informed consent to participate in the studies. The Regional Ethical Review Board in Gothenburg/Sweden, Kuopio/Finland and Linköping/Sweden approved ethical permission for the studies.

6.1.1 STAGING OF SEVERITY; THE INPH SCALE

For disease staging and also to quantify severity of symptoms and improvement after surgery, the iNPH scale, developed at the Hydrocephalus Unit, was used in the studies ²⁷. The scale covers the domains of gait, balance, cognition and continence and uses both ordinal scales and continuous

measures. Gait is given double weight given that it is reported as the major complaint of patients with iNPH, as well as their caregivers ¹²⁵.

The *gait* domain is measured by letting the patients walk 10 meters in a free manner. Number of steps and time taken were recorded. The test is performed twice, using the most favourable result. Additionally, an ordinal scale for measuring gait was applied. 1 = Normal, 2 = Slight disturbance of tandem walk and turning, 3 = Wide based gait with sway, without foot corrections, 4 = Tendency to fall, with foot corrections, 5 = Walking with cane, 6 = Bi-manual support needed, 7 = Aided, 8 = Wheelchair bound. The three different tests were converted into scores, added and divided by three (or the number of tests performed), thus adding up to a domain score.

Balance was measured using an ordinal scale where; 1 = Able to stand independently for more than 30 sec on either lower extremity alone, 2 = Ableto stand independently for less than 30 sec on either lower extremity alone, 3 = Able to stand independently with the feet together for less than 30 seconds, 5 = Able to stand independently with the feet apart (1 foot length) for more than 30 seconds, 6 = Able to stand independently with the feet apart for less than 30 seconds, 7 = Unable to stand without assistance. The rating score was then converted into a domain score.

Neuropsychology was measured by the Grooved pegboard, the Rey Auditory Verbal Learning Test (RAVLT) and the Swedish Stroop test ²⁶. For measuring manual dexterity, the grooved pegboard was used. The test was performed twice and the fastest time recorded. Verbal learning and recall was measured by the RAVLT. A total of five trials were performed and the sum of the total

trials recorded and later converted into scores. In the Stroop test selective attention, cognitive flexibility and processing speed are measured. Two areas are given points; colour naming and an interference task. Scores are converted and the total four scores are added and divided by four (or by the number of tests performed), thus adding up to a domain score.

Continence was covered by an ordinal scale, where the rating is given by the most reliable source, due to this somewhat delicate nature. 1 = Normal, 2 = Urgency without incontinence, 3 = Infrequent incontinence without napkin, 4 = Frequent incontinence with napkin, 5 = Bladder incontinence, 6 = Bladder and bowel incontinence. The result was then converted into a domain score.

In all, the domain scores were added up (gait given double weight) and divided by 5 (or the number of domains available).

The resulting scale is a measurement of severity of the disease, ranging from 0-100 where 0 is the most severe state. When constructing the scale, it was designed with reference to a group of healthy elderly individuals and 100 can thereby be seen as representing normality. Within the scale, the score is to be seen in relation to other iNPH patients, thus reflecting the severity of the disease in relation to other patients with the same disease. To define improvement following surgery an increase in \geq 5 points on the scale was used.

6.2 BIOCHEMICAL ANALYSIS

All of the chemical analyses were made by methods based on antibody detection.

Immunoassays were used for most of the analyses. Briefly, the immunoassay method is based on quantification of the analyte using specific antibodies. One antibody is coated in excess on a plate (capture antibody) in a well. Then, the sample is administered to the well, leading the analyte in the sample to react with the antibodies. After washing, another analyte-specific antibody (detection antibody) is administered and binds to a different epitope on the molecule, thus creating a complex (or a "sandwich") between the analyte and the two antibodies. The detection antibody will carry a label that allows for detection ¹²⁶⁻¹²⁸, either by an electrochemiluminescent plate-based assay or enzyme-linked immunosorbent assay (ELISA)

For some analysis, the XMap technology was used ¹²⁹. The multianalyte assay is developed as to be able to measure several analytes at once, thereby reducing the number of analyses and the amount of CSF needed. Monoclonal capture antibodies (Mab) specific for their epitope are constructed. Spectrally specific carboxylated beads are covalently coupled with the MAbs. A plate filled with several (in this case 96) wells are pre-washed. The beads are placed in wells together with biotinylated detector MAbs. CSF samples are applied and incubated over night. After washing, plates were read by Luminex 100, by flow cytometric separation of the different antibody-coated microspheres ^{129 130}

6.2.1 CSF SAMPLING

Lumbar CSF was obtained from the iNPH patients prior to surgery. All lumbar punctures were performed in the morning to avoid any influence on the result from possible diurnal fluctuations in biomarker levels. The lumbar puncture was made with the patient in the recumbent position.

Ventricular CSF was sampled through the catheter introduced in the right lateral ventricle at the time of shunt surgery. The first 2 mL of CSF were discarded and the next 8 mL were collected. Postoperative ventricular CSF was sampled at the postoperative re-examination through a puncture of the Rickham reservoir.

The CSF, collected in polypropylene tubes, was centrifuged at $2,000 \times \text{g}$ at room temperature for 10 min. The ensuing supernatant was aliquoted in screw-cap polypropylene tubes and stored at -80°C pending biochemical analyses.

6.2.2 ANALYTICAL METHODS

For these studies, the following analytical methods were used to determine CSF biomarker concentrations. For each study, all analyses were performed batch-wise in one round of experiments by board-certified laboratory technicians at the Clinical Neurochemistry Laboratory, Sahlgrenska University Hospital, Mölndal, Sweden. The laboratory technicians were blinded to clinical data In study I, II, and IV, NFL was measured by enzyme-linked immunosorbent assay (ELISA) technology using a commercial kit (UmanDiagnostics NFlight[®]) with a lower limit of detection of 50 ng/L as described in Norgren et al¹⁰⁹. In *study III*, NFL concentration was measured using an in house enzymelinked immunosorbent assay (ELISA) as previously described in Gaetani et al ¹³¹. In this method, monoclonal antibodies NfL21(coating) and NfL23 (detection), targeted at the core domain, are used. Lower limit of quantification (LLOQ) is 78 pg/mL and the upper limit of quantification (ULOQ) is 10,000 pg/mL. Coeficient of variation (CV) was below 13 %. The method from Uman Diagnostics and the in-house ELISA are strongly correlated (r = 0.9984, p < 0.001)¹³¹. The initial ELISA method for determining NFL was described by Rosengren et al. where polyclonal antisera were used ¹³². The method was later elaborated by Norgren et al with monoclonal antibodies which yielded higher sensitivity and specificity, no cross-reactivity with the NF-intermediate (NF-M) and heavy chain (NF-H) and the advantage of being able to establish a stable method using the monoclonal antibodies MAb 47:3 (coating) and MAb 2:1 (tracer).

The analysis of MBP (*I*, *IV*) was performed with an ELISA (Active[®] MBP, Diagnostic Systems Laboratories Inc., Webster, Texas, USA), according to the manufacturer's instructions.

CSF YKL-40 (II) concentration was measured by solid phase sandwich ELISA (R&D Systems, Inc., Minneapolis, Minnesota, USA) according to the manufacturer's instructions.

GFAP (*IV*) concentration was measured by an in-house sandwich ELISA method using antisera anti-GFAP IgG polyclonal antibodies from two species, rabbit anti-GFAP IgG and hen anti GFAP IgG. Goat anti-rabbit IgG was used as a detection antibody ¹²³.

Amyloid β isoforms (A β 38, A β 40, and A β 42), the sAPP isoforms (sAPP α and sAPP β) (*I*, *II*, *III* and *IV*) and the inflammatory markers IL-8, IL-10 (*I*) and MCP1 (*I*, *III*) were analyzed by electrochemiluminescence assays described by the kit manufacturer (Meso Scale Discovery, Gaithersburg, MD, USA) ¹³³. As for sAPP isoforms, the capture Ab for sAPP α is the Mab 6E10 and for sAPP β a neoepitope-specific antibody is used.

The APLP1-derived peptides APL1 β 25, APL1 β 27, and APL1 β 28 (*II*) were analyzed using a commercial ELISA (IBL International, Hamburg, Germany). The samples were analyzed according to the kit insert with minor modifications. The CSF samples were diluted 1:20 for APL1 β 25, 1:10 for APL1 β 27, and 1:5 for APL1 β 28 by the dilution buffer contained in the kit. All samples were analyzed in duplicate and CV for standards and samples was < 5 %.

CSF T-tau and P-tau (*I, III*) were measured with flow cytometry by the Luminex[®] xMAP[®] technology using the INNO-BIA AlzBio3 kit (Innogenetics, Ghent, Belgium), as previously described in detail in Olsson et al ¹²⁹. CV was below 10 %.

The concentrations of matrix metalloproteinase (MMP) -1, -2, -3, -9, -10 and tissue inhibitor of metalloproteinase 1 (TIMP1) (*IV*), were measured using single- or multiplex electrochemiluminescent ELISA (Meso Scale Discovery, Rockville, Maryland, USA), following the manufacturer's instructions with minor modifications. CV was below 15 % for all assays.

6.3 RADIOLOGICAL EVALUATION

In study II and IV, the extent of radiological white matter lesions in iNPH patients (II O(IV)) and patients with SSVD (IV) were staged according to the age-related white matter changes (ARWMC) scale ¹³⁴.

All patients had undergone radiological examination as a part of the diagnostic routine and the rating was performed on the images available. All iNPH patients had undergone MRI. In the SSVD group, patients had undergone MRI or CT. All radiological staging was made by the same observer (AJ).

The ARWMC scale is constructed to be able to be used for both computed tomography (CT) and MRI images. White matter change is defined as bright lesions ≥ 5 mm on T2, proton density (PD) or fluid attenuated inversion recovery (FLAIR) on MRI or hypodense areas of ≥ 5 mm on CT. Rating is made in five different domains: frontal, parieto-occipital, temporal, basal ganglia (striatum, globus pallidus, thalamus, internal/ external capsule and insula) and infratentorial/ cerebellum. In each region, the left and right

hemisphere is rated separately, giving a total of ten regions. In each region, the ARWMC is rated from 0 to 3. The scale is given in Table 1.

Table 1. The Age related white matter changes (ARWMC) scale.

| White matter lesions | |
|-----------------------|---|
| | |
| 0 | No lesions (including symmetrical, |
| | well-defined caps or bands) |
| 1 | Focal lesions |
| 2 | Beginning confluence of lesions |
| 3 | Diffuse involvement of the entire region, |
| | without involvement of U fibres |
| | |
| Basal ganglia lesions | |
| 0 | No lesions |
| 1 | 1 focal lesion (≥ 5 mm) |
| 2 | > 1 focal lesion |
| 3 | Confluent lesions |

6.4 STATISTICAL ANALYSIS

Due to non-symmetrical distribution of data, non-parametric statistics were used in most of the analysis (*I, II and IV*). Pairwise comparison was performed by the Wilcoxon Mann-Whitney U-test. The Kruskal Wallis test was used for

multiple comparisons. Changes between pre- and postoperative examinations and CSF concentrations were analyzed by the Wilcoxon signed rank test. For comparison of two proportions, the Fisher's exact test was used. For associations between two independent variables, the Spearman rank order correlation was chosen.

In *study III*, parametrical statistics was used to maximize the potential for constructing a combined predictive model. The One-way ANCOVA, corrected for age and sex, with Dunnett's multiple comparisons test was used to compare all groups to iNPH and HI. To construct the predictive model, univariable logistic regression analysis was performed for each individual CSF variable to separate iNPH vs non-iNPH disorders. Stepwise selection of the significant variables was used to select a multivariable logistic model and the chosen model was cross-validated. Area under ROC-curve (AUC-statistics) was calculated for description of goodness of models for iNPH vs HI, non-iNPH, cognitive disorders and movement disorders.

In all studies, significance tests were two-sided and alpha was set to p < 0.05. If not otherwise stated, no correction for the mass significance effect was made in order to avoid type II errors. Statistical analyses were made using IBM SPSS Statistics for Windows version 20 (*I*), 21 (*II*) and 25 (*III, IV*) (SPSS, Chicago, IL, USA), SAS Version 9 for Windows (SAS Institute, Cary, NC, USA) and GraphPad Prism[©] for Windows version 8.0.2. (GraphPad Software, La Jolla California USA, <u>www.graphpad.com</u>).

AJ performed the statistics in study *I*, *II* and *IV*. In study *III*, statistics were performed by Anders Pehrsson and Nils-Gunnar Pehrsson at Statistiska Konsultgruppen/ Gothenburg.

6.5 STUDY DESIGN AND PATIENT SELECTION

6.5.1 STUDY I

In study I, we included 27 patients with iNPH and 20 healthy elderly.

Patients were selected retrospectively, 15 men and 13 women, aged 57 to 79 and diagnosed according to standard protocol. All patients received a ventriculo-peritoneal shunt with a programmable valve with an anti-siphon device and a Rickham reservoir.

Lumbar CSF (LCSF) was obtained prior to surgery, at the time for clinical evaluation. Per-operative ventricular CSF (VCSFper) was sampled through the catheter introduced in the right lateral ventricle at the time of shunt surgery. The first 2 mL of CSF were discarded and the next 8 mL were collected. Postoperative ventricular CSF (VCSFpost) was sampled at the 6-month postoperative re-examination through a puncture of the Rickham reservoir.

Analyses for comparisons were made on previously gathered lumbar CSF samples from elderly healthy individuals, 11 men and 9 women ⁴⁹. These individuals were recruited from the population register of the City of Gothenburg and the Swedish retired people's organization, ages ranging from 64 to 76. Criteria of exclusion included neurological-, or psychiatric illnesses (including addiction of alcohol and drugs) or back- or spinal problems. All control subjects underwent neurological testing and blood tests measuring

liver- and kidney function, blood count, ions and blood sugar were performed, assuring that these tests came out within the normal range. None of the subjects chosen were treated with centrally working analgesics, or psychopharmacological drugs.

| pu | ile | |
|------|------|-----|
| - 0 | tart | |
| pre | rqu | |
| ng | nte | |
| agi | ıd i | |
|) st | ar | |
| 001 | lian | |
| 6 | пеа | |
| re (| I St | |
| sco | и с | |
| ule | give | |
| scc | is a | |
| ΡH | ng | |
| N | agi | |
| ols. | H SI | |
| ntrc | Π | |
| 00 | Π. | |
| put | snts | |
| Н | atie | |
| NP | d E | |
| ır i | Π | |
| e fi | r il | |
| elin | fo | |
| basi | эшс | |
| at l | utco | |
| se | 1 oi | |
| d a | anc | |
| an | ely | |
| Sex | ttiv | R) |
| 2. | era | δIJ |
| ble | top | lge |
| 3 | S | 1 |

| Table 2. Sex and age at be postoperatively and outcon range (IQR). | aseline for iNPH me for iNPH pa | l and controls. INPH tients. INPH staging | scale score (0-100) is given as median | staging pre- and and interquartile |
|--|------------------------------------|--|---|---------------------------------------|
| | Ŧ | INPH | | |
| | | Pre op | Post op | Outcome |
| | (n = 20) | (n = 27) | (n = 27) | (n = 27) |
| Female (n (%)) | 9 (46) | 13 (47) | | |
| Age (mean (SD)) | 70.6 (3.6) | 69.6 (6.6) | | |
| Gait | | 45 (31 to 82) | 77 (50 to 90) | 13 (3 to 31)*** |
| Neuropsychology | | 60 (42 to 77) | 80 (55 to87) | 8 (-2 to 15)** |
| Balance | | 67 (67 to 67) | 67 (67 to 83) | 0 (0 to16) NS |
| Continence | | 60 (0 to 100) | 80 (60 to 80) | 0 (0 to 20) NS |
| Total | | 59 (46 to 75) | 73 (58 to 84) | 13 (3 to 21)*** |
| ** p < 0.01, *** p < 0.001, N | <pre>IS; non-significal</pre> | nt. Significance calcul | ated by Man Whitney | / U test. |

6.5.2 STUDY II

For *study II*, we included 20 patients with iNPH and 20 neurologically healthy controls.

We selected 10 patients who improved substantially from surgery and 10 who did not. All patients were diagnosed with iNPH according to standard protocol, underwent surgery with installment of a ventriculo-peritoneal shunt with a programmable valve with an anti-siphon device and a Rickham reservoir.

Patients were selected retrospectively from our local database that at the time included 176 patients that had full pre- and postoperative iNPH scale scores and had sufficient amount of frozen CSF samples available. From that material, we selected the 10 patients that benefitted the most (defined as "total" outcome on the iNPH scale). In patients that did not improve (defined as < 5 p improvement in the iNPH scale at post-operative exam), medical records were scanned in order to establish that the shunts were functional at the time of re-evaluation. None had complications or other conditions that could explain that improvement was not reached. In that group, the 10 patients that benefitted the least were selected. Vascular risk factors were documented for all patients and white matter changes were scored using the age related white matter changes ARWMC-scale ¹³⁴.

As for controls, we selected twenty persons undergoing knee-surgery and that had given their consent to CSF sampling in conjunction with receiving spinal anesthesia. All controls had a normal mini mental state examination (MMSE) score and had a normal neurological status.

| | Pre op | | Post op | | Outcome | |
|------------------------|------------------|------------------------|--------------------|--------------------|---------------|------------------|
| | Improved | Non-improved | Improved | Non improved | Improved | Non-improved |
| | n = 10 | n = 10 | n = 10 | n = 10 | n = 10 | n = 10 |
| Gait domain | 33 (16 to 48) | 54 (35 to 69) NS | 84 (57 to 100) | 50 (34 to 81) NS | 49 (26 to 57) | -1 (-6 to 11)*** |
| Cognitive domain | 64 (37 to 73) | 60 (46 to 80) NS | 75 (57 to 85) | 69 (48 to 80) NS | 10 (6 to 21) | 3 (-6 to 11) NS |
| Continence domain | 60 (20 to 80) | 80 (60 to 80) NS | 90 (75 to 100) | 70 (55 to 80) NS | 30 (0 to 45) | 0 (-20 to 5)** |
| Balance domain | 67 (67 to 71) | 67 (67 to 83) NS | 75 (67 to 87) | 67 (67 to 83) NS | 0 (-4 to 20) | 0 (-16 to 0) NS |
| Total iNPH score | 50 (36 to 64) | 63 (56 to 70) NS | 77.3 (71 to 87) | 64 (52 to 71)* | 26 (21 to 30) | 1 (-3 to 3)*** |
| *p ≤ 0.05 ** p ≤ 0.01, | *** p ≤ 0.001, N | S; non-significant. \$ | Significance calcu | lated by Man Whitn | ey U test | |

Table 3. INPH scale score pre op, post op and outcome (median and IQR) in the improved and non-improved group.

| | Improved | Non-improved | |
|--|---------------------|---------------------|----|
| | n = 10 | n = 10 | |
| Age, mean (SD) | 70.3 (3.2) | 71.6 (8.0) | NS |
| Female, n (%) | 5 (50) | 3 (30) | NS |
| Sickness duration (months), mean (SD) | 42 (21) | 34 (28) | NS |
| Diabetes, n (%) | 2 (20) | 2 (20) | NS |
| Hypertension, n (%) | 5 (50) | 6 (60) | NS |
| Cardiovascular disease, n (%) | 2 (20) | 1 (10) | NS |
| MMSE, median (Q1-Q3) | 23 (22 to 28) | 26 (24 to 28) | NS |
| ARWMC, median (Q1-Q3) | 6 (4 to 10) | 11 (5 to 20) | NS |
| El, median (Q1-Q3) | 0.43 (0.38 to 0.46) | 0.39 (0.36 to 0.41) | NS |
| NS = non-significant. Significance calculated by M | an Whitney U test | | |

Table 4. Comparison of clinical data of improved and non-improved iNPH patients at baseline.

CSF biomarkers in idiopathic normal pressure hydrocephalus.

6.5.3 STUDY III

In *study III* we included 82 patients with iNPH, 70 with Parkinson's disease (PD), 34 with multiple systems atrophy (MSA), 34 with progressive supranuclear palsy (PSP), 15 with corticobasal degeneration (CBD), 50 with Alzheimer's disease (AD), 19 with frontotemporal dementia (FTD), 75 with vascular dementia (VAD) and 54 neurologically healthy individuals (HI).



Figure 4. An overview of the different patient cohorts.

The 82 patients with iNPH (diagnosed by standard protocol), were selected retrospectively from our database. Patients that had received the diagnosis of iNPH and had undergone pre- and postoperative examination were

consecutively included. All patients had received a ventriculo-peritoneal shunt with an adjustable valve, anti-siphon device and a Rickham reservoir.

From the Sahlgrenska University Hospitals' movement disorders unit, we included 153 patients with movement disorders. The movement disorders group included 70 patients with definite PD according to the United Kingdom Parkinson's Disease Society Brain Bank clinical diagnostic criteria ¹³⁵; 34 with probable multiple system atrophy (MSA) according to Gilman's criteria ¹³⁶; 34 with probable or definite progressive supranuclear palsy (PSP) according to the National Institute of Neurological Disorders and Stroke and Society for Progressive Supranuclear Palsy, Inc. clinical criteria ¹³⁷; and 15 with probable corticobasal degeneration (CBD) according to Armstrong et al.¹³⁸.

The cognitive disorders group included 144 patients. In this group, 50 patients with AD and 19 patients with frontotemporal lobar degeneration (FTLD) were diagnosed at the Department of Neurology at Kuopio University Hospital, Kuopio, Finland by an experienced neurologist specialized in memory disorders. All patients in the AD group met the NINCDS-ADRDA (National Institute of Neurological and Communicative Disorders and Stroke and the Alzheimer's Disease and Related Disorders Association) criteria for probable AD ¹³⁹ and FTLD patients were diagnosed according to the Neary criteria ¹⁴⁰. The 75 VAD patients were diagnosed at the Department of Geriatric Medicine at Linköping University Hospital, Sweden using the ICD-10 criteria ¹⁴¹ and to the NINDS-AIREN (Association sub grouped according

46

Internationale pour la Recherche et l'Enseignement en Neurosciences) criteria ¹⁴².

Fifty-four (54) HI were included in the analysis. The group was composed of 20 individuals from Kuopio University Hospital, Finland and 34 from Linköping University Hospital, Sweden. At both centers, neurologically healthy individuals with an MMSE score \geq 26, undergoing planned surgical orthopedic intervention with spinal anesthesia and provided their informed consent to CSF sampling were included.

| | Ŧ | HdNi | AD | FTLD | VAD | PD | MSA | PSP | CBD |
|---|---|--|--|---|---|---------------------------------------|--------------------------------|---------------------------------|----------------------------------|
| | n = 54 | n = 82 | n = 50 | n = 19 | n = 75 | n = 70 | n = 34 | n = 34 | n = 15 |
| Female, n (%) | 32 (59) | 29 (35) | 29 (58) | 14 (74) | 45 (60) | 23 (33) | 20 (59) | 20 (59) | 10 (67) |
| Age, mean (SD) | 71 (10) | 73 (7)*** | 71 (7) | (6) 69 | 79 (6)### ^{&&&} | 60 (12)### ^{&&&} | 65 (8)### | 70 (7) | 68 (9) |
| Age is presented as mean anc by Kruskal-Wallis one-way an 0.001 (versus iNPH); ^{&&&} P < v | l SD. Sex is p alysis of rank 0.001 (versus | resented as nu s. Pair-wise an HI). No corre | umber of fem lalysis was m ction for the r | ales and %. (ade by Wilco nass-significo | Significance testii xon-Mann-Whitn ance was made. | rg in comparison vey U-test and sho | with iNPH and wn as *** P . | d healthy con < 0.001 (all g | trols was done cups), ### P < |

Table 5. Sex and age in iNPH, contrast groups and healthy individuals.

"

CSF biomarkers in idiopathic normal pressure hydrocephalus.

6.5.4 STUDY IV

For *study IV*, we included 52 patients with iNPH, 17 patients with subcortical small vessel disease (SSVD) and 28 healthy individuals (HI).

The iNPH patients were diagnosed with iNPH according to standard protocol between 2007 and 2012 at the Hydrocephalus research unit and CSF samples were selected retrospectively. All patients had undergone preand postoperative evaluation according to protocol and had received a ventriculo-peritoneal shunt with an adjustable valve, anti-siphon device and a Rickham reservoir.

In collaboration with the memory clinic at Sahlgrenska University Hospital, we included patients with SSVD and HI. All were a part of the Gothenburg MCI study ¹⁴³.

SSVD patients were diagnosed using the Erkinjuntti criteria ¹⁴². More specifically, the patients were required to have mild, moderate or severe white matter changes (WMC) according to Fazekas classification ¹³⁴ and predominant frontosubcortical symptoms such as mental slowness, executive dysfunction and extrapyramidal motor signs but without pronounced memory loss.

Healthy individuals were primarily recruited through senior citizens organizations, e.g. at information meetings on dementia, and a small proportion were relatives of patients ¹⁴³. None of the HI had diseases known to cause cognitive impairment nor did they exhibit any cognitive decline.

49

| Table 6. Age, | sex and | MMSE in | iNPH, | SSVD | and | controls. |
|---------------|---------|---------|-------|------|-----|-----------|
|---------------|---------|---------|-------|------|-----|-----------|

| | iNPH | SSVD | HI |
|----------------|-------------------------------|---------------------------------------|------------|
| | n = 52 | n = 17 | n = 28 |
| Age, mean (SD) | 72 (7)*,# | 71 (7) | 68 (4) |
| Female, n (%) | 23 (44) | 12 (71) | 10 (36) |
| MMSE | 24 (22-27) ^{***,###} | 27 (25-28) ^{&&&} | 30 (29-30) |

Age is presented as mean and SD. Sex is presented as number of females and %. MMSE is presented as median and interquartile range (IQR). Significance testing in comparing all groups was done by Kruskal-Wallis oneway analysis of ranks. Pair-wise analysis was made by Wilcoxon-Mann-Whitney U-test. * P < 0.05, *** P < 0.001 (all groups), #P < 0.05, ##P < 0.001(iNPH versus controls); &&& P < 0.001 (SSVD versus controls). No correction for the mass-significance was made.

6.5.5 OVERLAP

Study design and patient selection differed between the different studies. Even so, there is a substantial overlap between them. In Table 7 the overlap is shown. Each column represents one study and each row shows how many patients in that study that are present also in the other studies, i.e. the number of patients common to the specific studies. The bottom line represents the total number of patients in each study.

| STUDY I | STUDY II | STUDY III | STUDY IV |
|---------|----------|-----------|----------|
| 4 | | | |
| 4 | 4 | 4 | |
| 4 | 4 | 4 | 4 |
| 9 | | 9 | |
| 1 | | | 1 |
| | 3 | 3 | |
| | 8 | 8 | 8 |
| | 1 | | 1 |
| | | 24 | |
| | | 24 | 24 |
| | | | 8 |
| 6 | | 6 | 6 |
| 28 | 20 | 82 | 52 |

Table 7. Overlap of patients in study I-IV.

7 RESULTS

7.1 SEPARATING INPH FROM HEALTHY INDIVIDUALS WITH CSF BIOMARKERS

The potential of separating iNPH patients from healthy individuals (HI) was explored in *study I, II, III and IV*.

In *study I*, we reported that iNPH patients exhibited elevated levels of NFL and MCP1 in combination with a lowering of all APP-derived proteins as well as tau proteins. The interleukins IL-8 and IL-10 were not elevated, nor was MBP in comparison with HI. Moreover, albumin levels (absolute concentrations and CSF/ plasma ratio) were measured and did not significantly differ between iNPH patients and HI. Results from *study I* are presented in Table 8.

| iNPH | HI |
|------------------|---|
| (n = 28) | (n = 20) |
| 1260 (840-2290) | 825 (653 -1243)* |
| 1.5 (1.1 -1.9) | 1.3 (1.0 1.5) NS |
| 637 (438-894) | 1641 (1231 -2173)*** |
| 5067 (3634-6573) | 10083 (7626 -12794)*** |
| 221 (156-325) | 498 (391 -669)*** |
| 505 (338-739) | 1110 (727 -1244)*** |
| 176 (110-258) | 414 (250 -545)*** |
| 39 (34 -50) | 84 (64-107)*** |
| 39 (33 -50) | 59 (47 -75)** |
| 34 (26-38) | 31 (26-40) NS |
| 0.66 (0 -0.9) | 0.67 (0 -0.8) NS |
| 746 (602-874) | 628 (564 -686)* |
| 287 (188 -408) | 232 (203-280) NS |
| 6.8 (5.0 -10) | 5.6 (4.5 -6.4) NS |
| | iNPH (n = 28) 1260 (840-2290) 1.5 (1.1 -1.9) 637 (438-894) 5067 (3634-6573) 221 (156-325) 505 (338-739) 176 (110-258) 39 (34 -50) 39 (33 -50) 34 (26-38) 0.66 (0 -0.9) 746 (602-874) 287 (188 -408) 6.8 (5.0 -10) |

Table 8. CSF biomarkers in iNPH and HI (I).

Analysis made by Wilcoxon Mann-Whitney U-test and shown as * $p \le 0.05$, ** $p \le 0.01$, *** $p \le 0.001$,NS; non-significant. Values are given as median and Q1-Q3 range.

In *study II*, the biomarker focus was placed on amyloid metabolism. Data supported the results from *study I* of a lowering of all APP-derived proteins in iNPH in relation to HI. The metabolic products of the APP homologues APP- like protein 1 (*APLP1*) were also determined. Here, APL1β28 was slightly reduced in iNPH while APL1β25 and -27 were elevated. NFL and YKL40 did not differ between iNPH and HI (Table 9).

| | iNPH | HI |
|----------------|------------------|----------------------|
| | n = 20 | n = 20 |
| NFL (ng/L) | 1185 (731-2103) | 938 (610-2141) NS |
| APL1β25 (ng/L) | 2591 (2296-2951) | 2180 (1898-2386) *** |
| APL1β27 (ng/L) | 1083 (887-1177) | 874 (796-964) *** |
| APL1β28 (ng/L) | 1423 (1317-1550) | 1621 (1422-1797) ** |
| Aβ38 (ng/L) | 502 (266-625) | 1114 (819-1445) *** |
| Aβ40 (ng/L) | 3676 (2190-4748) | 7682 (6366-9809) *** |
| Aβ42 (ng/L) | 241 (144-405) | 754 (493-1058) *** |
| sAPPα (ng/mL) | 207 (157-259) | 416 (323-665) *** |
| sAPPβ (ng/mL) | 119 (92-170) | 280 (182-389) *** |
| YKL40 (ng/mL) | 122 (90-167) | 137 (104-177) NS |

Table 9. CSF biomarkers in iNPH and HI (II)

Analysis made by Wilcoxon Mann-Whitney U-test and shown as ** $p \le 0.01$, *** $p \le 0.001$,NS; non-significant. Values are given as median and Q1-Q3 range.

Results from *study III* are presented in Table 10. Here we used the significant biomarkers from *study I*. The study could replicate the results from *study I*, with a lowering of all APP-derived proteins and tau proteins whereas MCP1 was elevated. NFL was elevated at a trend level but did not reach statistical significance.

| | iNPH | н |
|---------------|------------------|---------------------|
| | n = 82 | n = 54 |
| NFL (pg/mL) | 1155 (821-1676) | 754 (540-1771) NS |
| MCP1 (pg/mL) | 477 (421-561) | 387 (341-477)* |
| Aβ38 (pg/mL) | 1543 (1089-1824) | 2148 (1681-2591)*** |
| Aβ40 (pg/mL) | 3727 (2808-4723) | 5123 (4313-6494)*** |
| Aβ42 (pg/mL) | 345 (274-438) | 477 (364-707)*** |
| sAPPα (pg/mL) | 429 (312-562) | 679 (471-796)*** |
| sAPPβ (pg/mL) | 324 (238-387) | 482 (363-634)*** |
| T-tau (pg/mL) | 206 (157-307) | 296 (209-477)NS |
| P-tau (pg/mL) | 30 (24-37) | 44 (35-59)*** |

Table 10. CSF biomarkers in iNPH and HI (III)

Analysis made by One-way ANCOVA with Dunnett's multiple comparisons test corrected for age and sex and shown as * p \leq 0.05, *** p \leq 0.001, NS; non-significant. Values are given as median and Q1-Q3 range

APP-derived proteins and biomarkers of damage to the subcortical zone were the focus of *study IV*. All APP-derived proteins were lower in iNPH than in HI. NFL, MBP and GFAP were all elevated. MMP-10 was slightly elevated but the concentration of markers of extracellular matrix remodeling did not differ between iNPH and HI.

| | iNPH | HI |
|----------------|----------------------|-------------------------|
| | n = 52 | n = 28 |
| sAPPα (pg/mL) | 384 (303-593) | 850 (694-1207)*** |
| sAPPβ (pg/mL) | 227 (170-325) | 516 (446-664)*** |
| Aβ38 (pg/mL) | 1333 (823-1928) | 2855 (2266-3261)*** |
| Aβ40 (pg/mL) | 3541 (2206-5648) | 7009 (5570-7814)*** |
| Aβ42 (pg/mL) | 361 (232-496) | 693 (510-931)*** |
| NFL (pg/mL) | 1592 (1012-2519) | 889 (694-1072)*** |
| GFAP (pg/mL) | 876 (659-1146) | 559 (381-718)*** |
| MBP (pg/mL) | 1,997 (1,407-2,503) | 1,446 (1,228-1,632)*** |
| MMP-1 (pg/mL) | 26 (16-47) | 24 (19-33) NS |
| MMP-2 (pg/mL) | 21190 (18965-23600) | 21317 (18423-23549) NS |
| MMP-3 (pg/mL) | 221 (162-322) | 238 (201-344) NS |
| MMP-9 (pg/mL) | 160 (114-205) | 129 (89-160) NS |
| MMP-10 (pg/mL) | 49 (38-67) | 42 (31-49)* |
| TIMP-1 (pg/mL) | 99329 (87306-113161) | 86094 (78696-107987) NS |

Table 11. CSF biomarkers in iNPH and HI (IV)

Analysis made by Wilcoxon Mann-Whitney U-test and shown as * $p \le 0.05$, *** $p \le 0.001$,NS; non-significant. Values are given as median and Q1-Q3 range.

In Table 12, results displaying the difference between iNPH and HI are shown schematically. It can be seen that the results with lower levels of APP-derived proteins and tau proteins were replicated consistently. There was an elevation of MCP1. The significance of an elevated NFL and MBP differed between the studies.
| | I | II | III | IV |
|-------------|--------------|--------------|--------------|--------------|
| NFL | 1 | NS | NS | 1 |
| MBP | NS | - | - | ↑ |
| GFAP | - | - | - | 1 |
| YKL40 | - | NS | - | - |
| MCP1 | ↑ | - | ↑ | - |
| Albumin CSF | NS | - | - | - |
| Alb ratio | NS | - | - | - |
| IL-8 | NS | - | - | - |
| IL-10 | NS | - | - | - |
| MMP-1 | - | - | - | NS |
| MMP-2 | - | - | - | NS |
| MMP-3 | - | - | - | NS |
| MMP-9 | - | - | - | NS |
| MMP-10 | - | - | - | 1 |
| TIMP-1 | - | - | - | NS |
| Αβ38 | \downarrow | \downarrow | \downarrow | \downarrow |
| Αβ40 | \downarrow | \downarrow | \downarrow | \downarrow |
| Αβ42 | \downarrow | \downarrow | \downarrow | \downarrow |
| sAPPα | \downarrow | \downarrow | \downarrow | \downarrow |
| sAPPβ | \downarrow | \downarrow | \downarrow | \downarrow |
| APL1β25 | - | 1 | - | - |
| APL1β27 | - | 1 | - | - |
| ΑΡL1β28 | - | \downarrow | - | - |
| T-tau | \downarrow | - | \downarrow | - |
| P-tau | \downarrow | - | \downarrow | - |

Table 12. An overview of the difference in CSF biomarker concentration between iNPH patients and HI.

Direction of arrow indicates level in iNPH as compared to HI. NS = non-significant

7.2 THE DIFFERENTIAL DIAGNOSTIC CAPACITY OF CSF BIOMARKERS

In *study III*, we tested the CSF biomarkers that differed significantly between iNPH and HI in *study I* on a material consisting of some of the most common iNPH mimics; AD, FTLD, VAD, PD, MSA, PSP and CBD. We could show that concentrations of APP-derived proteins remained low in iNPH. The sAPPs were lower in iNPH than in AD, FTLD, VAD, PD and PSP but did not reach statistical significance in comparison with MSA and CBD. Most of the A β s were lower in iNPH with A β 42 equaling the level in AD, VAD and CBD. Tau proteins distinguished iNPH from the cognitive disorders, but did not separate iNPH from the movement disorders. MCP1 remained elevated in iNPH compared to the other disorders but this result was only significant vs PD and MSA when correcting for the effect of age and sex (Table 13).

Similarities and differences between iNPH and SSVD were analyzed in *study IV*. These two disorders share much of the vascular risk factor profile and the clinical (subcortical) picture. Hence, the discrimination between these two disorders remains the most clinically challenging. We could conclude that APP-derived proteins (except A β 42) were lower in iNPH than in patients with SSVD. There was no statistical difference in biomarkers of white matter damage (NFL or MBP) nor in astroglia activation (GFAP) or extracellular matrix remodeling markers (MMP-1, -2, -3, -9, -10 and TIMP-1) (Table 14).

| $\hat{\mathbf{x}}$ |
|--------------------|
| (III |
| CBD |
| and |
| PSP |
| MSA, |
| PD, |
| VAD, |
| FTLD, |
| 4D, |
| .put |
| NPH d |
| in i |
| concentrations |
| markers |
| \overline{f} bio |
| CSI |
| 13. |
| Table |

| | INPH | AD | FTLD | VAD | PD | MSA | PSP | CBD |
|-------------|--------------------|------------------------|--------------------------|------------------------|-------------------|-----------------|------------------|---------------|
| | n = 82 | n = 50 | n = 19 | n = 75 | n = 70 | n = 34 | n = 34 | n = 15 |
| T-tau | 245 (131) | 980 (333)*** | 342 (146) | 651 (594)*** | 224 (105) | 315 (200) | 329 (247) | 366 (230) |
| P-tau | 32 (12) | 96 (27)*** | 45 (13) | 63 (41)*** | 33 (12) | 35 (18) | 46 (38)* | 42 (19) |
| (bg/mL) | | | | | | | | |
| NFL | 1717(1963) | 1977 (3104) | 2089 (1401) | 2646 (3475) | 839 (622) | 2322 (987) | 2219 (2761) | 2137 (1178) |
| (bg/mL) | | | | | | | | |
| Aβ38 | 1526 (519) | 2710 (807)*** | 2324 (608)*** | 2136 (672)*** | 2056 (624)*** | 1888 (856)* | 2125 (1076)*** | 2091 (684)* |
| (bg/mL) | | | | | | | | |
| Aβ40 | 3800 (1193) | 6541 (1654)*** | 5801 (1222)*** | 5477 (1540)*** | 5067 (1391)*** | 4650 (1815)* | 5099 (1965)*** | 5197 (1561)** |
| (bg/mL) | | | | | | | | |
| Aβ42 | 364 (138) | 318 (95) | 569 (204)*** | 387 (191) | 548 (187)*** | 489 (203)* | 488 (170)* | 505 (216) |
| (bg/mL) | | | | | | | | |
| sAPPα | 446 (178) | 865 (310)*** | 738 (279)*** | 631 (262)*** | 715 (262)*** | 597 (209) | 650 (309)** | 585 (224) |
| (bg/mL) | | | | | | | | |
| sAPPB | 321 (121) | 599 (192)*** | 502 (166)** | 484 (188)*** | 503 (178)*** | 414 (142) | 470 (212)*** | 452 (154) |
| (bg/mL) | | | | | | | | |
| MCP1 | 492 (109) | 436 (162) | 400 (101) | 456 (115) | 382 (128)** | 365 (70)** | 410 (121) | 448 (272) |
| (bg/mL) | | | | | | | | |
| CSF bioma | rker concentration | is are shown as mea | n and SD. Significand | ce testing in compari | son with iNPH was | done by One-way | ANCOVA corrected | for age |
| and sex wit | h Dunnett's multip | ole comparisons test a | and shown as $* P < 0$. | 05, ** P < 0.01, *** P | o < 0.001. | | | , |

| | iNPH | SSVD |
|--|--|--|
| | n = 52 | n = 17 |
| sAPPα (pg/mL) | 384 (303-593) | 683 (475-847)** |
| sAPPß (pg/mL) | 227 (170-325) | 417 (232-458)** |
| Aβ38 (pg/mL) | 1333 (823-1928) | 2196 (1749-2505)** |
| Aβ40 (pg/mL) | 3541 (2206-5648) | 5428 (4678-6838)** |
| Aβ42 (pg/mL) | 361 (232-496) | 474 (320-558) NS |
| NFL (pg/mL) | 1592 (1012-2519) | 1638 (1150-3149) NS |
| GFAP (pg/mL) | 876 (659-1146) | 820 (472-976) NS |
| MBP (pg/mL) | 1,997 (1,407-2,503) | 1,691 (1,461-2,351) NS |
| MMP-1 (pg/mL) | 26 (16-47) | 34 (20-54) NS |
| MMP-2 (pg/mL) | 21190 (18965-23600) | 22244 (21146-25104) NS |
| MMP-3 (pg/mL) | 221 (162-322) | 250 (186-372) NS |
| MMP-9 (pg/mL) | 160 (114-205) | 163 (107-193) NS |
| MMP-10 (pg/mL) | 49 (38-67) | 63 (40-76) NS |
| TIMP-1 (pg/mL) | 99329 (87306-113161) | 105464 (87590-142345) NS |
| CSF biomarker concentrations are shown as by Wilcoxon-Mann-Whitney U-test and show | s medians and interquartile ranges (IQR). Sig m as ** P < 0.01, *** P < 0.001, NS; non-sign | inificance testing was made inficant. |

Table 14. CSF biomarkers in iNPH and SSVD (IV)

CSF biomarkers in idiopathic normal pressure hydrocephalus.

| | AD | FTLD | VAD | SSVD | PD | MSA | PSP | CBD |
|---------------------|-----------------|-----------------|------------------------|-------------------|---------------|----------------|-----------------|--------------|
| NFL | NS | NS | NS | NS | NS | NS | NS | NS |
| MBP | ı | | ı | NS | | | | |
| GFAP | ı | ı | ı | NS | , | ı | 1 | , |
| MCP-1 | NS | NS | NS | | \rightarrow | \rightarrow | NS | NS |
| MMP-1 | ı | | ı | NS | , | ı | 1 | , |
| MMP-2 | ı | ı | ı | NS | , | ı | 1 | , |
| MMP-3 | ı | ı | ı | NS | | | | |
| MMP-9 | ı | ı | ı | NS | , | 1 | 1 | , |
| MMP-10 | I | ı | ı | NS | | | | |
| TIMP-1 | ı | | ı | NS | | | | |
| Aβ38 | ← | ← | ← | ÷ | ← | ← | ← | ← |
| Aβ40 | ← | ← | ← | ÷ | ← | ← | ← | ← |
| Aβ42 | NS | ← | NS | NS | ← | ← | ← | NS |
| sAPPα | ← | ← | ~ | ÷ | ← | NS | ← | NS |
| sAPPβ | ~ | ← | ~ | ← | ← | NS | ~ | NS |
| T-tau | ~ | NS | ~ | | NS | NS | NS | NS |
| P-tau | ~ | NS | ~ | ı | NS | NS | ~ | NS |
| Arrows indicate lev | el in compariso | on with iNPH. I | <u> VS = non-signi</u> | ificant. Except f | or iNPH vs SS | VD, results ar | e corrected for | age and sex. |

Table 15. Schematic overview of biomarker concentrations in iNPH mimics in comparison with iNPH.

Anna Jeppsson

In *study III*, we constructed a predictive model for iNPH. The model consisted of T-tau, $A\beta40$ and MCP-1 (simplified model 10*MCP1 - Ab40 - 5*T-tau). ROC-curve (AUC-statistics) was calculated and yielded an AUC of 0.87 (iNPH vs HI), AUC 0.86 (iNPH vs non-iNPH disorders), AUC 0.80 (iNPH vs movement disorders) and AUC 0.92 (iNPH vs cognitive disorders). A prediction plot was constructed and is shown in Fig 5.



Figure 5. Prediction plot for estimating the probability of a patient suffering from iNPH or a non-iNPH disorder. T-tau is given in 4 concentrations, $A\beta 40$ is given in 8 different intervals whereas MCP-1 is shown as a continuous variable on the X-axes. Estimated probability of iNPH is given on the Y-axes.

7.3 CSF BIOMARKERS IN VENTRICULAR CSF

In *study I*, we compared the concentration of the biomarkers in lumbar with ventricular CSF, the rostro-caudal gradient (RCG). The APP-derived proteins, NFL and IL-8 were higher in lumbar than in ventricular CSF whereas the tau proteins, MBP and MCP-1 were higher in ventricular CSF. Results are shown in Fig 6 and 7.



Figure 6. Concentrations of NFL, MBP IL-8, IL-10 and MCP-1 in ventricular CSF per op and lumbar CSF pre op. Significance is calculated by the Wilcoxon Mann-Whitney U-test (I).



Figure 7. Concentrations of APP-derived proteins and tau proteins in ventricular CSF per op and lumbar CSF pre op. Significance is calculated by the Wilcoxon Mann-Whitney U-test (I).

The biomarkers change in ventricular CSF following surgery (fig 8 and 9). The APP-derived proteins and P-tau did increase. Results on T-tau are more conflicting. At a group level, levels did decrease but looking at individual patients, it rather seems as if most of the patients remained relatively unchanged, with some patients showing a more pronounced reduction. For NFL, the levels slightly increased although many patients remained stable in their levels. MBP did decrease.



Figure 8. Concentration of NFL, MBP, IL-8, IL-10 and MCP-1 in ventricular CSF per- and post surgery. Significance is calculated by the Wilcoxon signed rank test (I).



Figure 9. Concentration of APP-derived proteins and tau proteins in ventricular CSF per- and post surgery. Significance is calculated by the Wilcoxon signed rank test (I).

7.4 PREDICTING SHUNT RESPONSE BY CSF BIOMARKERS

In *study II*, we aimed to examine if the biomarker profile differed depending on whether the patient was improved by surgery or not. However, even in these heavily dichotomized groups, none of the CSF biomarker levels at baseline differed between the groups (Table 16).

| | Improved | Non-improved |
|----------------|---------------------|------------------------|
| | n = 10 | n = 10 |
| NFL (ng/L) | 1186 (869 to 1670) | 1085 (699 to 2432) NS |
| APL1β25 (ng/L) | 2532 (2174 to 2958) | 2820 (2401 to 2954) NS |
| APL1β27 (ng/L) | 1067 (900 to 1157) | 1085 (867 to 1214) NS |
| APL1β28 (ng/L) | 1423 (1264 to 1568) | 1458 (1291 to 1562) NS |
| Aβ38 (ng/L) | 500 (308 to 605) | 503 (224 to 677) NS |
| Aβ40 (ng/L) | 3731 (2642 to 4740) | 3677 (1522 to 4789) NS |
| Aβ42 (ng/L) | 241 (155 to 370) | 244 (122 to 438) NS |
| sAPPα (ng/mL) | 205 (175 to 279) | 212 (144 to 297) NS |
| sAPPβ (ng/mL) | 114 (95 to 155) | 127 (75 to 181) NS |
| YKL40 (ng/mL) | 122 (99 to 153) | 134 (84 to 180) NS |

Table 16. CSF biomarker levels in improved vs non-improved iNPH patients.

Analysis is made by Man Whitney U test., NS; non-significant. Values are given as median and IQ-range.

7.5 RADIOLOGICAL WHITE MATTER CHANGES

White matter changes were assessed in *study II* and *IV*. ARWMC were most abundant in the frontal and parietal-occipital areas. In *study II*, the patients were divided into improved and non-improved. Non-improved patients had slightly more ARWMC (median (IQR) 11, (5-20)) than improved patients (median (IQR) 6, (4-10)) but the difference did not reach statistical significance.

Compared to patients with SSVD, iNPH patients displayed less ARWMC. Ventricle size, though, was slightly larger in iNPH than in SSVD.

In table 17 and 18, ARWMC scores are presented for each sub-region and as a total score. In every domain ARWMC is reported for right (R) and left (L) hemisphere separately.

| | iNPH |
|--------------------|-------------------|
| Brain region | n = 20 |
| Frontal | |
| R | 1.5 (1 -3) |
| L | 1.5 (1 - 3) |
| Parietal-occipital | |
| R | 1 (1 - 2) |
| L | 1 (1 - 2) |
| Temporal | |
| R | 0 (0 - 1) |
| L | 0 (0 - 1) |
| Basal ganglia | |
| R | 0 (0 - 1) |
| L | 0 (0 - 1) |
| Infratentorial | |
| R | 0 (0 - 2) |
| L | 0 (0 - 2) |
| Total | 9 (4 - 13) |
| Evans' Index | 0.4 (0.36 - 0.45) |

Table 17. ARWMC in iNPH (II).

Values are given as median and Q1-Q3.

| | iNPH | SSVD |
|--------------------|-----------------|---------------------|
| Brain region | n = 51 | n = 14 |
| Frontal | | |
| R | 1 (1-2) | 2 (2-3) |
| L | 1 (1-2) | 2 (2-3) |
| Parietal-occipital | | |
| R | 1 (1-2) | 2.5 (1-3) |
| L | 1 (1-2) | 2.5 (1-3) |
| Temporal | | |
| R | 0 (0-1) | 0 0-0) |
| L | 0 (0-1) | 0 (0-0.25) |
| Basal ganglia | | |
| R | 0 (0-1) | 1 (0-2.25) |
| L | 0 (0-0) | 1 (0-2.25) |
| Infratentorial | | |
| R | 0 (0-1) | 1 (0-2) |
| L | 0 (0-1) | 0.5 (0-2) |
| Total | 7 (4-11) | 12 (8-22)** |
| Evans' Index | 0.4 (0.37-0.44) | 0.31 (0.27-0.38)*** |

Table 18. ARWMC in iNPH and SSVD (IV).

Values are given as median and Q1-Q3. ** P < 0.01, *** P < 0.001.

8 DISCUSSION

The overall aim of this thesis was to explore the diagnostic and prognostic potential of CSF biomarkers in iNPH and hereby elucidate underlying pathophysiologic mechanisms of the disorder. Here, I will start by discussing the pathophysiological implications following the CSF biomarker studies.

8.1 AMYLOIDS IN INPH

In all of our studies, iNPH patients presented with lower levels of all APPderived proteins in comparison with HI. Being the surrogate marker for AD plaque pathology, A β 42 is the marker studied the most, together with the other component of plaques: A β 40. Most studies have found lower A β 42^{69 144-152} and A β 40^{145 149 152} in NPH/ iNPH in comparison to HI even if some groups presented compatible levels between iNPH and HI of A β 42¹⁵³⁻¹⁵⁶ and A β 40^{146 155}. However, it is noteworthy that nearly all studies that showed compatible levels are from the same research group, indicating a possible overlap in these studies. All studies on CSF biomarkers in iNPH so far have showed lower levels of sAPP α and $-\beta$ in iNPH ^{146 149 152 153 155 156} as well as A β 38^{149 152 155}.

APL1 is a homologue to APP ⁹⁹. In *study II* we could show that the cleavage products of APL1, APLP1β, were not affected to the same extent, nor in

the same manner as APP. These findings are supported by the only other study on the subject ¹⁵⁵. As such, we would like to argue that reduced levels of APP-derived proteins are specific and characterize the CSF biomarker profile of patients with iNPH in comparison with HI.

There have been theories proposing a common pathological state in NPH and AD. According to these theories, NPH and AD share a common pathophysiological aetiology in that reduced clearance of A β would lead to AD-like pathology in iNPH-patients ⁹⁷ ⁹⁸, a theory in part based on observations of rats with Kaolin-induced hydrocephalus (i.e. rather sNPH than iNPH).

We believe that the results from studies expanding the APP-derived biomarkers studied here, clearly speaks against this hypothesis of a common pathophysiology. In AD, there is an isolated reduction of A β 42 in CSF whereas the other A β and sAPP fragments remain unaltered or even elevated. In iNPH there is a more general reduction of APP-derived proteins in CSF. We believe that this indicates disturbances in different parts of the amyloid cascade, where AD is a downstream phenomenon in the cascade. iNPH on the other hand, seems to suffer from an upstream disturbance or/ together with a reduced clearance affecting all fragments. In CSF, when measuring A β 42 levels between iNPH and AD, our results (*III*) as well as results from several other studies will indicate that levels of A β 42 do not differ between the groups ^{78 144 150 151 153 157}. Some studies have also shown an increase ^{145 155 156 158} or a decrease of A β 42 ^{147 148} in iNPH in comparison with AD. A β 38 is lower in iNPH than in AD ¹⁵⁵ and in contrary to our study, A β 40 has found to be in the same level as in AD ^{145 155}. As for the sAPPs, our study (*III*) and all other studies show that patients with iNPH have lower levels than patients with AD ^{153 155 156}.

Around 30 % of patients with iNPH do exhibit AD-like pathology in cortical biopsies ¹⁵⁹. As AD is a common comorbidity or differential diagnosis, we argue that it is of importance not to base the evaluation on A β 42 solely, but to simultaneously look at different parts of the cascade to aid in diagnostic queries.

There have been previous attempts in treating patients with AD with a low flow ventriculoperitoneal shunt with the underlying assumption that this would facilitate CSF turnover and hence drain the CSF of accumulated amyloid. However, these attempts, although initially promising, have not been successful ^{46 160 161}.

If APP clearance is hindered by the elevated centrifugal pressure in iNPH and if this would explain the lower concentrations, it might be that APPs can be cleared from a stagnant pool in the ECS once the force applied is being removed. This has led to some objections for the usage of CSF biomarkers in iNPH and that this reduction of ECS and thus clearance of amyloids into the CSF might hinder the interpretation of differences in biomarker concentrations analysis between iNPH and AD ¹⁶².

There are few studies that have addressed the relation between ISF and CSF levels of CSF biomarkers. Herruka et al reported the level of Ab1-42 to be quite similar between ISF and ventricular CSF (VCSF) levels ¹⁶³. Tau proteins were higher in ISF than VCSF. Aβ42 and P-tau remained quite

stable in ISF in a 21 h period during microdialysis, whereas T-tau was increased initially and falling to a stable plateau within hours. This could indicate that the levels in CSF are actually mirroring the actual amyloid levels, arguing against an isolated mechanistic drainage dysfunction. We believe that such a dysfunction would also hinder the drainage of APL1 derived proteins which does not seem to be the case ^{149 155}. Larger studies analysing the relation between ISF/CSF concentrations are needed.

A common objection to the suggested CSF biomarker findings in iNPH is that of a possible dilution mechanism due to increased CSF amount. However, there seems to be no correlation between ventricle volume and CSF biomarkers ¹⁶⁴. Also, the finding that not all proteins behave in the same way would argue against a dilution mechanism that accounts for the low APP-derived proteins.

The slight increase of APL1 β 25 and 27 seen in *study II* could indicate increased cleavage of APLP-1 by γ -secretase as a response to a reduction of its primary substrate APP. In iNPH an increase in γ -secretase activity in cortical biopsies in patients with iNPH with A β -pathology has been shown ¹⁶⁵ whereas in Downs syndrome (where APP is overly-expressed) there is a decreased level of all APL1 β which could be in accordance with a less available γ -secretase ¹⁶⁶. These findings support the notion of less available APP proteins in iNPH.

Something happens with APP-derived proteins after shunt surgery. In *study* I, concentrations in ventricular CSF increased post op ¹⁵². Tarnaris et al, studying biomarker changes in iNPH during ELD, showed that A β 42

increased during lumbar drainage ⁷². The same increase of A β post op has been reported earlier ¹⁵⁵. However, post op values are not so easily interpreted. The shunt would probably change the CSF flow dynamics as a new route of low resistance is being introduced. This would also presumably affect the manner in which amyloids are drained if the studies indicating the importance of the glymphatic system for amyloid drainage are proven right ⁵⁶.

More studies of biomarkers pre- and post shunt surgery are warranted to bring the dynamic changes caused by shunt surgery into clarification.

8.2 CORTICAL PATHOLOGY IN INPH?

T-tau indicates cortical neural damage whereas P-tau is indicative of misphosforylation of tau which is seen in neurofibrillary tangles in AD ^{67 106}. In *study I* and *III* we measured both T-tau and P-tau in iNPH and they were found to be lower in iNPH than in HI. A number of studies have shown compatible ^{144-148 150 153-155 167} or lower ⁶⁹ levels of T-tau and compatible ^{78 146} ^{147 150 151 153 156} or lower ^{69 148 154 155} levels of P-tau. There are two studies with contradictory results. Kudo et al and Kapaki et al have showed elevated levels of T-tau ^{73 78}. In the first study, the cohort was mixed between idiopathic and secondary cases with aetiologies known to increase tau levels. In the second study, T-tau was significantly increased but looking closer at the results, there were two patients with iNPH that had very high

levels of T-tau whereas the rest of the patients seemed to have values well in line with the control sample which might have distorted the statistics.

In *study III* we could show that the movement disorders (PD, PSP, MSA and CBD) exhibited levels of T- and P-tau in the same range as iNPH. The same has been shown previously ^{144 148 150 154} and tau-proteins do not seem to separate iNPH from movement disorders. The cognitive disorders did however have elevated tau proteins. AD patients exhibit higher levels of T- ^{78 144 145 147 148 150 153-155 157 158} and P-tau ^{78 147 148 150 151 153 155-158} than iNPH. The same can be said when comparing to FTLD ^{145 150}. Dementias of vascular origin show more mixed results. Previous results on VAD showed levels similar to iNPH ¹⁵⁰ whereas the VAD-patients in our study had higher levels. SAE have shown higher levels ⁶⁹. We know that there are individuals with VAD in our study that have cortical infarctions which might explain higher tau levels.

Patients with mild cognitive impairment (MCI) have shown higher levels of tau proteins than patients with iNPH ¹⁴⁷. A large proportion of MCI patients go on to develop AD and thus it is likely that presence of AD would obscure the image.

Given these results, we believe that iNPH does not involve any extensive damage to cortical structures and that elevated tau levels in patients with iNPH would indicate presence of cortical pathology (e.g. AD) that needs to be taken into consideration by the clinician.

8.3 DAMAGE TO WHITE MATTER AND GLIA ACTIVATION

MBP, a marker of oligodendroglia, is in hydrocephalus presumed to reflect myelin destruction in the periventricular zone ¹¹². There is an increase of MBP in CSF in iNPH (*I*, *IV*) (even if not reaching significance in *study I*). Previously, only one study has measured MBP in hydrocephalic (i.e. not NPH) patients and 80 % exhibited increased levels, even if this mixed cohort of high- and low pressure hydrocephalus might not represent an iNPH cohort ¹¹². Following surgery, MBP concentrations are lowered which we interpret as a reduction of myelin destruction.

As previously stated, there are many clinical similarities between iNPH and SSVD, thought to arise from subcortical structures ¹⁶⁸. Studies on SSVD had indicated a CSF biomarker subcortical profile ¹⁶⁹ and we wanted to try this panel for our iNPH patients. The markers of extracellular matrix remodelling (MMPs and TIMP1) were not elevated in iNPH in comparison to HI nor to SSVD (IV). Thus, in this study, we could not reproduce the suggested profile for subcortical CSF biomarkers ¹⁶⁹. No other study has measured these markers in iNPH.

NFL, an axonal degeneration marker ¹⁰⁶ was elevated in *study I* and *IV* but not significantly so in *II* and *III*. NFL has been shown to be increased in iNPH in comparison with HI before ^{69 167 170 171} and the levels seem compatible to those in SSVD/SAE/BD (*IV*) ^{69 167 171}. NFL was initially regarded as mainly representing large, periventricular axons but is now believed to be a more general marker of axonal degeneration ^{108 109}.

According to our clinical experience, we often find that iNPH patients present with NFL levels in the higher span of the reference interval but we seldom see very high levels in iNPH. One possible explanation to the high NFL concentrations seen in earlier studies could be that these studies included a mixture of idiopathic and secondary cases of NPH ^{167 170 171}. These results are in line with the notion that NFL is rather a marker of disease intensity than marker of a specific aetiology ¹¹⁰.

Astrocyte activation is in these studies mirrored by YKL40 (*II*) and GFAP (*IV*). YKL40 was not found elevated (*II*) whereas GFAP was elevated in comparison with HI but not with SSVD (*IV*). Elevated GFAP in iNPH has been shown before ^{167 170 172} and also the findings of compatible levels in SSVD ¹⁶⁷. It could be speculated that the reason for the discrepancy might be the low neuroinflammatory component in iNPH that does not trigger YKL40 ¹²⁴ production. Astrocyte activation has also been implicated in iNPH by elevated Leucine-rich α 2-glycoprotein (LRG) which is thought to be expressed by peri-capillary astrocytes ^{154 173-175}.

MCP-1 is also a cytokine which acts as a chemoattractant of astroglia, apart from the recruitment of monocytes from the periphery ¹¹⁵. It seems to be elevated in CSF of iNPH patients as compared to HI as shown in *study I* and *III*. It was also included in the predictive model to capture astroglial activation. Considering that no other group has measured MCP-1, these results needs replication.

Taken together, the CSF biomarker profile in iNPH is supportive of a subcortical profile with gliosis and possibly with astroglial activation. In

addition, the ARWMC staging in *II* and *IV*, are supportive of such a position with WMC being quite extensive even if they radiologically do not match those of SSVD. Postoperatively, MBP levels decreased in ventricular CSF (III) and previous studies have shown that also the WMC tend to improve post surgery ^{37 167}.

8.4 VASCULAR CHANGES IN INPH

Following the results in *study I*, we shaped the hypothesis of a biomarker pattern in iNPH reflecting a decreased periventricular metabolism which is also supported by MRI findings ^{176 177}. In *study IV*, we looked closer at this periventricular zone by exploring similarities and differencies between iNPH and SSVD, which we know both exhibit periventricular changes. The clinical pictures of iNPH and SSVD can often be very similar. The cognitive features are both fronto-subcortical (as opposed to posterior in e.g. AD) 168 and gait and urinary symptoms are frequent in both disorders. These similarities are not surprising, if we assume that the clinical manifestations in both disorders are partially the results of periventricular changes. In study IV, we saw that many patients with SSVD show ventriculomegaly and WMC are extensive in both disorders, even if they are more pronounced in SSVD. There is an increasing body of evidence focusing on the importance of vascular risk factors in patients with iNPH ¹⁷⁸⁻¹⁸⁰. To complicate matters even further, we know that patients with iNPH and extensive white-matter changes (Binswangers disease) can

respond to shunt treatment ^{37 181}. As for CSF biomarker changes, we can see that the two disorders do share patterns of biomarker alterations even if the changes in the iNPH patients are more pronounced.

Even if incompletely understood, the aetiology of SSVD is thought to arise from arteriolar dysfunction which mainly affects highly vascularized tissues, such as the brain's white matter ¹⁶⁸ ¹⁸²⁻¹⁸⁴. Contemporary research on NPH is separating sNPH cases from iNPH cases to a larger extent than earlier. We can see that this shift has somewhat changed our view on some of the CSF biomarkers. Presumably, focusing on pressure dynamics might have over-stressed the similarities of s- and iNPH in favor of looking at the underlying factors and that the aetiologies might be more different than previously thought. It may well be that the idiopathic form of NPH, as opposed to the secondary form, might arise from vascular changes. The nature of these changes are not completely understood. The new hypothesis on the glymphatic system might provide a piece of the puzzle. One could speculate that iNPH patients have a down regulation of APP proteins in response to a reduced parenchymal clearance of AB due to reduced turnover and this is what we see in lumbar CSF. We believe that further research into vascular changes and glymphatics will be an important future line of research in iNPH.

8.5 PREDICTING OUTCOME?

The question of predictability was addressed in *study II*. We could not find any CSF biomarker, nor any clinical marker that could be used to predict which patient that would benefit from shunt surgery. Non-improved patients presented with more ARWMC than the improved. In this small material, the difference between these heavily dichotomized groups was not significant but this might be attributed to a type II error due to small sample size. Regardless, patients with vascular changes improve to the same extent as those without and even extensive white matter changes should not be used as an argument against shunt surgery ^{37 171}.

There have been numerous attempts at finding objective criteria that could be used to select patients that would benefit from surgery, or rather not expose patients to surgery that we know would *not* benefit. As previously discussed, several tests can aid in patient selection, the most widely used being the CSF tap-test or other tests based on clinical response to CSF removal ³⁹. None of them have yet been able to show a specificity high enough to safely exclude patients from shunt surgery ⁴³. In an attempt to find markers of shunt responsiveness, Luikku et al used the disease state index (DSI) which consisted of demographical, clinical, radiological, and biochemical data from 284 patients (54 non-responders) to analyze which patients would benefit from shunt surgery. They concluded that the AUC of predicting shunt responsiveness was still 0.58 and thus remains extremely challenging ¹⁸⁵. Judging from the evidence it seems that to *diagnose* and to *predict reversibility* are two different tasks in iNPH. There are many

iNPH researchers that would oppose this view. In many centers worldwide, TT is still being used for diagnosing iNPH and in the Japanese guidelines, a positive TT is required to be diagnosed with probable iNPH ¹⁷. We do believe that the diagnosis of iNPH can still be correct even if the patient is non-responsive to shunt treatment, and that outcome after shunting is mere a measure of reversibility, not diagnosis.

Co-morbidity is, needless to say, an important factor when attempting to predict the disease course for the individual patients. There are some studies indicating that CSF biomarkers could be used to map co-morbid neurodegenerative disorders and thus to predict long-term cognitive outcome ^{151 186 187}. Neurodegenerative co-morbidity does not seem to impair the short term results ^{38 186} and it seems that even if co-morbidities are present, the "iNPH part" might still be reversible even if the course of the neurodegenerative disorder progresses over time as it would without shunt surgery. To that extent, CSF biomarkers could offer a good tool to signal the presence of co-morbidities that might influence the outcome.

8.6 CAN WE USE CSF BIOMARKERS TO DIAGNOSE INPH?

Our first research question was whether CSF biomarkers can aid in diagnosing iNPH. There are several answers to this question. We can conclude that the results from these studies lend support to the idea that iNPH has pathophysiological features that are discernible in the CSF biomarker pattern. But, it is rather the pattern than the individual marker that seems to hold the greatest promise of a diagnostic tool for iNPH. Following the studies in this thesis and of others research, we propose that the biomarker pattern in iNPH is summarized with a lowering of all APPderived proteins, no elevation of tau proteins and possibly a slight elevation of NFL and MCP1.

As a diagnostic team, we face a pre-selected group of persons that present with complaints that *could* be attributed to iNPH. As such, the clinical importance of CSF biomarkers is rather to distinguish which patients that have a pattern that can be attributed to iNPH from those that do not. In *study I* we were able to show this pattern and in *study III* we were able to repeat the findings and show that the combination of T-tau, A β 40 and MCP-1 separated iNPH patients from patients with movement- and cognitive disorders of different origin than iNPH with a high sensitivity and specificity. In sum, we do believe that CSF can aid as an additional tool when diagnosing iNPH.

8.7 GENERAL METHODOLOGICAL CONSIDERATIONS

Some general methodological considerations that need to be addressed have in part been mentioned earlier but will be elaborated upon here.

As for building a model to predict if a person can be said to suffer from iNPH, we have to consider how representative our sample is. Even if the CSF biomarkers are analysed using different samples, they are all from the same pre-selected setting of patients referred to the unit with the suspicion of iNPH. This would render the prevalence of iNPH much higher than in the general population (with an estimated prevalence of up to 2 % of persons over 65 years ¹²). Going even further, our contrast diagnoses are, even if at a specialized neurological hospital clinic, not under the suspicion of iNPH. In the real clinical setting, boundaries between different disorders are far more blurred. In addition, as stated in the 'methods' section, there is an overlap of patients in the different studies. Taken together it is of great importance that the results reported here are tested on other iNPH cohorts and that we carefully consider which patients the findings might be applicable to. This being said, we do think that the results can be of use. We do not claim to have found a diagnostic marker for iNPH that can be used in a preclinical setting (yet). But, going from the results presented herein, we do believe that we are ready to try the results prospectively at our clinic. This will be one of the upcoming studies.

Moreover, it is possible that our iNPH cohort might in turn represent a sub-population as the exact algorithm for diagnosing iNPH differs across the globe. Therefore, we do believe that it is of great importance to carefully examine CSF biomarkers in different iNPH cohorts to bring this into clarity.

In order to deliver diagnostic CSF biomarkers there are some concrete methodological limitations. Many of the assays are not as of yet in clinical practice but are rather developed for research purposes. The between run variability is too large to be able to compare absolute levels between different studies, making it, at this stage, impossible to discuss cut-off levels for the biomarkers chosen. We know that it can be done, as in the case with core AD-biomarkers, but we are not there yet.

9 CONCLUSIONS AND FUTURE PERSPECTIVES

This thesis indicates that iNPH exhibits a CSF biomarker pattern that distinguishes patients with iNPH from healthy individuals of the same age but also from many of their mimics. This profile is characterized by a lowering of all APP-derived proteins, no elevation of tau proteins and an elevation of MBP, GFAP and MCP1.

We believe that these CSF changes reflect pathophysiological processes characteristic of iNPH. The lowering of APP-proteins seems specific and might be a consequence of a reduced production or/ in combination with a reduced clearance, specific for APP metabolites. No elevation of tau proteins indicates that there is no substantial cortical pathology in iNPH. The elevation of MBP indicates destruction of white matter and elevated GFAP and MCP1 indicates astrocyte activation.

As we view CSF biomarkers as a tool to study pathophysiological processes in iNPH we want to continue elaborating on these findings and combine them with additional biomarkers reflecting other aspects of pathophysiology. We plan to look more closely at synaptic function (in importance to APP) in iNPH by analyzing novel CSF biomarkers of synaptic function. Also, we want to continue exploring the relation between ISF and CSF. In order to do this, we would need to combine biochemical analysis (presumably with microdialysis and ventricular CSF) with radiological techniques, both MRI and possibly Positron emission tomography (PET). This combination of biomarkers (neuroimaging and biochemical) will also be combined with clinical data in order to explore if there is a close association between our biomarkers and clinical symptoms.

The biochemical changes in CSF of patients with iNPH seem to share common features with patients with SSVD. This could indicate that the underlying pathophysiology in iNPH and SSVD share common features, possibly connected to shared vascular risk factors and burden of vascular disease. These findings need elaborating, preferably by strengthening the collaboration with research milieus studying SSVD with the aim to explore longitudinal development of disease as well as similarities and differences that might be related to the reversibility of iNPH. It might even be that a proportion of the patients today classified as SSVD alone might actually benefit from shunt placement. This would relate to identifying biomarkers of reversibility. Even if we are not there yet, novel knowledge on pathophysiological processes might provide windows to explore this further. As of today, we have not found any CSF biomarker that can aid in the selection of patients eligible for shunt surgery but we can see that the CSF profile changes after surgery indicating a substantial effect from the shunt. More studies on CSF biomarkers in lumbar CSF pre- and post-surgery are needed as such studies are lacking in the literature. We want to understand how the shunt procedure affects the biomarker concentrations and also to see if alterations in CSF biomarkers are linked to improvement from surgery. We are collecting both lumbar pre – and post surgery CSF samples and also ventricular CSF per- and post surgery in order to investigate these relations further.

In order to combine these pathophysiological findings from the studies, we constructed a CSF biomarker algorithm of T-tau, A β 40 and MCP-1 that could prove useful for distinguishing iNPH from cognitive and movement disorders at specialized clinics. As for diagnostic purposes, we wish to elaborate on the model that we have shaped here. One step would be to assess the model on a different iNPH cohort. We have initiated a collaboration with another iNPH center for this purpose. We are also in the process of collecting CSF material prospectively on our cohort with no overlaps to the studies herein. We would also encourage other researchers to replicate the results presented in this thesis.

As the biochemical techniques advance quickly, many of the methods are now sensitive enough to measure CNS derived proteins in peripheral blood. We are already working on some of the CSF markers in plasma and/or serum and we hope that this field will expand in the future, opening up for blood tests aiding diagnostics in the not so distant future.

We hope that the results from this thesis and the studies to come will prove useful for our patients and their kin. We owe it to our patients to continue trying to render iNPH less idiopathic.

ACKNOWLEDGEMENTS

To all the iNPH patients whom in giving their consent to the studies made it possible to begin with. Thank you.

I have had the privilege of having four supervisors, which has been a blessing (and, at times, a curse);

Mats Tullberg, my main supervisor who has gently guided me through my process as a PhD student with a kindness and a patience that anyone could learn from. Moreover, you forced me to step out of my comfort zone and carry this thesis full term despite my many tricks to get away. It was worth it, thank you.

Carsten Wikkelsø, my co-supervisor, who was also my first supervisor when I started in this group. I owe you gratitude that extends far beyond the research projects. You believed in me in times when frankly no one else would. You provided me with a space of my own where I could think and get away in many situations when I really needed one. And of course, you showed how fun research can be.

Henrik Zetterberg, whose enthusiasm and curiosity cannot be exceeded. This seems to extend not only to science which is a true inspiration.

Kaj Blennow, who initially scared me a bit but whose competence in his subject and unpreceded ability to see the pathophysiological workings behind its CSF trails is truly remarkable.

I would like to thank all the co-authors of the manuscripts included in this thesis: *Mikko Holtta*, *Radu Constantinescu*, *Anne M Remes, Sanna-Kaisa Herukka*, *Tuomas Rauramaa*, *Katarina Nägga*, *Ville Leinonen*, *Maria Bjerke*, *Per Hellström*, *Petronella Kettunen* and *Anders Wallin*.

My colleagues at the Hydrocephalus research unit who also became my friends (and especially for the supportive attitude, all the laughter and all the good food); *Simon Agerskov, Gunilla Ahl-Börjesson, Kerstin Andrén* (whom needs an extra thank you for her fantastic ability to help me visualize what I am thinking), *Mikael Edsbagge, Dan Faramand, Per Hellström, Daniel Jaraj, Madeleine Johnsson, Lena Kollén, Tove Rasmussen, Maria Wallin* and Doerthe Ziegelitz.

The highly skillful staff at the neurochemical laboratory, not the least *Mariann Wall*.

For help with the English language editing, I would like to thank Judith Klecki.

To my friends- You are too many that I would like to thank than can fit into these pages. For that, I am sorry. I love you nevertheless.

Malin Hägglund, my friend and ally. Without you, I would go bonkers

Cecilia Verdinelli-Peralta. Mi hermana. As for this thesis, I owe you thanks for language and layout editing, for making the text readable and providing me with vivid images of how others before me have struggled with the same (and that throwing manuscripts in dumpsters is not necessarily a rational act).

For one of my absolutely longest lasting friendship: *Karin Svedberg* who has seen, and yet stood by, me.

Jomi 'Juttan'' Jutlöv, for the loyal friendship, the tears, the laughter and the fantastic illustrations in this thesis.

Anna Thulin, for support in keeping track of the important stuff.

For strategic choices, my go-to person; *Lina Holmqvist* (everyone should have one).

To the Linda "the library wizard" Hammarbäck.

My father, Olle, my mother Astrid, my younger brother Jojjo (with family), my older brothers Baltzar, Niklas and Jesper. Family: it's complicated.

To my aunt *Kerstin* who let me use her work, to my cousins Asa and *Frederik*.

To my godmother *Annika* and her sister *Gunilla* who have been a true support beyond what could be expected from anyone when my mother fell ill.

To the book-circle that forces me to read more than research papers and reminding of the world beyond: *Helga, Linda, Frida, Malin* and *Cecilia*.

To Alma, Anton and Viggo;

You mean the world to me.

Mama loves you.

Always.
REFERENCES

- 1. International. AsD. Policy Brief for G8 Heads of Government. The Global Impact of Dementia 2013-2050. London. *Alzheimer's Disease International* 2013
- Hakim S, Adams RD. The special clinical problem of symptomatic hydrocephalus with normal cerebrospinal fluid pressure. Observations on cerebrospinal fluid hydrodynamics. *Journal of the Neurological Sciences* 1965;2(4):307-27.
- 3. Adams RD, Fisher CM, Hakim S, et al. Symptomatic Occult Hydrocephalus with Normal Cerebrospinal-Fluid Pressure - a Treatable Syndrome. *New England Journal of Medicine* 1965;273(3):117-26.
- 4. Fisher CM. The clinical picture in occult hydrocephalus. *Clinical Neurosurgery* 1977;24:270-84.
- 5. Wallenstein MB, McKhann GM, 2nd. Salomon Hakim and the discovery of normal-pressure hydrocephalus. *Neurosurgery* 2010;67(1):155-9.
- 6. Marmarou A, Bergsneider M, Relkin N, et al. Development of guidelines for idiopathic normal-pressure hydrocephalus: introduction. *Neurosurgery* 2005;57(3 Suppl):S1-3.
- Tisell M, Hoglund M, Wikkelso C. National and regional incidence of surgery for adult hydrocephalus in Sweden. *Acta Neurologica Scandinavica* 2005;112(2):72-5.
- Klinge P, Hellstrom P, Tans J, et al. One-year outcome in the European multicentre study on iNPH. *Acta Neurologica Scandinavica* 2012;126(3):145-53.
- 9. Hiraoka K, Meguro K, Mori E. Prevalence of idiopathic normal-pressure hydrocephalus in the elderly population of a Japanese rural community. *Neurologia Medico-Chirurgica* 2008;48(5):197-99.
- Tanaka N, Yamaguchi S, Ishikawa H, et al. Prevalence of possible idiopathic normal-pressure hydrocephalus in Japan: the Osaki-Tajiri project. *Neuroepidemiology* 2009;32(3):171-5.

- 11. Brean A, Eide PK. Prevalence of probable idiopathic normal pressure hydrocephalus in a Norwegian population. *Acta Neurologica Scandinavica* 2008;118(1):48-53.
- 12. Jaraj D, Rabiei K, Marlow T, et al. Prevalence of idiopathic normalpressure hydrocephalus. *Neurology* 2014;82(16):1449-54.
- 13. Tarnaris A, Toma AK, Chapman MD, et al. Use of cerebrospinal fluid amyloid-beta and total tau protein to predict favorable surgical outcomes in patients with idiopathic normal pressure hydrocephalus. *Journal of Neurosurgery* 2011;115(1):145-50.
- 14. Toma AK, Stapleton S, Papadopoulos MC, et al. Natural history of idiopathic normal-pressure hydrocephalus. *Neurosurgical Review* 2011;34(4):433-9.
- 15. Andren K, Wikkelso C, Tisell M, et al. Natural course of idiopathic normal pressure hydrocephalus. *Journal of Neurology, Neurosurgery and Psychiatry* 2014;85(7):806-10.
- 16. Relkin N, Marmarou A, Klinge P, et al. Diagnosing idiopathic normalpressure hydrocephalus. *Neurosurgery* 2005;57(3 Suppl):4-16.
- 17. Ishikawa M, Hashimoto M, Kuwana N, et al. Guidelines for management of idiopathic normal pressure hydrocephalus: Guidelines from the Guidelines committee of idiopathic normal pressure hydrocephalus, the Japanese society of normal pressure hydrocephalus. *Neurologia Medico-Chirurgica* 2008;48:1-23.
- 18. Hashimoto M, Ishikawa M, Mori E, et al. Diagnosis of idiopathic normal pressure hydrocephalus is supported by MRI-based scheme: a prospective cohort study. *Cerebrospinal Fluid Research* 2010;7:18.
- 19. Stolze H, Kuhtz-Buschbeck JP, Drucke H, et al. Gait analysis in idiopathic normal pressure hydrocephalus--which parameters respond to the CSF tap test? *Clinical Neurophysiology* 2000;111(9):1678-86.
- 20. Knutsson E, Lying-Tunell U. Gait apraxia in normal-pressure hydrocephalus: patterns of movement and muscle activation. *Neurology* 1985;35(2):155-60.

- 21. Blomsterwall E, Svantesson U, Carlsson U, et al. Postural disturbance in patients with normal pressure hydrocephalus. *Acta Neurologica Scandinavica* 2000;102(5):284-91.
- 22. Blomsterwall E, Frisen L, Wikkelso C. Postural function and subjective eye level in patients with idiopathic normal pressure hydrocephalus. *Journal of Neurology* 2011;258(7):1341-6.
- 23. Wikkelso C, Blomsterwall E, Frisen L. Subjective visual vertical and Romberg's test correlations in hydrocephalus. *Journal of Neurology* 2003;250(6):741-5.
- 24. Devito EE, Pickard JD, Salmond CH, et al. The neuropsychology of normal pressure hydrocephalus (NPH). *British Journal of Neurosurgery* 2005;19(3):217-24.
- 25. Iddon JL, Pickard JD, Cross JJ, et al. Specific patterns of cognitive impairment in patients with idiopathic normal pressure hydrocephalus and Alzheimer's disease: a pilot study. *Journal of Neurology, Neurosurgery and Psychiatry* 1999;67(6):723-32.
- 26. Hellstrom P, Klinge P, Tans J, et al. The neuropsychology of iNPH: findings and evaluation of tests in the European multicentre study. *Clinical Neurology and Neurosurgery* 2012;114(2):130-4.
- Hellstrom P, Klinge P, Tans J, et al. A new scale for assessment of severity and outcome in iNPH. *Acta Neurologica Scandinavica* 2012;126(4):229-37.
- Lindqvist G, Andersson H, Bilting M, et al. Normal pressure hydrocephalus: psychiatric findings before and after shunt operation classified in a new diagnostic system for organic psychiatry. *Acta Psychiatrica Scandinavica Supplementum* 1993;373:18-32.
- 29. Hellström P. The neuropsychology of idiopathic normal pressure hydrocephalus. Institute of Neuroscience and Physiology, Department of Clinical Neuroscience and Rehabilitation, University of Gothenburg, 2011.
- 30. Hellstrom P, Edsbagge M, Blomsterwall E, et al. Neuropsychological effects of shunt treatment in idiopathic normal pressure hydrocephalus. *Neurosurgery* 2008;63(3):527-35.

- 31. Tullberg M, Hellstrom P, Piechnik SK, et al. Impaired wakefulness is associated with reduced anterior cingulate CBF in patients with normal pressure hydrocephalus. *Acta Neurologica Scandinavica* 2004;110(5):322-30.
- 32. Hellstrom P, Edsbagge M, Archer T, et al. The neuropsychology of patients with clinically diagnosed idiopathic normal pressure hydrocephalus. *Neurosurgery* 2007;61(6):1219-26.
- 33. Jonas S, Brown J. Neurogenic bladder in normal pressure hydrocephalus. *Urology* 1975;5(1):44-50.
- 34. Sakakibara R, Kanda T, Sekido T, et al. Mechanism of bladder dysfunction in idiopathic normal pressure hydrocephalus. *Neurourology and Urodynamics* 2008;27(6):507-10.
- 35. Blomsterwall E, Bilting M, Stephensen H, et al. Gait abnormality is not the only motor disturbance in normal pressure hydrocephalus. *Scandinavian Journal of Rehabilitation Medicine* 1995;27(4):205-9.
- 36. Agerskov S, Hellstrom P, Andren K, et al. The phenotype of idiopathic normal pressure hydrocephalus-a single center study of 429 patients. *Journal of the Neurological Sciences* 2018;391(1878-5883):54-60.
- 37. Tullberg M, Jensen C, Ekholm S, et al. Normal pressure hydrocephalus: Vascular white matter changes on MR images must not exclude patients from shunt surgery. *American Journal of Neuroradiology* 2001;22(9):1665-73.
- 38. Craven CL, Baudracco I, Zetterberg H, et al. The predictive value of T-tau and AB1-42 levels in idiopathic normal pressure hydrocephalus. *Acta Neurochirurgica* 2017;159(12):2293-300.
- 39. Wikkelso C, Andersson H, Blomstrand C, et al. Normal pressure hydrocephalus. Predictive value of the cerebrospinal fluid tap-test. *Acta Neurologica Scandinavica* 1986;73(6):566-73.
- 40. Kilic K, Czorny A, Auque J, et al. Predicting the outcome of shunt surgery in normal pressure hydrocephalus. *Journal of Clinical Neuroscience* 2007;14(8):729-36.

- 41. Ishikawa M, Hashimoto M, Mori E, et al. The value of the cerebrospinal fluid tap test for predicting shunt effectiveness in idiopathic normal pressure hydrocephalus. *Fluids Barriers CNS* 2012;9(1):1.
- 42. Vanneste JA. Diagnosis and management of normal-pressure hydrocephalus. *Journal of Neurology* 2000;247(1):5-14.
- 43. Marmarou A, Bergsneider M, Klinge P, et al. The value of supplemental prognostic tests for the preoperative assessment of idiopathic normal-pressure hydrocephalus. *Neurosurgery* 2005;57(3 Suppl):17-28;.
- 44. McComb JG. Recent research into the nature of cerebrospinal fluid formation and absorption. *Journal of Neurosurgery* 1983;59(3):369-83.
- 45. Davson H, Welch K, Segal M. Physiology and pathophysiology of the cerebrospinal fluid. Edinburgh: Churchill Livingstone 1987.
- 46. Silverberg GD, Mayo M, Saul T, et al. Novel ventriculo-peritoneal shunt in Alzheimer's disease cerebrospinal fluid biomarkers. *Expert Review of Neurotherapeutics* 2004;4(1):97-107.
- 47. Borgesen SE, Gjerris F. Relationships between Intracranial-Pressure, Ventricular Size, and Resistance to Csf Outflow. *Journal of Neurosurgery* 1987;67(4):535-39.
- 48. Arango C, McMahon RP, Lefkowitz DM, et al. Patterns of cranial, brain and sulcal CSF volumes in male and female deficit and nondeficit patients with schizophrenia. *Psychiatry Research* 2008;162(2):91-100.
- 49. Edsbagge M, Starck G, Zetterberg H, et al. Spinal cerebrospinal fluid volume in healthy elderly individuals. *Clinical Anatomy* 2011;24(6):733-40.
- 50. Battal B, Kocaoglu M, Bulakbasi N, et al. Cerebrospinal fluid flow imaging by using phase-contrast MR technique. *British Journal of Radiology* 2011;84(1004):758-65.
- 51. Edsbagge M, Tisell M, Jacobsson L, et al. Spinal CSF absorption in healthy individuals. *American Journal of Physiology: Regulatory, Integrative and Comparative Physiology* 2004;287(6):1450-5.

- 52. Abbott NJ. Evidence for bulk flow of brain interstitial fluid: significance for physiology and pathology. *Neurochemistry International* 2004;45(4):545-52.
- 53. Rekate HL. The definition and classification of hydrocephalus: a personal recommendation to stimulate debate. *Cerebrospinal Fluid Research* 2008;5:2.
- 54. Deo-Narine V, Gomez DG, Vullo T, et al. Direct In Vivo Observation of Transventricular Absorption in the Hydrocephalic Dog Using Magnetic Resonance Imaging. *Investigative Radiology* 1994;29(3):287-93.
- 55. Brinker T, Stopa E, Morrison J, et al. A new look at cerebrospinal fluid circulation. *Fluids Barriers CNS* 2014;11:10.
- 56. Iliff JJ, Wang MH, Liao YH, et al. A Paravascular Pathway Facilitates CSF Flow Through the Brain Parenchyma and the Clearance of Interstitial Solutes, Including Amyloid beta. *Science Translational Medicine* 2012;4(147)
- 57. Louveau A, Plog BA, Antila S, et al. Understanding the functions and relationships of the glymphatic system and meningeal lymphatics. *Journal of Clinical Investigation* 2017;127(9):3210-19.
- 58. Absinta M, Ha SK, Nair G, et al. Human and nonhuman primate meninges harbor lymphatic vessels that can be visualized noninvasively by MRI. *eLife* 2017;6
- 59. Mathiisen TM, Lehre KP, Danbolt NC, et al. The perivascular astroglial sheath provides a complete covering of the brain microvessels: an electron microscopic 3D reconstruction. *Glia* 2010;58(9):1094-103.
- 60. Papadopoulos MC, Verkman AS. Aquaporin water channels in the nervous system. *Nature Reviews: Neuroscience* 2013;14(4):265-77.
- 61. Thrane AS, Rangroo Thrane V, Nedergaard M. Drowning stars: reassessing the role of astrocytes in brain edema. *Trends in Neurosciences* 2014;37(11):620-8.
- 62. Kress BT, Iliff JJ, Xia M, et al. Impairment of paravascular clearance pathways in the aging brain. *Annals of Neurology* 2014;76(6):845-61.
- 63. Xie L, Kang H, Xu Q, et al. Sleep drives metabolite clearance from the adult brain. *Science* 2013;342(6156):373-7.

- 64. Ding F, O'Donnell J, Xu Q, et al. Changes in the composition of brain interstitial ions control the sleep-wake cycle. *Science* 2016;352(6285):550-5.
- 65. Biomarkers Definitions Working G. Biomarkers and surrogate endpoints: preferred definitions and conceptual framework. *Clinical Pharmacology and Therapeutics* 2001;69(3):89-95.
- 66. Tarnaris A, Toma AK, Chapman MD, et al. Rostrocaudal dynamics of CSF biomarkers. *Neurochemical Research* 2011;36(3):528-32.
- 67. Blennow K, Hampel H. CSF markers for incipient Alzheimer's disease. Lancet Neurology 2003;2(10):605-13.
- 68. Dubois B, Feldman HH, Jacova C, et al. Advancing research diagnostic criteria for Alzheimer's disease: the IWG-2 criteria. *Lancet Neurology* 2014;13(6):614-29.
- 69. Agren-Wilsson A, Lekman A, Sjoberg W, et al. CSF biomarkers in the evaluation of idiopathic normal pressure hydrocephalus. *Acta Neurologica Scandinavica* 2007;116(5):333-9.
- 70. Tarnaris A, Watkins LD, Kitchen ND. Biomarkers in chronic adult hydrocephalus. *Cerebrospinal Fluid Research* 2006;3:11.
- 71. Tarnaris A, Toma AK, Kitchen ND, et al. Ongoing search for diagnostic biomarkers in idiopathic normal pressure hydrocephalus. *Biomarkers in Medicine* 2009;3(6):787-805.
- 72. Tarnaris A, Toma AK, Chapman MD, et al. The longitudinal profile of CSF markers during external lumbar drainage. *Journal of Neurology, Neurosurgery and Psychiatry* 2009;80(10):1130-3.
- 73. Kudo T, Mima T, Hashimoto R, et al. Tau protein is a potential biological marker for normal pressure hydrocephalus. *Psychiatry and Clinical Neurosciences* 2000;54(2):199-202.
- 74. Leinonen V, Menon LG, Carroll RS, et al. Cerebrospinal fluid biomarkers in idiopathic normal pressure hydrocephalus. *International Journal of Alzheimer's Disease* 2011;2011:312526.
- 75. Tullberg M, Blennow K, Mansson JE, et al. Ventricular cerebrospinal fluid neurofilament protein levels decrease in parallel with white matter pathology after shunt surgery in normal pressure hydrocephalus. *European Journal of Neurology* 2007;14(3):248-54.

- 76. Patel S, Lee EB, Xie SX, et al. Phosphorylated tau/amyloid beta 1-42 ratio in ventricular cerebrospinal fluid reflects outcome in idiopathic normal pressure hydrocephalus. *Fluids Barriers CNS* 2012;9(1):7.
- 77. Nakajima M, Miyajima M, Ogino I, et al. Leucine-rich alpha-2glycoprotein is a marker for idiopathic normal pressure hydrocephalus. *Acta Neurochirurgica* 2011;153(6):1339-46.
- 78. Kapaki EN, Paraskevas GP, Tzerakis NG, et al. Cerebrospinal fluid tau, phospho-tau181 and beta-amyloid1-42 in idiopathic normal pressure hydrocephalus: a discrimination from Alzheimer's disease. *European Journal of Neurology* 2007;14(2):168-73.
- 79. Muller UC, Deller T, Korte M. Not just amyloid: physiological functions of the amyloid precursor protein family. *Nature Reviews: Neuroscience* 2017;18(5):281-98.
- Kang J, Lemaire HG, Unterbeck A, et al. The precursor of Alzheimer's disease amyloid A4 protein resembles a cell-surface receptor. *Nature* 1987;325(6106):733-6.
- 81. Vassar R, Bennett BD, Babu-Khan S, et al. Beta-secretase cleavage of Alzheimer's amyloid precursor protein by the transmembrane aspartic protease BACE. *Science* 1999;286(5440):735-41.
- 82. Englund H. Soluble amyloid-β aggregates in Alzheimer´s disease. Acta Universitatis Upsaliensis :, 2009.
- Haass C, Selkoe DJ. Soluble protein oligomers in neurodegeneration: lessons from the Alzheimer's amyloid beta-peptide. *Nature Reviews: Molecular Cell Biology* 2007;8(2):101-12.
- 84. Gabelle A, Roche S, Geny C, et al. Correlations between soluble alpha/beta forms of amyloid precursor protein and Abeta38, 40, and 42 in human cerebrospinal fluid. *Brain Research* 2010;1357:175-83.
- 85. Okochi M, Tagami S, Yanagida K, et al. gamma-secretase modulators and presenilin 1 mutants act differently on presenilin/gammasecretase function to cleave Abeta42 and Abeta43. *Cell Rep* 2013;3(1):42-51.
- 86. Steiner H, Fluhrer R, Haass C. Intramembrane proteolysis by gammasecretase. *Journal of Biological Chemistry* 2008;283(44):29627-31.

- 87. Blennow K, de Leon MJ, Zetterberg H. Alzheimer's disease. *Lancet* 2006;368(9533):387-403.
- 88. Masters CL, Simms G, Weinman NA, et al. Amyloid plaque core protein in Alzheimer disease and Down syndrome. *Proceedings of the National Academy of Sciences of the United States of America* 1985;82(12):4245-9.
- 89. Rosen C, Andreasson U, Mattsson N, et al. Cerebrospinal fluid profiles of amyloid beta-related biomarkers in Alzheimer's disease. *Neuromolecular Medicine* 2012;14(1):65-73.
- 90. Hong S, Quintero-Monzon O, Ostaszewski BL, et al. Dynamic analysis of amyloid beta-protein in behaving mice reveals opposing changes in ISF versus parenchymal Abeta during age-related plaque formation. *Journal of Neuroscience* 2011;31(44):15861-9.
- 91. Rodrigue KM, Kennedy KM, Devous MD, Sr., et al. beta-Amyloid burden in healthy aging: regional distribution and cognitive consequences. *Neurology* 2012;78(6):387-95.
- 92. Selkoe DJ, Hardy J. The amyloid hypothesis of Alzheimer's disease at 25 years. *EMBO Molecular Medicine* 2016;8(6):595-608.
- 93. van der Kant R, Goldstein LS. Cellular functions of the amyloid precursor protein from development to dementia. *Developmental Cell* 2015;32(4):502-15.
- 94. Taylor CJ, Ireland DR, Ballagh I, et al. Endogenous secreted amyloid precursor protein-alpha regulates hippocampal NMDA receptor function, long-term potentiation and spatial memory. *Neurobiology* of Disease 2008;31(2):250-60.
- 95. Cousins SL, Dai W, Stephenson FA. APLP1 and APLP2, members of the APP family of proteins, behave similarly to APP in that they associate with NMDA receptors and enhance NMDA receptor surface expression. *Journal of Neurochemistry* 2015;133(6):879-85.
- Klinge PM, Samii A, Niescken S, et al. Brain amyloid accumulates in aged rats with kaolin-induced hydrocephalus. *Neuroreport* 2006;17(6):657-60.

- 97. Silverberg GD, Miller MC, Pascale CL, et al. Kaolin-induced chronic hydrocephalus accelerates amyloid deposition and vascular disease in transgenic rats expressing high levels of human APP. *Fluids Barriers CNS* 2015;12(1):2.
- 98. Silverberg GD, Mayo M, Saul T, et al. Alzheimer's disease, normalpressure hydrocephalus, and senescent changes in CSF circulatory physiology: a hypothesis. *Lancet Neurology* 2003;2(8):506-11.
- 99. Eggert S, Paliga K, Soba P, et al. The proteolytic processing of the amyloid precursor protein gene family members APLP-1 and APLP-2 involves alpha-, beta-, gamma-, and epsilon-like cleavages: modulation of APLP-1 processing by n-glycosylation. *Journal of Biological Chemistry* 2004;279(18):18146-56.
- 100. Yanagida K, Okochi M, Tagami S, et al. The 28-amino acid form of an APLP1-derived Abeta-like peptide is a surrogate marker for Abeta42 production in the central nervous system. *EMBO Molecular Medicine* 2009;1(4):223-35.
- 101. Shariati SA, De Strooper B. Redundancy and divergence in the amyloid precursor protein family. *FEBS Letters* 2013;587(13):2036-45.
- 102. Jacobsen KT, Iverfeldt K. Amyloid precursor protein and its homologues: a family of proteolysis-dependent receptors. *Cellular and Molecular Life Sciences* 2009;66(14):2299-318.
- 103. Tagami S, Okochi M, Yanagida K, et al. Relative ratio and level of amyloid-beta 42 surrogate in cerebrospinal fluid of familial Alzheimer disease patients with presenilin 1 mutations. *Neurodegenerative Diseases* 2014;13(2-3):166-70.
- 104. Reiber H. Dynamics of brain-derived proteins in cerebrospinal fluid. *Clinica Chimica Acta* 2001;310(2):173-86.
- 105. Lee VM, Goedert M, Trojanowski JQ. Neurodegenerative tauopathies. *Annual Review of Neuroscience* 2001;24:1121-59.
- 106. Mattsson N. Cerebrospinal fluid biomarkers reflecting β-amyloid and axonal pathology in Alzheimer's disease and related conditions. Institute of Neuroscience and Physiology, Dept. of Psychiatry and Neurochemistry, The Sahlgrenska Academy, University of Gothenburg, 2011.

- 107. Deisenhammer F, Egg R, Giovannoni G, et al. EFNS guidelines on disease-specific CSF investigations. *European Journal of Neurology* 2009;16(6):760-70.
- 108. Zetterberg H. Neurofilament Light: A Dynamic Cross-Disease Fluid Biomarker for Neurodegeneration. *Neuron* 2016;91(1):1-3.
- 109. Norgren N, Rosengren L, Stigbrand T. Elevated neurofilament levels in neurological diseases. *Brain Research* 2003;987(1):25-31.
- 110. Zetterberg H, Skillback T, Mattsson N, et al. Association of Cerebrospinal Fluid Neurofilament Light Concentration With Alzheimer Disease Progression. *JAMA Neurol* 2016;73(1):60-7.
- 111. Pfeiffer SE, Warrington AE, Bansal R. The oligodendrocyte and its many cellular processes. *Trends in Cell Biology* 1993;3(6):191-7.
- 112. Sutton LN, Wood JH, Brooks BR, et al. Cerebrospinal fluid myelin basic protein in hydrocephalus. *Journal of Neurosurgery* 1983;59(3):467-70.
- 113. Wyss-Coray T, Mucke L. Inflammation in neurodegenerative disease--a double-edged sword. *Neuron* 2002;35(3):419-32.
- 114. Perry VH, Nicoll JA, Holmes C. Microglia in neurodegenerative disease. *Nature Reviews: Neurology* 2010;6(4):193-201.
- 115. Semple BD, Bye N, Rancan M, et al. Role of CCL2 (MCP-1) in traumatic brain injury (TBI): evidence from severe TBI patients and CCL2-/- mice. *Journal of Cerebral Blood Flow and Metabolism* 2010;30(4):769-82.
- 116. Gerszten RE, Garcia-Zepeda EA, Lim YC, et al. MCP-1 and IL-8 trigger firm adhesion of monocytes to vascular endothelium under flow conditions. *Nature* 1999;398(6729):718-23.
- 117. Packard RR, Lichtman AH, Libby P. Innate and adaptive immunity in atherosclerosis. *Seminars in Immunopathology* 2009;31(1):5-22.
- 118. Mölne J, Wold A. Inflammation. Stockholm: Liber 2007.
- 119. Wyss-Coray T, Loike JD, Brionne TC, et al. Adult mouse astrocytes degrade amyloid-beta in vitro and in situ. *Nature Medicine* 2003;9(4):453-7.

- 120. Mattsson N, Tabatabaei S, Johansson P, et al. Cerebrospinal fluid microglial markers in Alzheimer's disease: elevated chitotriosidase activity but lack of diagnostic utility. *Neuromolecular Medicine* 2011;13(2):151-9.
- 121. Glass CK, Saijo K, Winner B, et al. Mechanisms underlying inflammation in neurodegeneration. *Cell* 2010;140(6):918-34.
- 122. Rosengren LE, Ahlsen G, Belfrage M, et al. A sensitive ELISA for glial fibrillary acidic protein: application in CSF of children. *Journal of Neuroscience Methods* 1992;44(2-3):113-9.
- 123. Rosengren LE, Wikkelso C, Hagberg L. A sensitive ELISA for glial fibrillary acidic protein: application in CSF of adults. *Journal of Neuroscience Methods* 1994;51(2):197-204.
- 124. Bonneh-Barkay D, Wang G, Starkey A, et al. In vivo CHI3L1 (YKL-40) expression in astrocytes in acute and chronic neurological diseases. *Journal of Neuroinflammation* 2010;7:34.
- 125. Toma AK, Tarnaris A, Kitchen ND, et al. Working towards patient oriented outcome assessment in normal pressure hydrocephalus, what is the most important? *Acta Neurochirurgica* 2011;153(1):177-80.
- 126. Walker JM, Rapley R. Medical biomethods handbook. Totowa, N.J.: Humana Press 2005.
- 127. Engvall E, Perlmann P. Enzyme-linked immunosorbent assay (ELISA). Quantitative assay of immunoglobulin G. *Immunochemistry* 1971;8(9):871-4.
- 128. Voller A, Bartlett A, Bidwell DE. Enzyme immunoassays with special reference to ELISA techniques. *Journal of Clinical Pathology* 1978;31(6):507-20.
- 129. Olsson A, Vanderstichele H, Andreasen N, et al. Simultaneous measurement of beta-amyloid(1-42), total tau, and phosphorylated tau (Thr181) in cerebrospinal fluid by the xMAP technology. *Clinical Chemistry* 2005;51(2):336-45.
- 130. Hansson O, Zetterberg H, Buchhave P, et al. Association between CSF biomarkers and incipient Alzheimer's disease in patients with mild cognitive impairment: a follow-up study. *Lancet Neurology* 2006;5(3):228-34.

- 131. Gaetani L, Hoglund K, Parnetti L, et al. A new enzyme-linked immunosorbent assay for neurofilament light in cerebrospinal fluid: analytical validation and clinical evaluation. *Alzheimer's Research and Therapy* 2018;10(1):8.
- 132. Rosengren LE, Karlsson JE, Karlsson JO, et al. Patients with amyotrophic lateral sclerosis and other neurodegenerative diseases have increased levels of neurofilament protein in CSF. *Journal of Neurochemistry* 1996;67(5):2013-18.
- 133. Zetterberg H, Andreasson U, Hansson O, et al. Elevated cerebrospinal fluid BACE1 activity in incipient Alzheimer disease. *Archives of Neurology* 2008;65(8):1102-7.
- 134. Wahlund LO, Barkhof F, Fazekas F, et al. A new rating scale for agerelated white matter changes applicable to MRI and CT. *Stroke* 2001;32(6):1318-22.
- 135. Hughes AJ, Daniel SE, Kilford L, et al. Accuracy of clinical diagnosis of idiopathic Parkinson's disease: a clinico-pathological study of 100 cases. *Journal of Neurology, Neurosurgery and Psychiatry* 1992;55(3):181-4.
- 136. Gilman S, Wenning GK, Low PA, et al. Second consensus statement on the diagnosis of multiple system atrophy. *Neurology* 2008;71(9):670-6.
- 137. Litvan I, Bhatia KP, Burn DJ, et al. Movement Disorders Society Scientific Issues Committee report: SIC Task Force appraisal of clinical diagnostic criteria for Parkinsonian disorders. *Movement Disorders* 2003;18(5):467-86.
- 138. Armstrong MJ, Litvan I, Lang AE, et al. Criteria for the diagnosis of corticobasal degeneration. *Neurology* 2013;80(5):496-503.
- 139. McKhann G, Drachman D, Folstein M, et al. Clinical diagnosis of Alzheimer's disease: report of the NINCDS-ADRDA Work Group under the auspices of Department of Health and Human Services Task Force on Alzheimer's Disease. *Neurology* 1984;34(7):939-44.
- 140. Neary D, Snowden JS, Gustafson L, et al. Frontotemporal lobar degeneration: a consensus on clinical diagnostic criteria. *Neurology* 1998;51(6):1546-54.

- 141. Organization WH. The ICD-10 Classification of Mental and Behavioural Disorders. *World Health Organization* 1993
- 142. Roman GC, Tatemichi TK, Erkinjuntti T, et al. Vascular dementia: diagnostic criteria for research studies. Report of the NINDS-AIREN International Workshop. *Neurology* 1993;43(2):250-60.
- 143. Wallin A, Nordlund A, Jonsson M, et al. The Gothenburg MCI study: Design and distribution of Alzheimer's disease and subcortical vascular disease diagnoses from baseline to 6-year follow-up. *Journal of Cerebral Blood Flow and Metabolism* 2016;36(1):114-31.
- 144. Lins H, Wichart I, Bancher C, et al. Immunoreactivities of amyloid beta peptide((1-42)) and total tau protein in lumbar cerebrospinal fluid of patients with normal pressure hydrocephalus. J Neural Transm (Vienna) 2004;111(3):273-80.
- 145. Gloeckner SF, Meyne F, Wagner F, et al. Quantitative analysis of transthyretin, tau and amyloid-beta in patients with dementia. *Journal of Alzheimer's Disease* 2008;14(1):17-25.
- 146. Ray B, Reyes PF, Lahiri DK. Biochemical studies in Normal Pressure Hydrocephalus (NPH) patients: change in CSF levels of amyloid precursor protein (APP), amyloid-beta (Abeta) peptide and phospho-tau. *Journal of Psychiatric Research* 2011;45(4):539-47.
- 147. Poulsen K, Bahl JM, Simonsen AH, et al. Distinct transthyretin oxidation isoform profile in spinal fluid from patients with Alzheimer's disease and mild cognitive impairment. *Clinical Proteomics* 2014;11(1):12.
- 148. Schirinzi T, Sancesario GM, Ialongo C, et al. A clinical and biochemical analysis in the differential diagnosis of idiopathic normal pressure hydrocephalus. *Frontiers in Neurology* 2015;6:86.
- 149. Jeppsson A, Holtta M, Zetterberg H, et al. Amyloid mis-metabolism in idiopathic normal pressure hydrocephalus. *Fluids Barriers CNS* 2016;13(1):13.
- 150. Santangelo R, Cecchetti G, Bernasconi MP, et al. Cerebrospinal Fluid Amyloid-beta 42, Total Tau and Phosphorylated Tau are Low in Patients with Normal Pressure Hydrocephalus: Analogies and Differences with Alzheimer's Disease. *Journal of Alzheimer's Disease* 2017;60(1):183-200.

- 151. Akiba C, Nakajima M, Miyajima M, et al. Change of Amyloid-beta 1-42 Toxic Conformer Ratio After Cerebrospinal Fluid Diversion Predicts Long-Term Cognitive Outcome in Patients with Idiopathic Normal Pressure Hydrocephalus. *Journal of Alzheimer's Disease* 2018;63(3):989-1002.
- 152. Jeppsson A, Zetterberg H, Blennow K, et al. Idiopathic normalpressure hydrocephalus: pathophysiology and diagnosis by CSF biomarkers. *Neurology* 2013;80(15):1385-92.
- 153. Miyajima M, Nakajima M, Ogino I, et al. Soluble amyloid precursor protein alpha in the cerebrospinal fluid as a diagnostic and prognostic biomarker for idiopathic normal pressure hydrocephalus. *European Journal of Neurology* 2013;20(2):236-42.
- 154. Miyajima M, Nakajima M, Motoi Y, et al. Leucine-rich alpha2glycoprotein is a novel biomarker of neurodegenerative disease in human cerebrospinal fluid and causes neurodegeneration in mouse cerebral cortex. *PLoS One* 2013;8(9):e74453.
- 155. Moriya M, Miyajima M, Nakajima M, et al. Impact of cerebrospinal fluid shunting for idiopathic normal pressure hydrocephalus on the amyloid cascade. *PloS One* 2015;10(3):e0119973.
- 156. Jurjevic I, Miyajima M, Ogino I, et al. Decreased Expression of hsamiR-4274 in Cerebrospinal Fluid of Normal Pressure Hydrocephalus Mimics with Parkinsonian Syndromes. *Journal of Alzheimer's Disease* 2017;56(1):317-25.
- 157. Tsai A, Malek-Ahmadi M, Kahlon V, et al. Differences in Cerebrospinal Fluid Biomarkers between Clinically Diagnosed Idiopathic Normal Pressure Hydrocephalus and Alzheimer's Disease. J Alzheimers Dis Parkinsonism 2014;4(4).(pii):1000150.
- 158. Jingami N, Asada-Utsugi M, Uemura K, et al. Idiopathic Normal Pressure Hydrocephalus has a Different Cerebrospinal Fluid Biomarker Profile from Alzheimer's Disease. *Journal of Alzheimers Disease* 2015;45(1):109-15.
- 159. Pyykko OT, Lumela M, Rummukainen J, et al. Cerebrospinal fluid biomarker and brain biopsy findings in idiopathic normal pressure hydrocephalus. *PloS One* 2014;9(3):e91974.

- 160. Silverberg GD, Mayo M, Saul T, et al. Continuous CSF drainage in AD: results of a double-blind, randomized, placebo-controlled study. *Neurology* 2008;71(3):202-9.
- 161. Silverberg GD, Levinthal E, Sullivan EV, et al. Assessment of lowflow CSF drainage as a treatment for AD: results of a randomized pilot study. *Neurology* 2002;59(8):1139-45.
- 162. Graff-Radford NR. Alzheimer CSF biomarkers may be misleading in normal-pressure hydrocephalus. *Neurology* 2014;83(17):1573-5.
- 163. Herukka SK, Rummukainen J, Ihalainen J, et al. Amyloid-beta and Tau Dynamics in Human Brain Interstitial Fluid in Patients with Suspected Normal Pressure Hydrocephalus. *Journal of Alzheimer's Disease* 2015;46(1):261-9.
- 164. Edsbagge M, Andreasson U, Ambarki K, et al. Alzheimer's Disease-Associated Cerebrospinal Fluid (CSF) Biomarkers do not Correlate with CSF Volumes or CSF Production Rate. *Journal of Alzheimers Disease* 2017;58(3):821-28.
- 165. Laitera T, Kurki MI, Pursiheimo JP, et al. The Expression of Transthyretin and Amyloid-beta Protein Precursor is Altered in the Brain of Idiopathic Normal Pressure Hydrocephalus Patients. *Journal of Alzheimer's Disease* 2015;48(4):959-68.
- 166. Portelius E, Holtta M, Soininen H, et al. Altered cerebrospinal fluid levels of amyloid beta and amyloid precursor-like protein 1 peptides in Down's syndrome. *Neuromolecular Medicine* 2014;16(2):510-6.
- 167. Tullberg M, Mansson JE, Fredman P, et al. CSF sulfatide distinguishes between normal pressure hydrocephalus and subcortical arteriosclerotic encephalopathy. *Journal of Neurology Neurosurgery and Psychiatry* 2000;69(1):74-81.
- 168. Wallin A, Roman GC, Esiri M, et al. Update on Vascular Cognitive Impairment Associated with Subcortical Small-Vessel Disease. *Journal of Alzheimer's Disease* 2018;62(3):1417-41.
- 169. Bjerke M, Zetterberg H, Edman A, et al. Cerebrospinal fluid matrix metalloproteinases and tissue inhibitor of metalloproteinases in combination with subcortical and cortical biomarkers in vascular dementia and Alzheimer's disease. *Journal of Alzheimer's Disease* 2011;27(3):665-76.

- 170. Tullberg M, Rosengren L, Blomsterwall E, et al. CSF neurofilament and glial fibrillary acidic protein in normal pressure hydrocephalus. *Neurology* 1998;50(4):1122-7.
- 171. Tullberg M, Hultin L, Ekholm S, et al. White matter changes in normal pressure hydrocephalus and Binswanger disease: specificity, predictive value and correlations to axonal degeneration and demyelination. *Acta Neurologica Scandinavica* 2002;105(6):417-26.
- 172. Albrechtsen M, Sorensen PS, Gjerris F, et al. High cerebrospinal fluid concentration of glial fibrillary acidic protein (GFAP) in patients with normal pressure hydrocephalus. *Journal of the Neurological Sciences* 1985;70(3):269-74.
- 173. Li X, Miyajima M, Mineki R, et al. Analysis of potential diagnostic biomarkers in cerebrospinal fluid of idiopathic normal pressure hydrocephalus by proteomics. *Acta Neurochirurgica* 2006;148(8):859-64.
- 174. Li X, Miyajima M, Jiang C, et al. Expression of TGF-betas and TGFbeta type II receptor in cerebrospinal fluid of patients with idiopathic normal pressure hydrocephalus. *Neuroscience Letters* 2007;413(2):141-4.
- 175. Nakajima M, Miyajima M, Ogino I, et al. Brain localization of leucinerich alpha2-glycoprotein and its role. *Acta Neurochirurgica Supplement* 2012;113:97-101.
- 176. Momjian S, Owler BK, Czosnyka Z, et al. Pattern of white matter regional cerebral blood flow and autoregulation in normal pressure hydrocephalus. *Brain* 2004;127(5):965-72.
- 177. Larsson A, Arlig A, Bergh AC, et al. Quantitative SPECT cisternography in normal pressure hydrocephalus. *Acta Neurologica Scandinavica* 1994;90(3):190-6.
- 178. Andren K, Wikkelso C, Sundstrom N, et al. Long-term effects of complications and vascular comorbidity in idiopathic normal pressure hydrocephalus: a quality registry study. *Journal of Neurology* 2018;265(1):178-86.
- 179. Jaraj D, Agerskov S, Rabiei K, et al. Vascular factors in suspected normal pressure hydrocephalus: A population-based study. *Neurology* 2016;86(7):592-9.

- 180. Israelsson H, Carlberg B, Wikkelso C, et al. Vascular risk factors in INPH: A prospective case-control study (the INPH-CRasH study). *Neurology* 2017;88(6):577-85.
- 181. Tisell M, Tullberg M, Hellstrom P, et al. Shunt surgery in patients with hydrocephalus and white matter changes. *Journal of Neurosurgery* 2011;114(5):1432-8.
- 182. Roman GC, Erkinjuntti T, Wallin A, et al. Subcortical ischaemic vascular dementia. *Lancet Neurology* 2002;1(7):426-36.
- 183. Thompson CS, Hakim AM. Living Beyond Our Physiological Means Small Vessel Disease of the Brain Is an Expression of a Systemic Failure in Arteriolar Function: A Unifying Hypothesis. *Stroke* 2009;40(5):322-30.
- 184. Wong SM, Jansen JFA, Zhang CE, et al. Blood-brain barrier impairment and hypoperfusion are linked in cerebral small vessel disease. *Neurology* 2019;92(15):1669-77.
- 185. Luikku AJ, Hall A, Nerg O, et al. Multimodal analysis to predict shunt surgery outcome of 284 patients with suspected idiopathic normal pressure hydrocephalus. *Acta Neurochirurgica* 2016;158(12):2311-19.
- 186. Nakajima M, Miyajima M, Ogino I, et al. Preoperative Phosphorylated Tau Concentration in the Cerebrospinal Fluid Can Predict Cognitive Function Three Years after Shunt Surgery in Patients with Idiopathic Normal Pressure Hydrocephalus. *Journal of Alzheimers Disease* 2018;66(1):319-31.
- 187. Kazui H, Kanemoto H, Yoshiyama K, et al. Association between high biomarker probability of Alzheimer's disease and improvement of clinical outcomes after shunt surgery in patients with idiopathic normal pressure hydrocephalus. *Journal of the Neurological Sciences* 2016;369(1878-5883):236-41.